

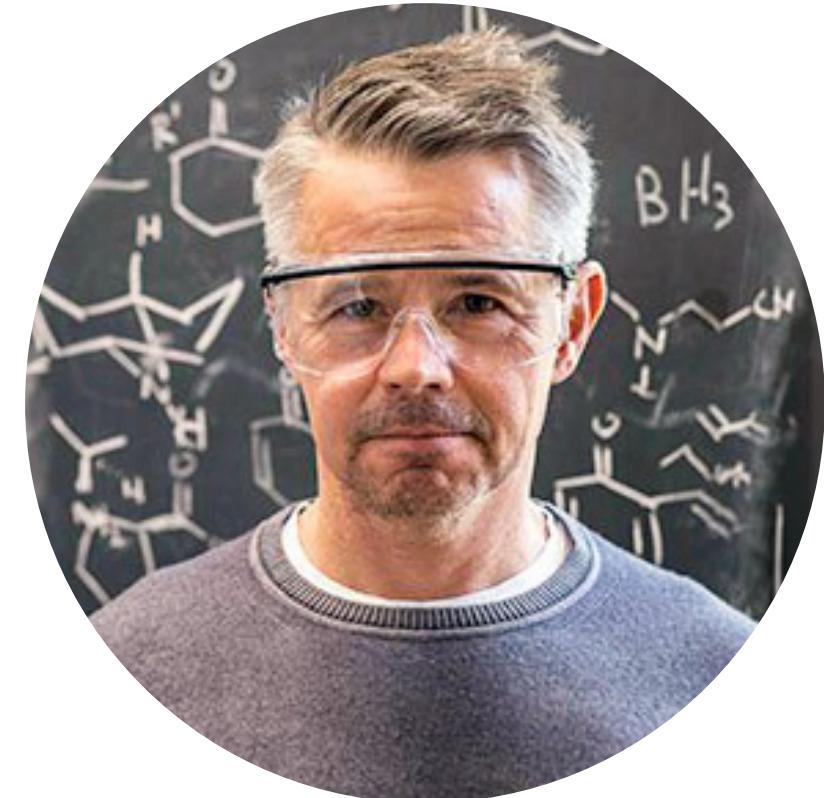
# Andrew G. Myers

Kevin Zong

Saturday Group Meeting - Seminar

September 27th, 2025

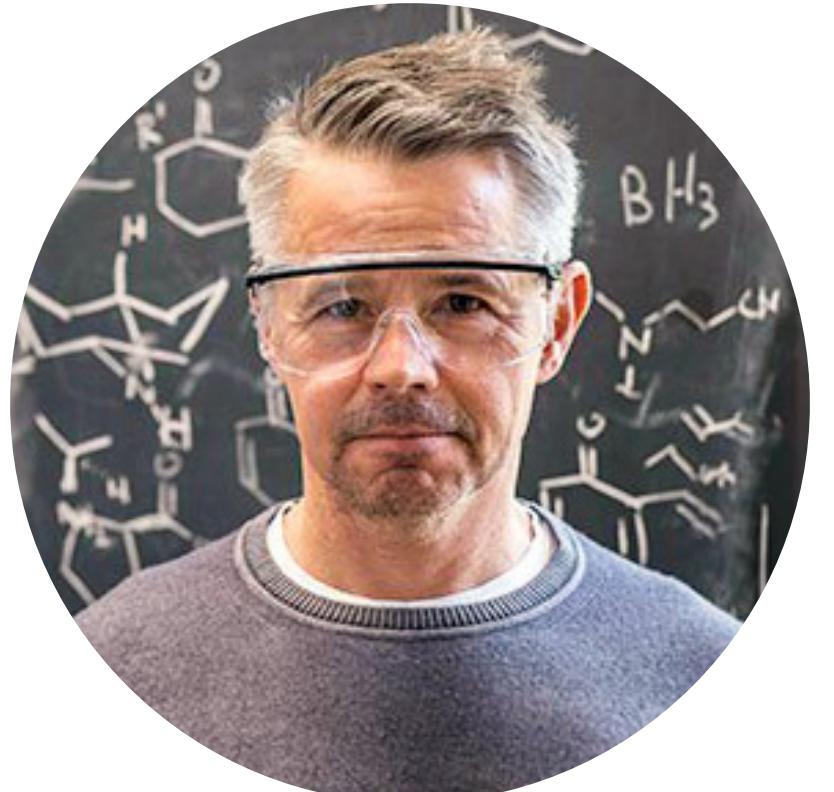
# Andrew G. Myers



# Andrew G. Myers



B.Sc.

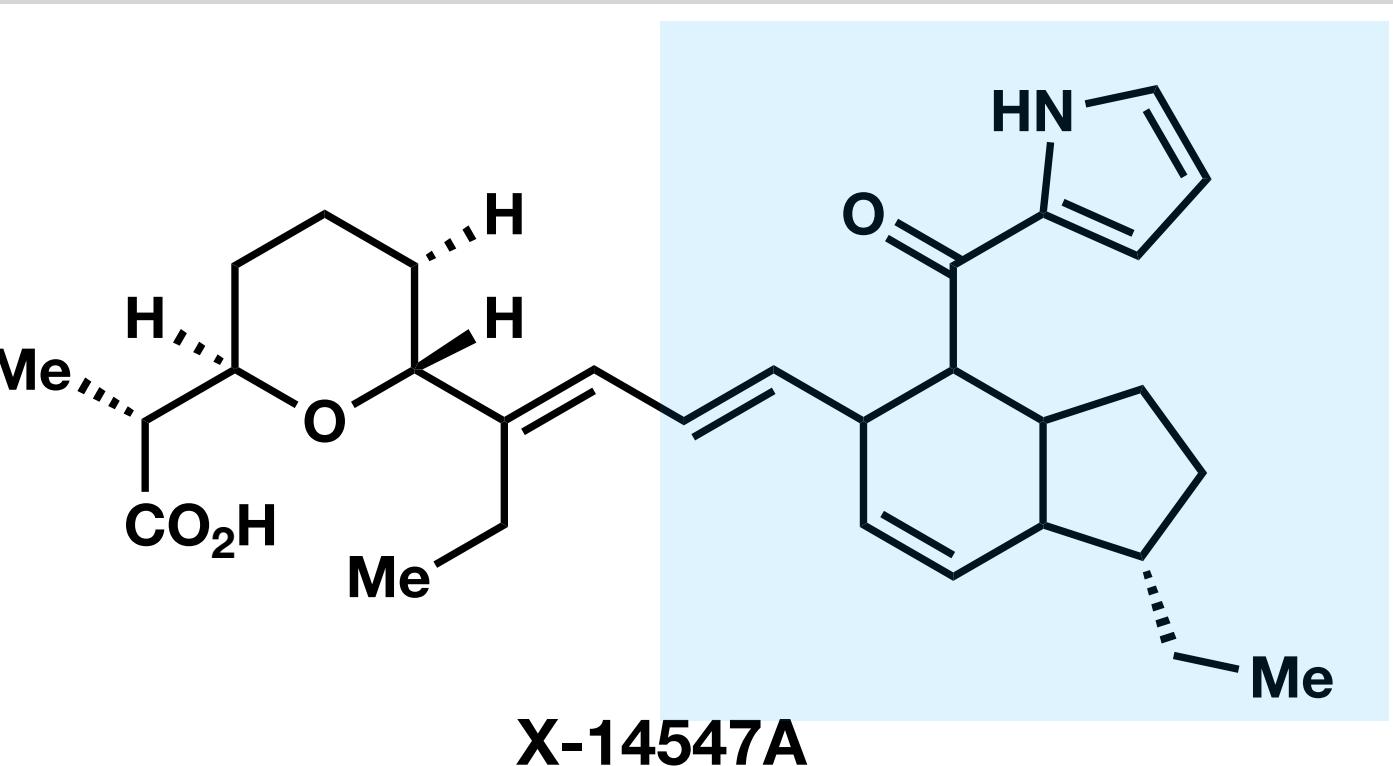


1977

1981

William R. Roush

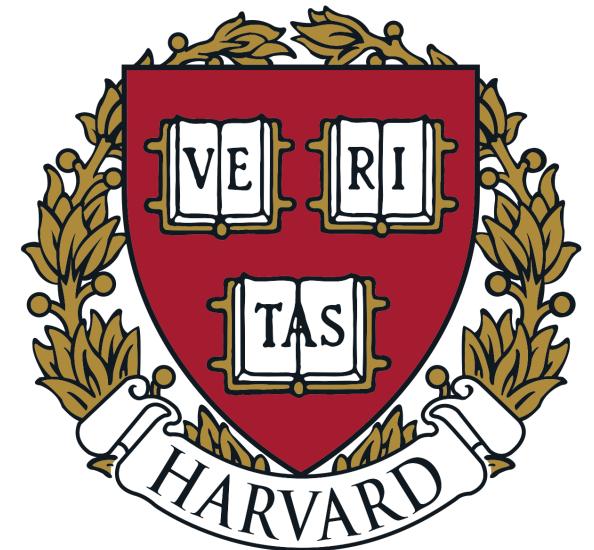
Total Synthesis of the Right-Hand Half of Antibiotic X-14547A



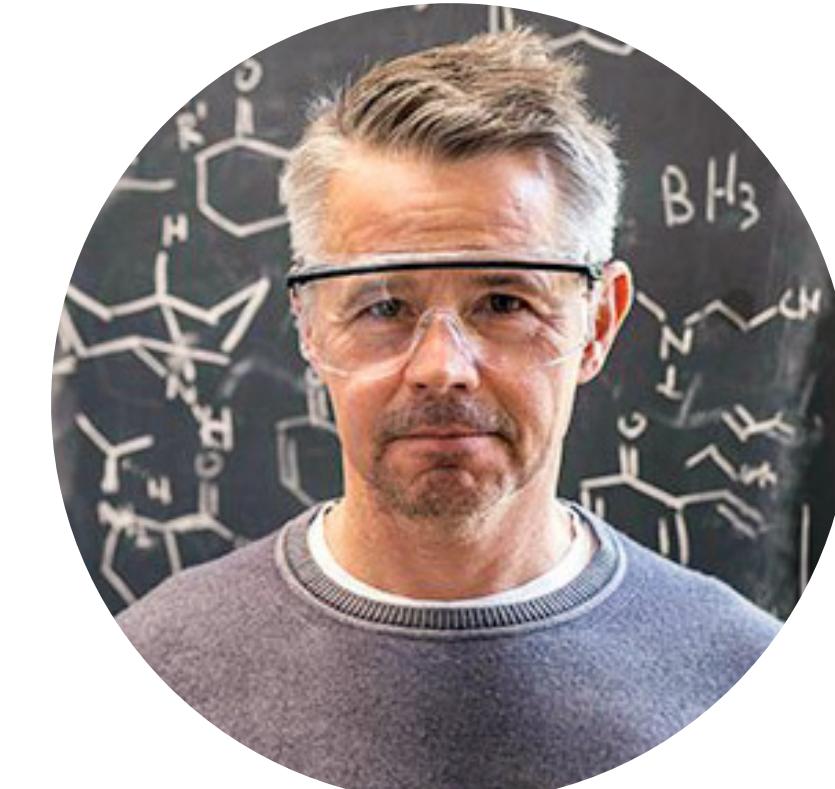
antibiotic activity against gram-positive bacteria

J. Org. Chem. 1981, 46, 1509-1511 3

# Andrew G. Myers



PhD and Post Doc

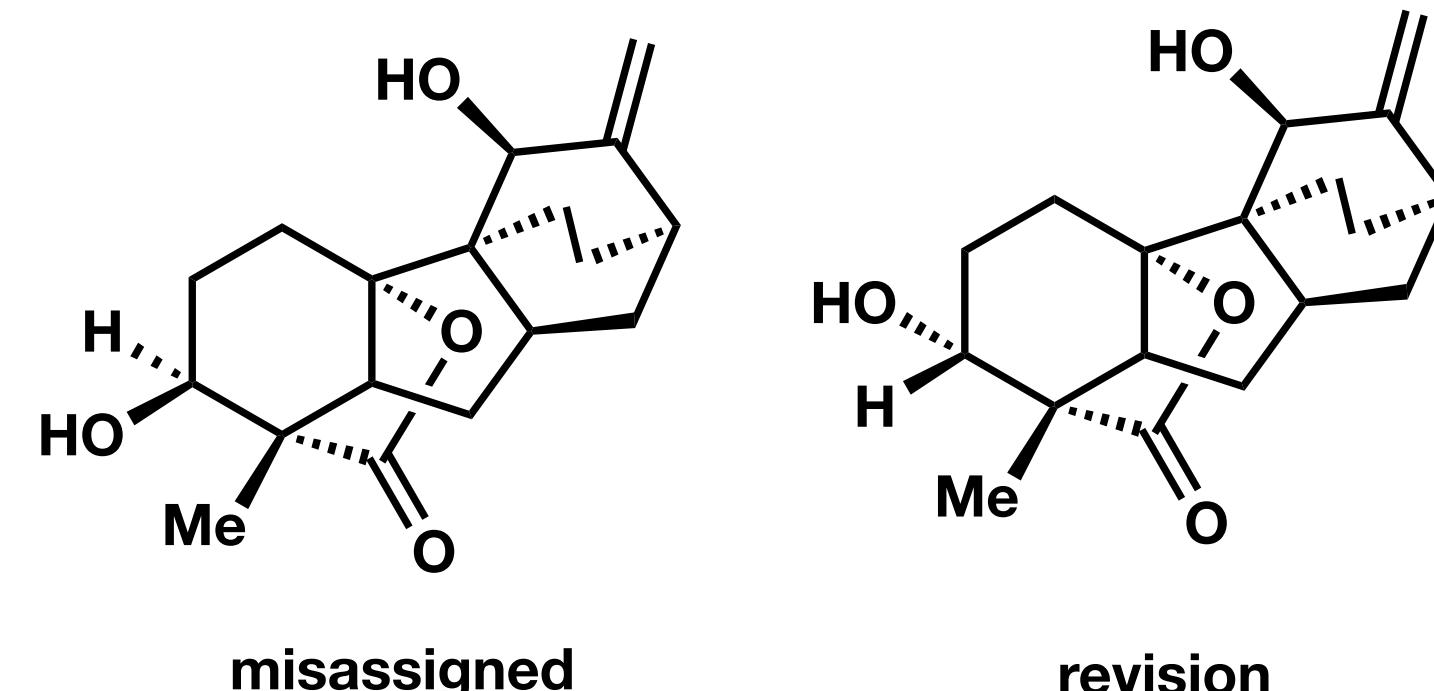


1981

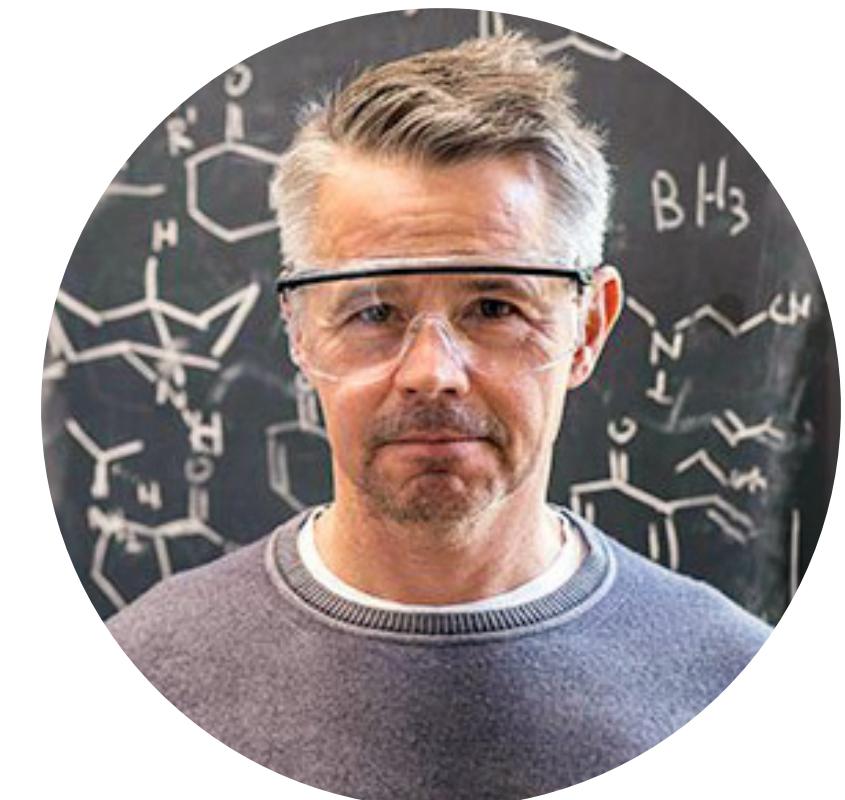
1986

E. J. Corey

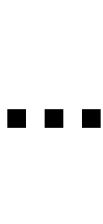
Total Synthesis and Structural Revision of ( $\pm$ ) -Antheridium-Inducing Factor ( $A_{An}, 2$ )



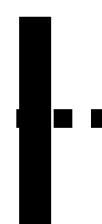
# Andrew G. Myers



Assistant, Associate, and  
then Full Professor (1994)



1986

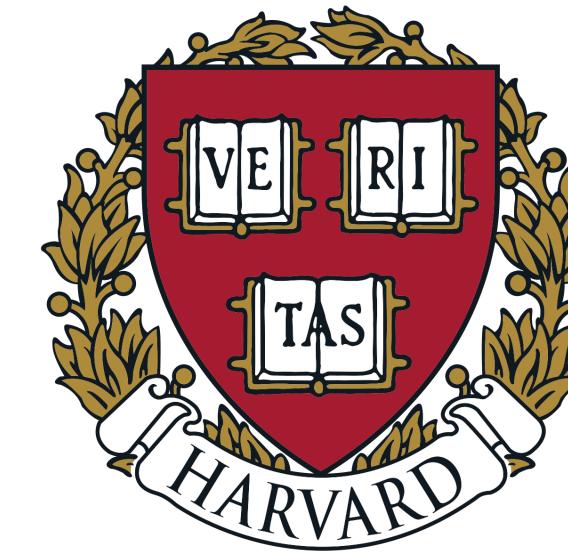
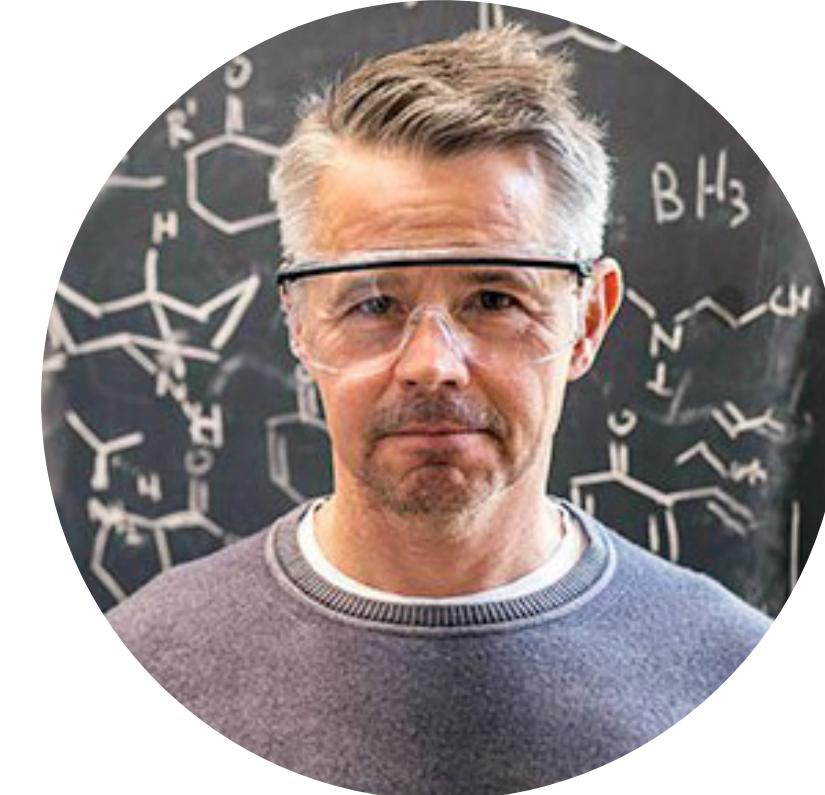


1998

# Andrew G. Myers



Assistant, Associate, and  
then Full Professor (1994)



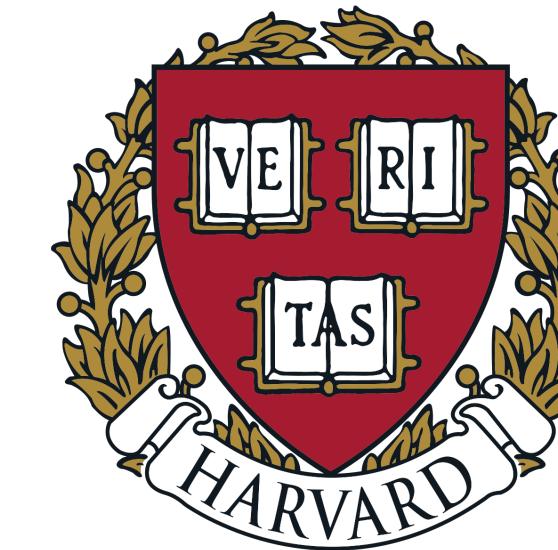
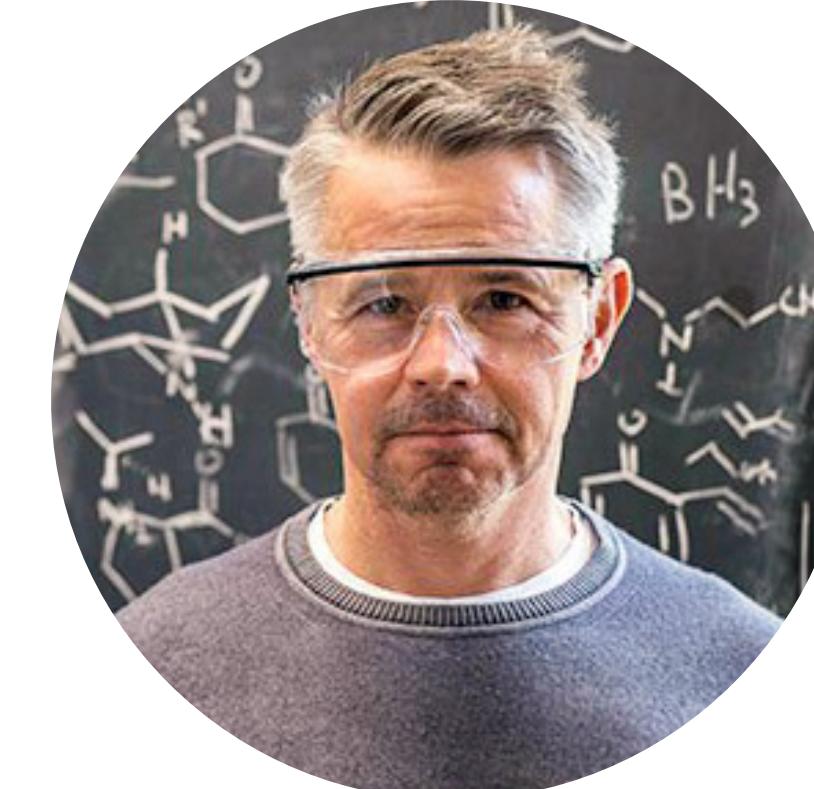
Department Chair 2007-2010



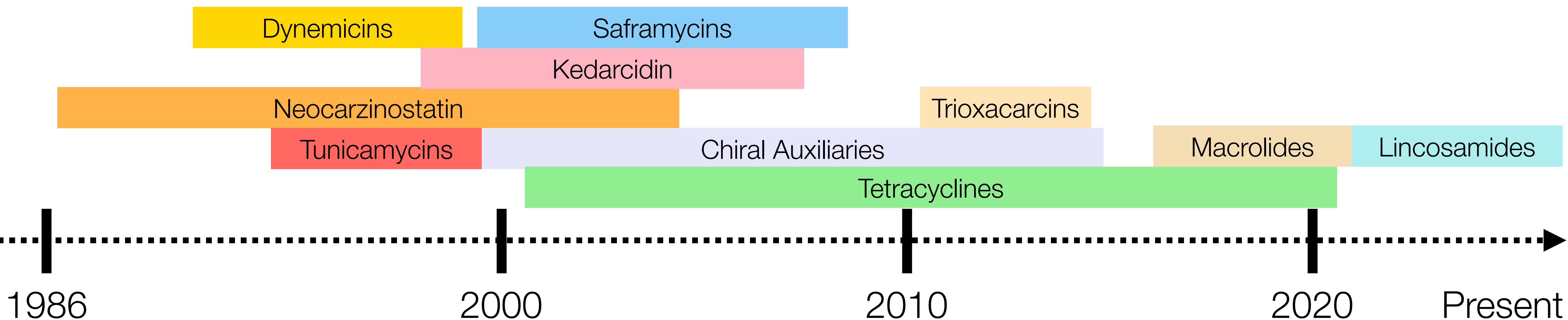
# Andrew G. Myers



Assistant, Associate, and  
then Full Professor (1994)



Department Chair 2007-2010



# Not Covered Today

avrainvillamide and stephacidin

cytochalasin

cortistatin

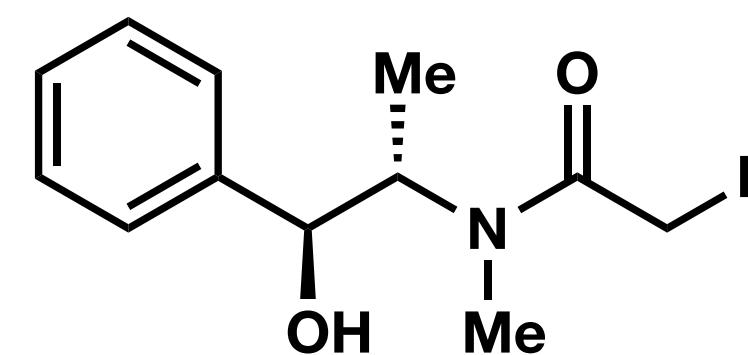
salinosporamide A

tunicamycins

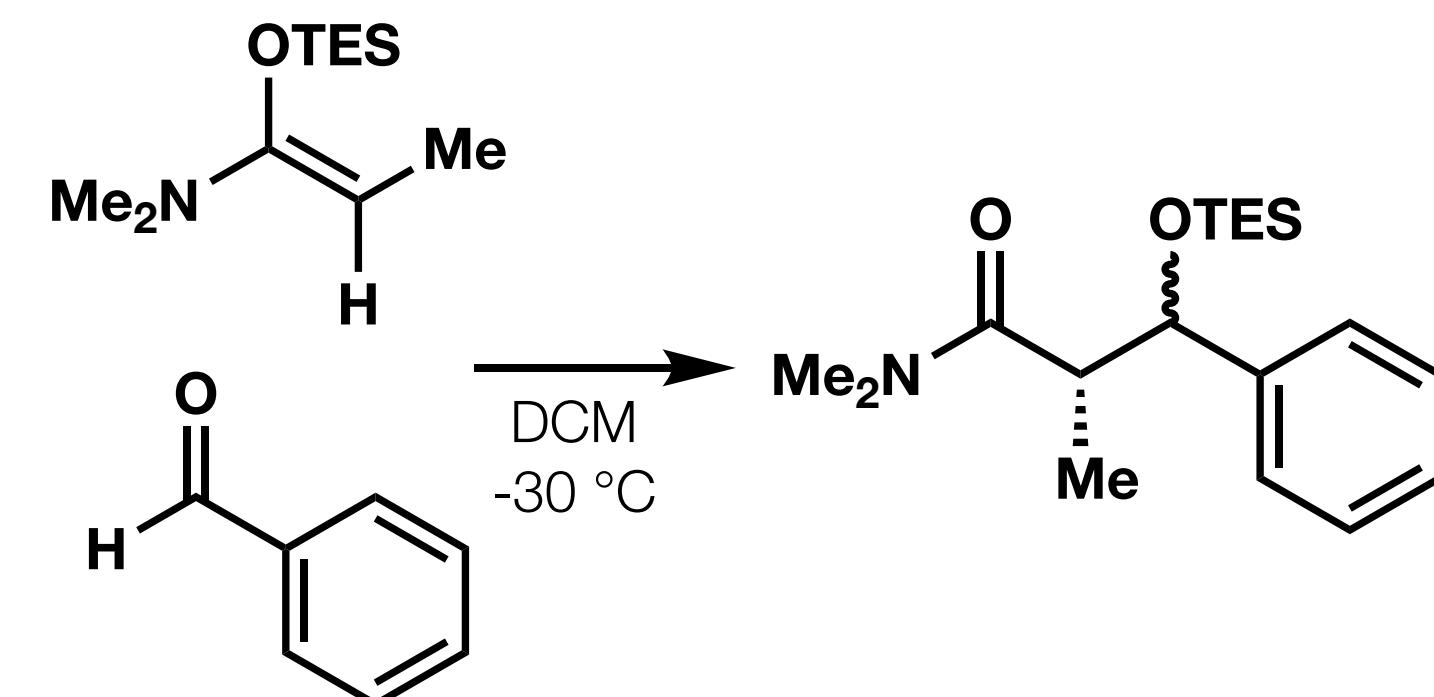
most method development and reagent development

# Method Development

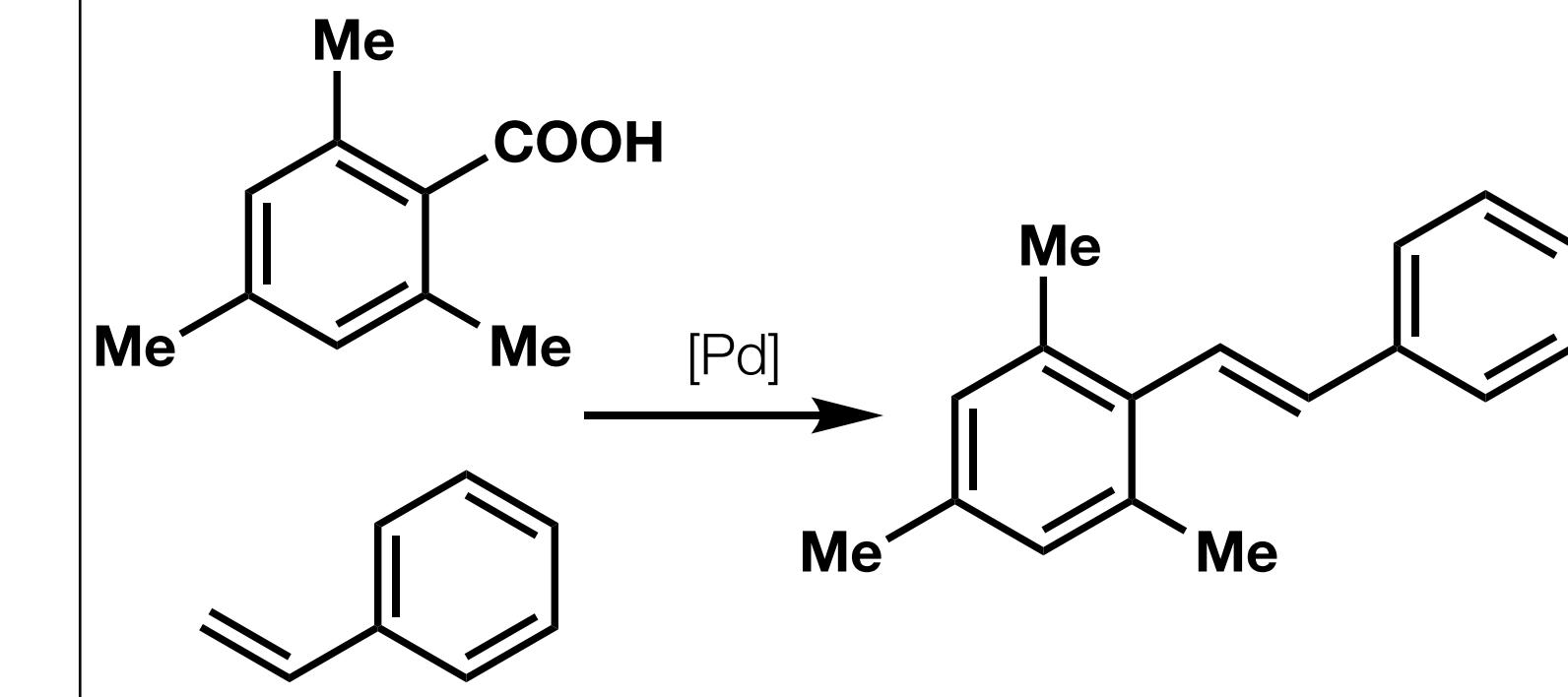
## Chiral Auxiliary



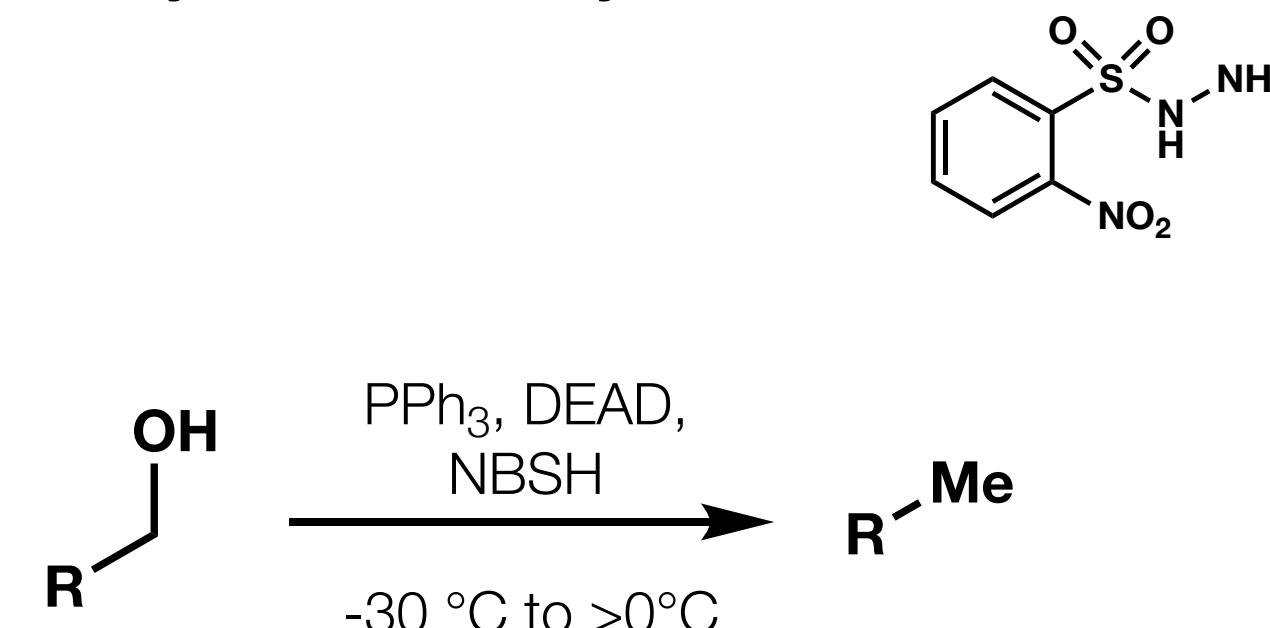
## Silicon-Directed Aldol



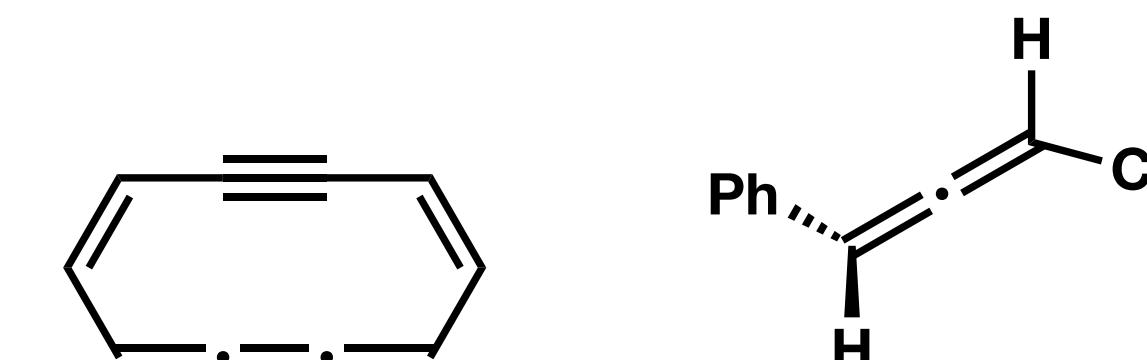
## Decarboxylative Palladiation



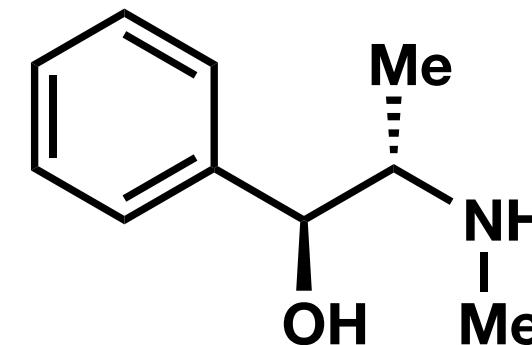
## Hydrazine/Hydrazone



## Allene and Ene-Diyne Synthesis



# Pseudoephedrine Auxiliary



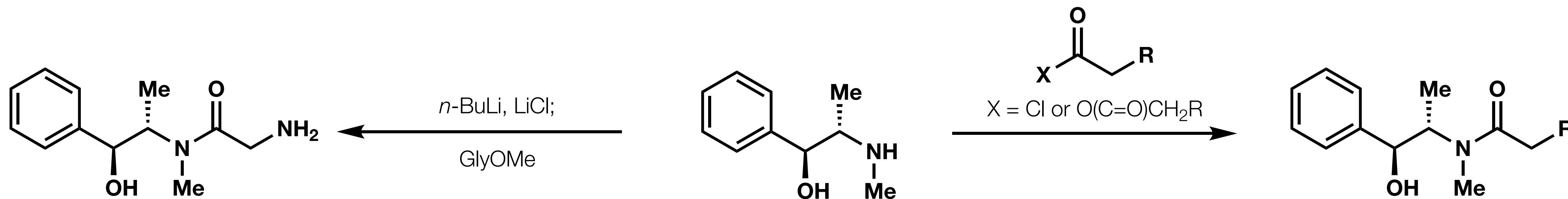
pseudoephedrine

The racemic mixture is also known as Sudafed, which is commonly used as an oral decongestant

Used to be widely available and cheap, but since it can be used to make meth, it is now quite hard to purchase

First reported as an effective auxiliary in 1994 by Myers

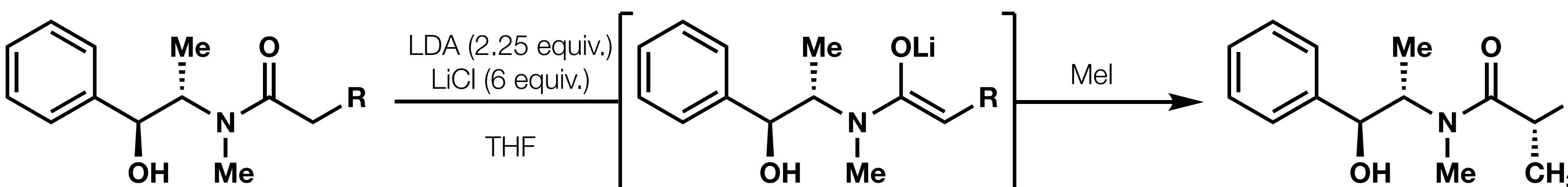
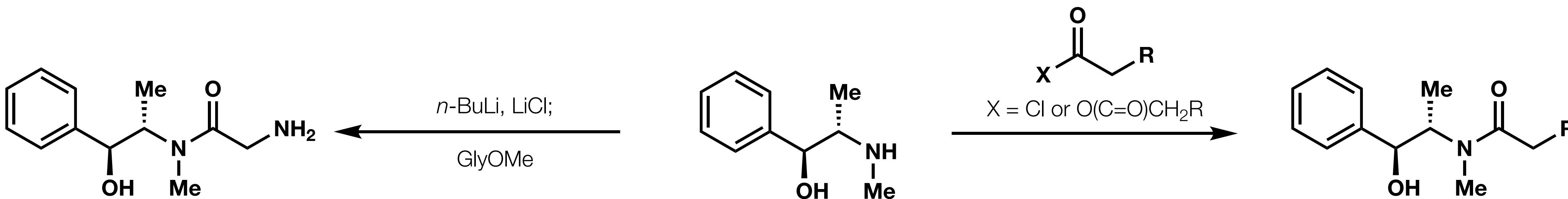
# Pseudoephedrine Auxiliary



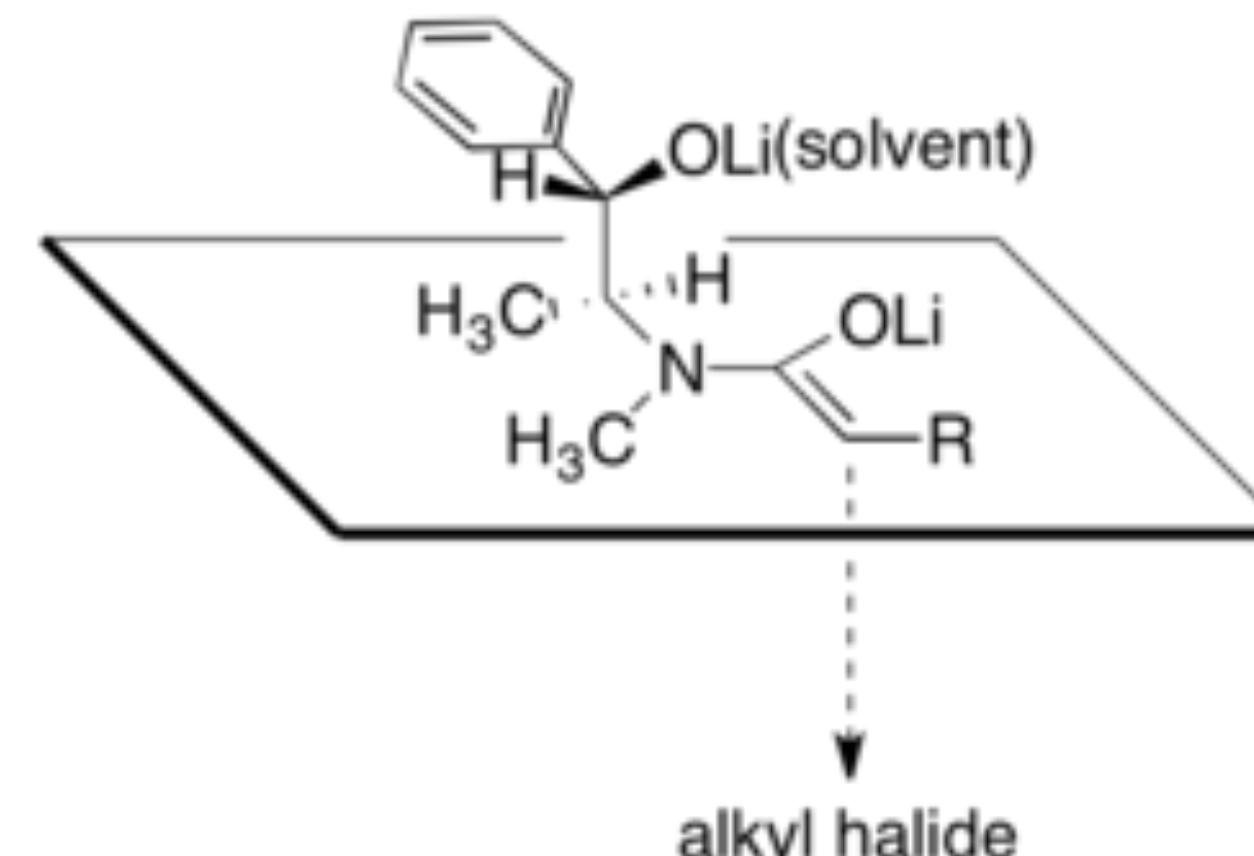
*dehydrated starting material shown, but hydrate  
can also be used directly with LiHMDS*

Tet. Lett. **1995**; 36(26): 4555–4558.  
JACS. **1995**; 117(32): 8488–8489.  
Tet. Lett. **1995**; 36(52): 9429–9432.  
JOC. **1999**; 64(9): 3322–3327.

# Pseudoephedrine Auxiliary



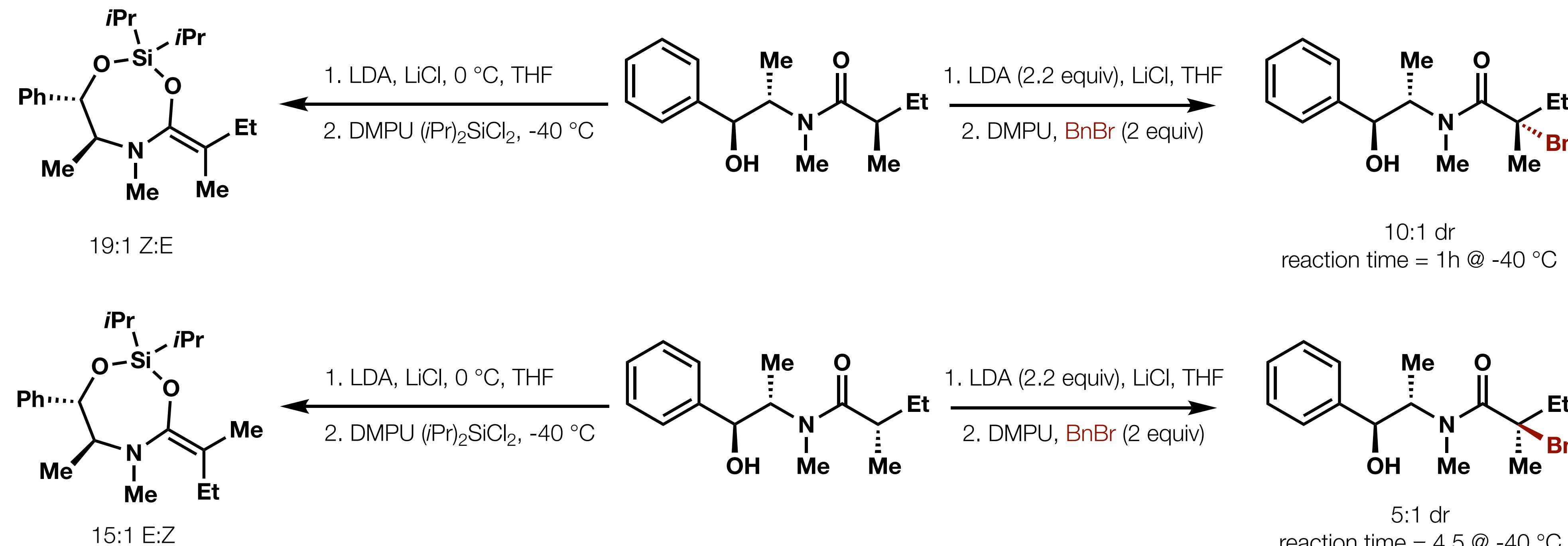
94% yield  
94% d.e.



\*Reactions in the presence of less than 4 equiv of LiCl are slower and do not proceed to completion; the diastereoselectivity of the alkylation reactions do not appear to be greatly affected by the concentration of LiCl

*Tet. Lett.* **1995**; 36(26): 4555–4558.  
*JACS.* **1995**; 117(32): 8488–8489.  
*Tet. Lett.* **1995**; 36(52): 9429–9432.  
*JOC.* **1999**; 64(9): 3322–3327.

# Quaternary Centers

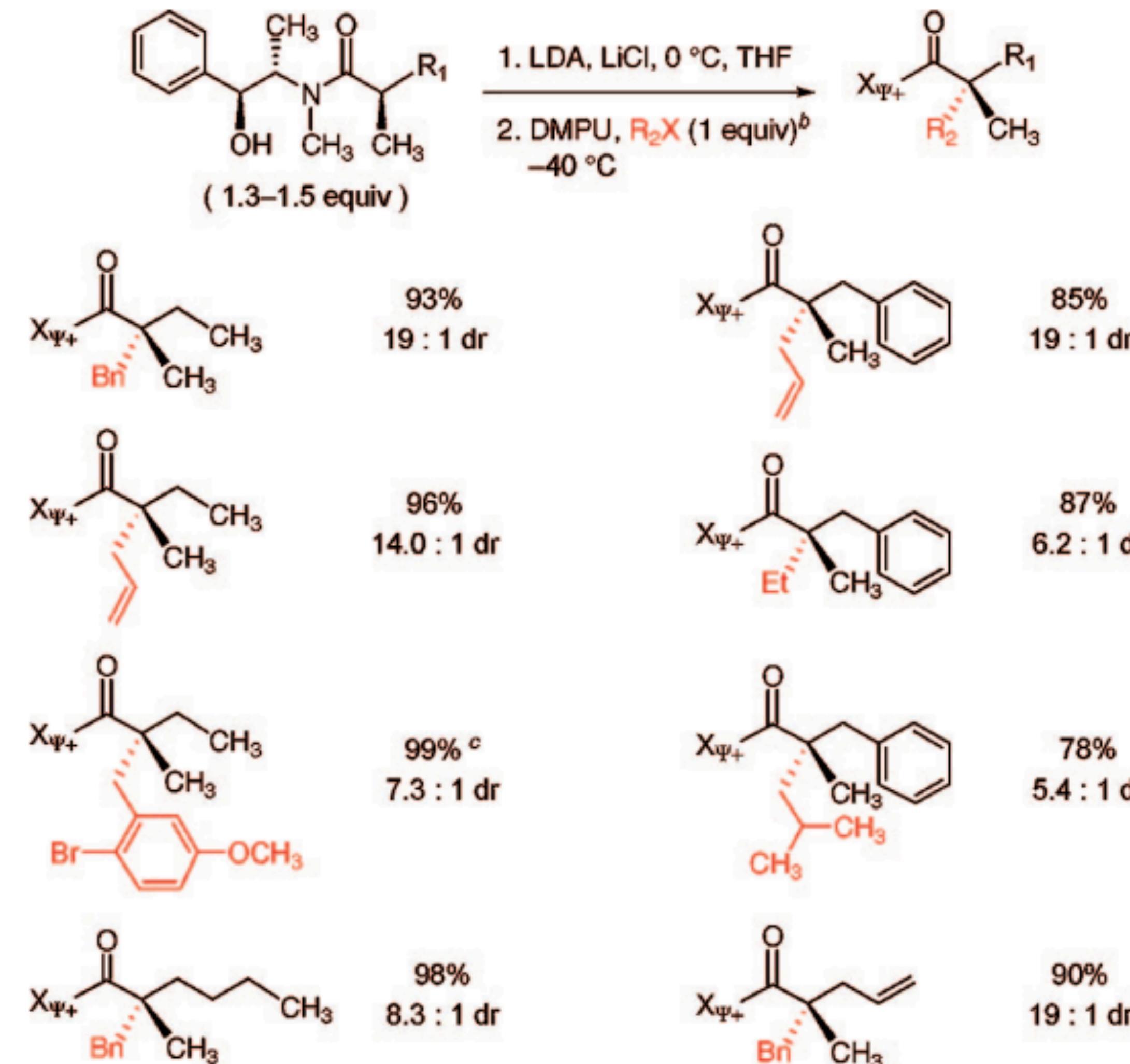


The E and Z enolates are both alkylated from the enolate π-face opposite the alkoxide side chain

Interesting Observation: The Z enolate reacts about 4x faster than the E enolate and is more diastereoselective at lower conversion, which led the authors to use excess enolate for higher diastereoselectivity

# Quaternary Centers

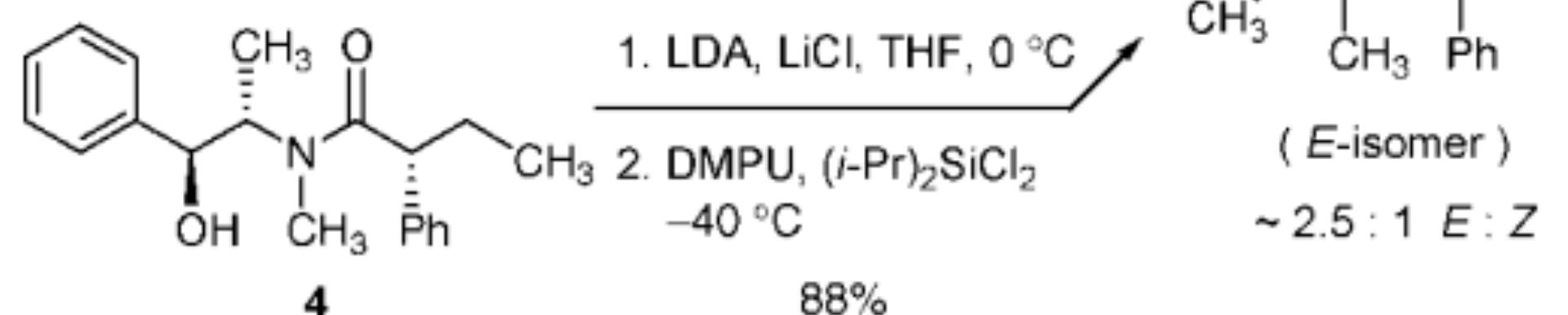
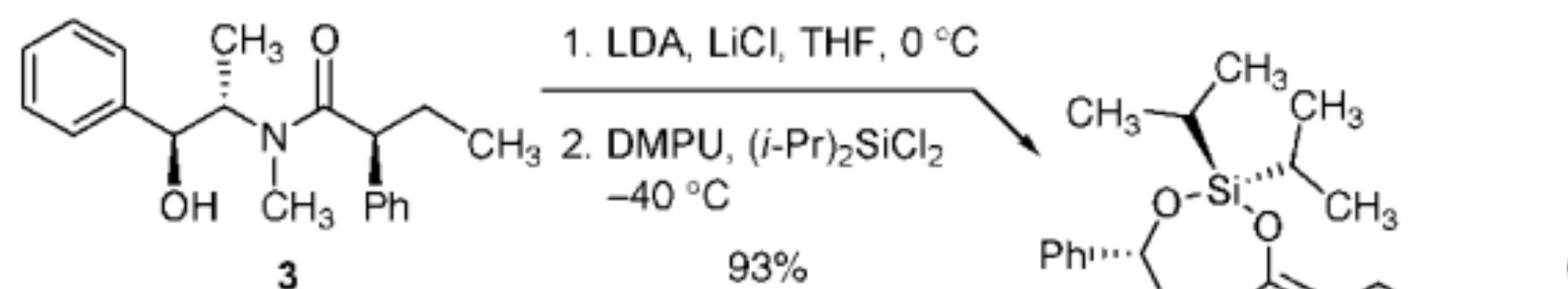
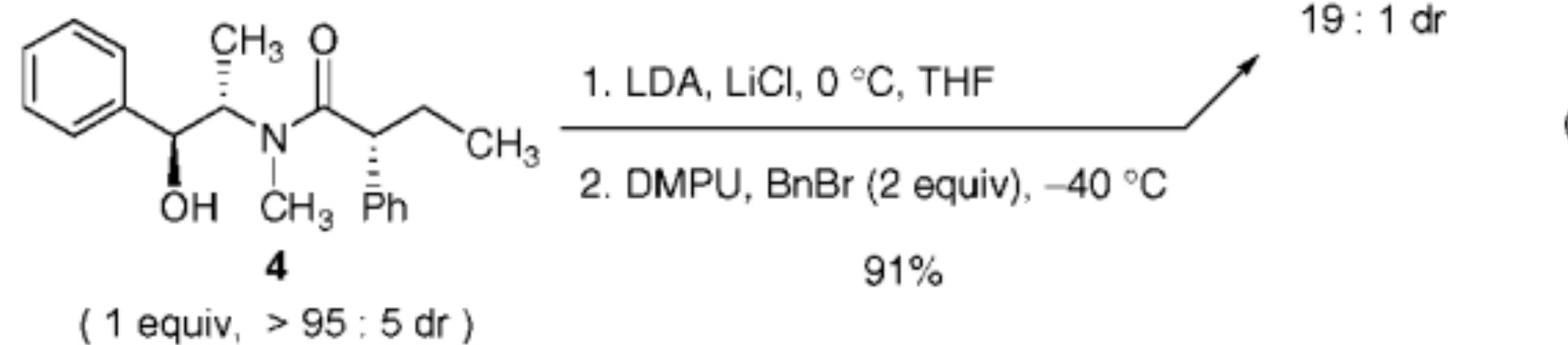
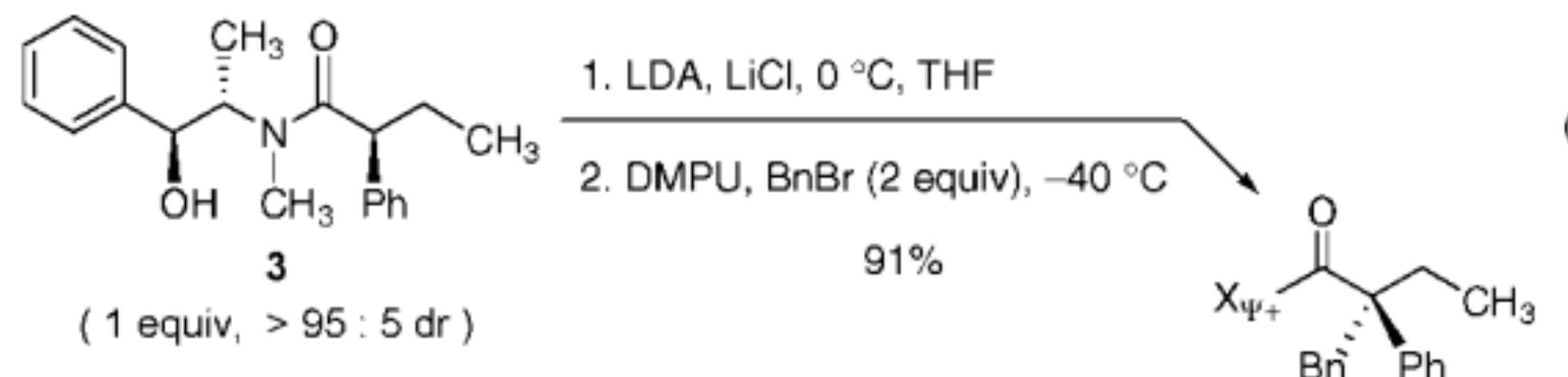
Interesting Observation: The Z enolate reacts about 4x faster than the E enolate and is more diastereoselective at lower conversion, which led the authors to use excess enolate for higher diastereoselectivity



JACS. **2008**, 130, 13231–13233

# Quaternary Centers

Another Interesting Observation: Both diastereomers of **alpha-phenyl amides** converge to the same product, suggesting that the E- and Z-alpha-phenyl-alpha-ethyl enolate intermediates might interconvert under the reaction conditions. This is supported by silanol trapping



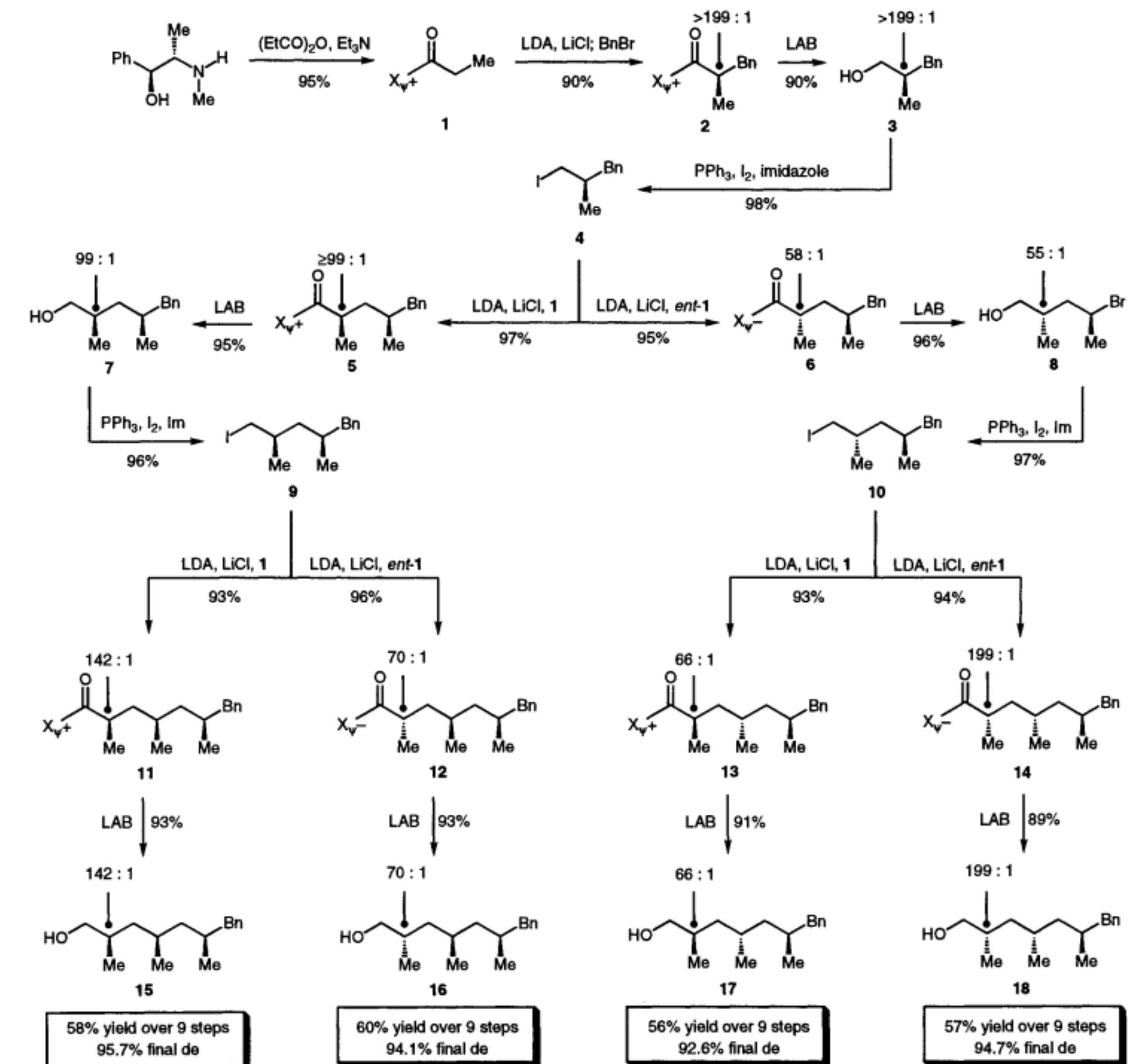
# Iteration

Iterative addition to set multiple stereo centers

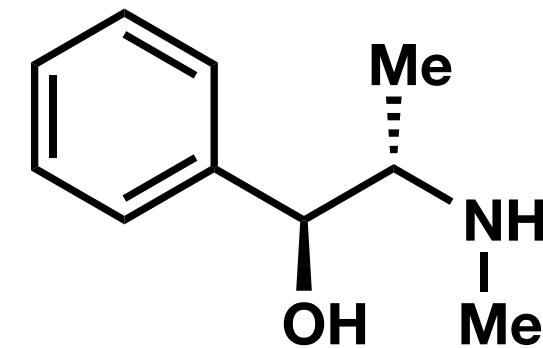
Strategy could be used to iteratively alkylate,  
reduce, alkylate, repeat

Enables asymmetric synthesis of 1,3-dialkyl  
substituted carbon chains of any stereochemical  
configuration

Facilitated by the development of LAB  
(LiH<sub>2</sub>NBH<sub>3</sub>), a reductant that can selectively  
reduce tertiary amides to primary alcohols

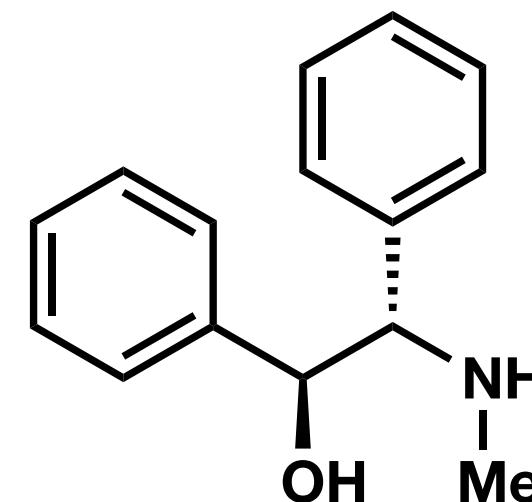


# Pseudoephedrine Auxiliary



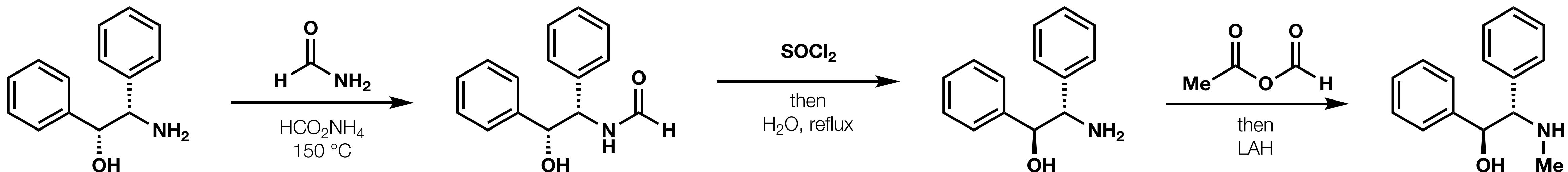
Restricted

# Pseudoephedrine Auxiliary



Pseudoephedrine

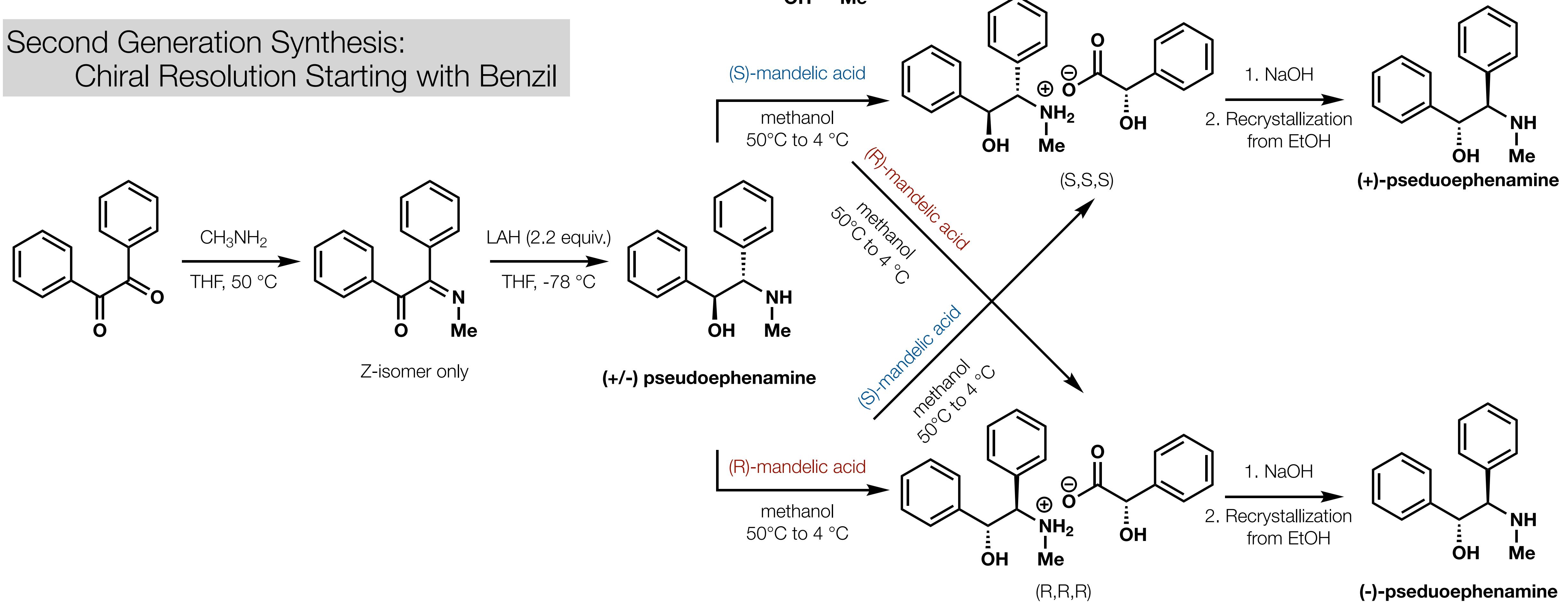
## First Generation Synthesis



(1R, 2S)-1,2-diphenyl-2-aminoethanol  
~\$1-2 per gram

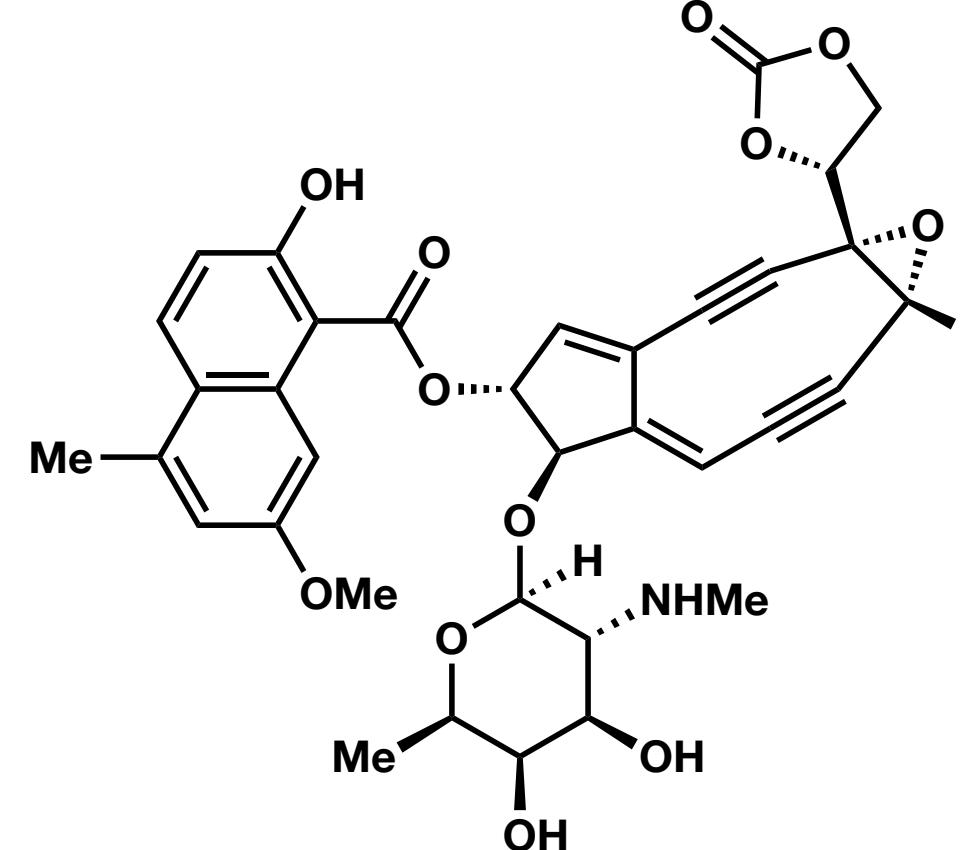
# Pseudoephedrine Auxiliary

Second Generation Synthesis:  
Chiral Resolution Starting with Benzil

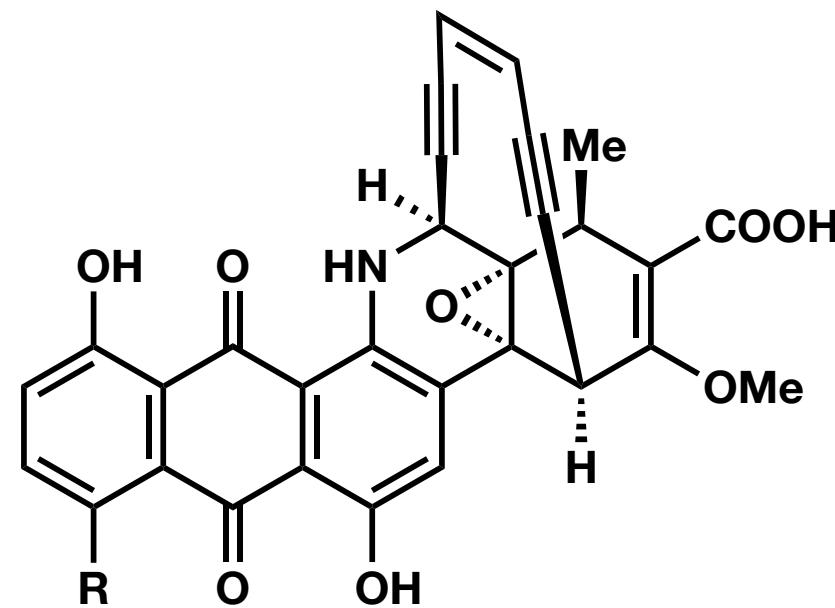


# Ene-diyynes

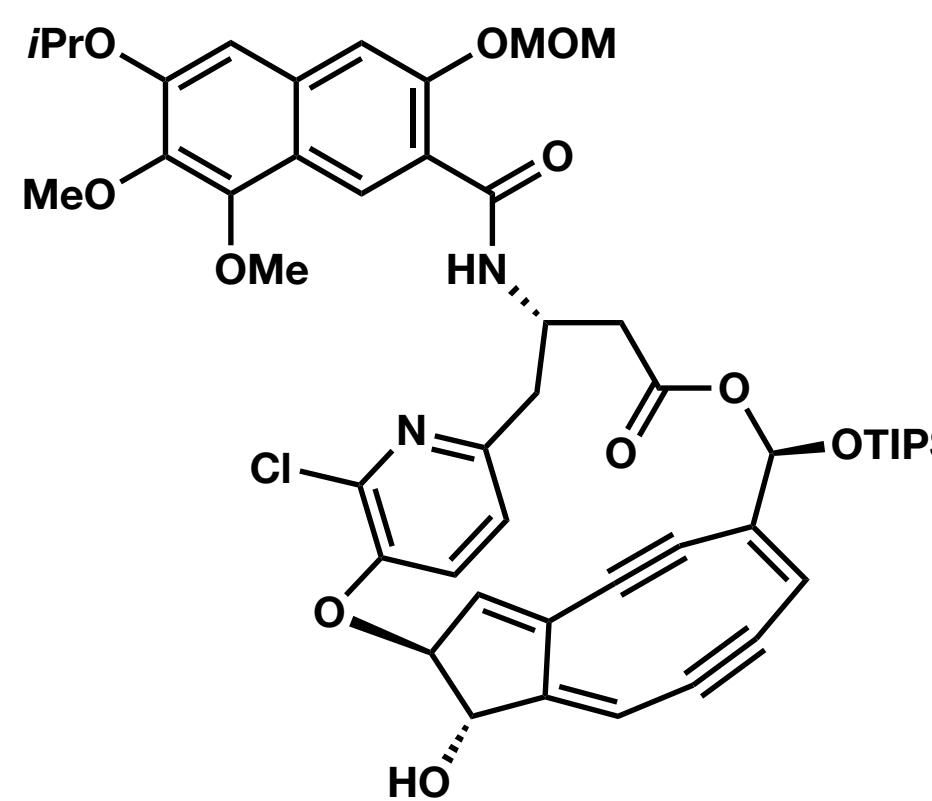
neocarzinostatin chromophore



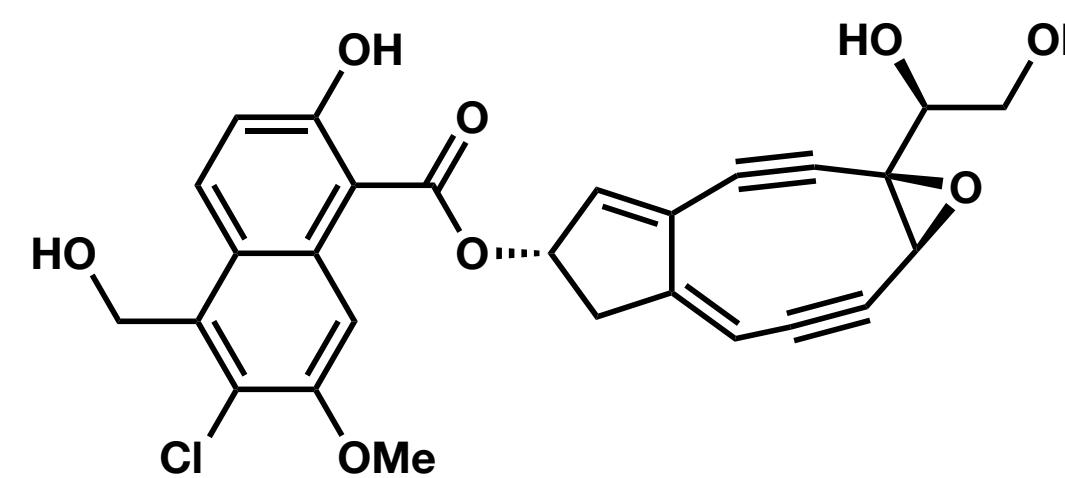
dynemicin A



kedarcidin chromophore



N1999A2



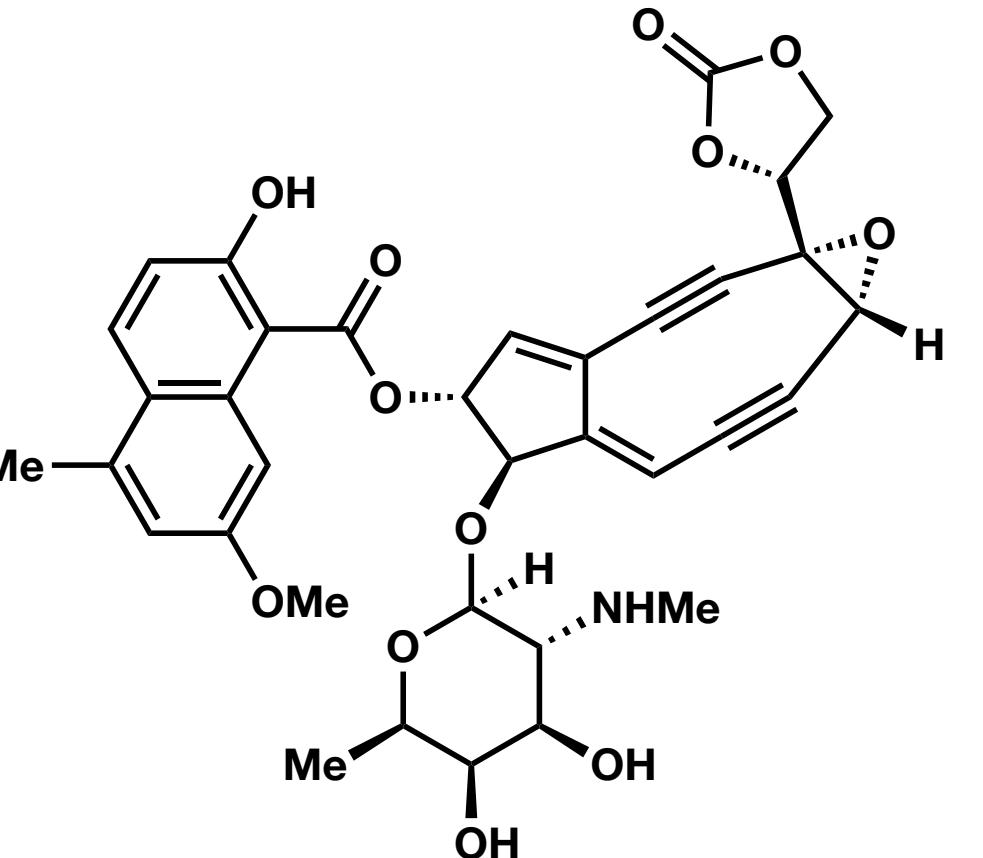
# Ene-diyynes

## Synthetically Challenging

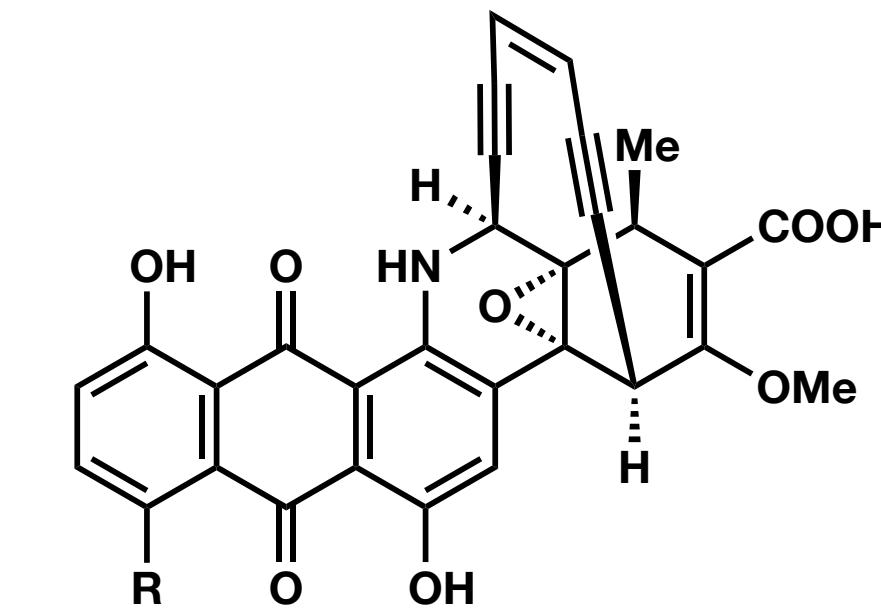
Chemically **unstable** compounds - often cannot be stored neat in the absence of radical inhibitors

Analog development demands flexible, **convergent** synthetic approaches

neocarzinostatin chromophore



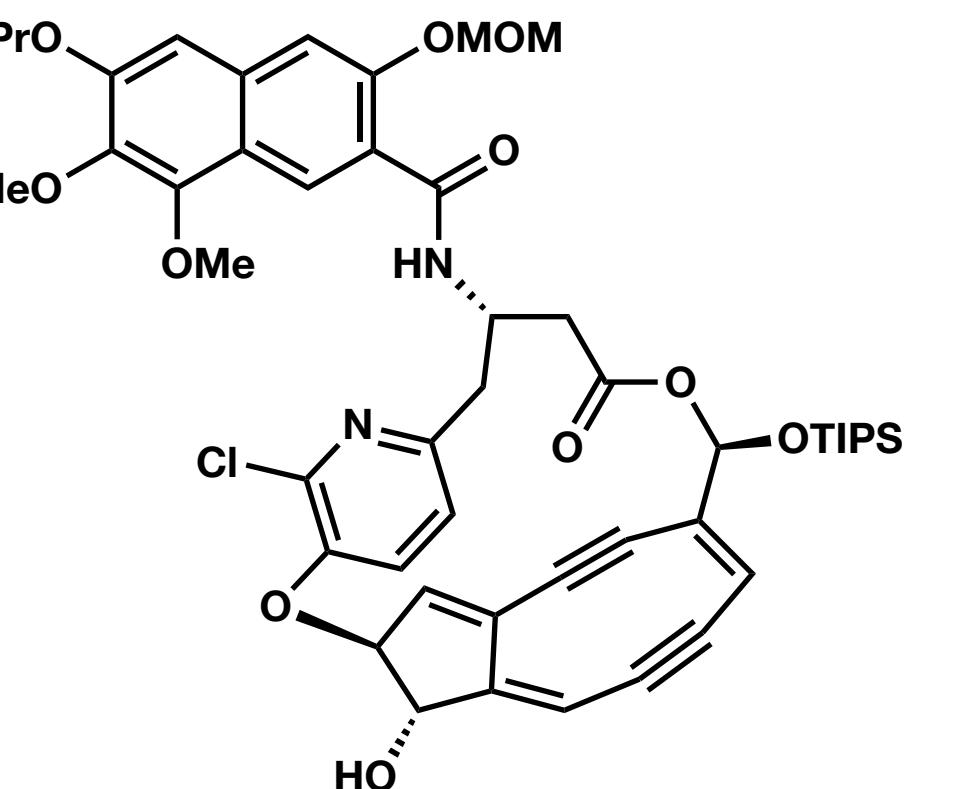
dynemicin A



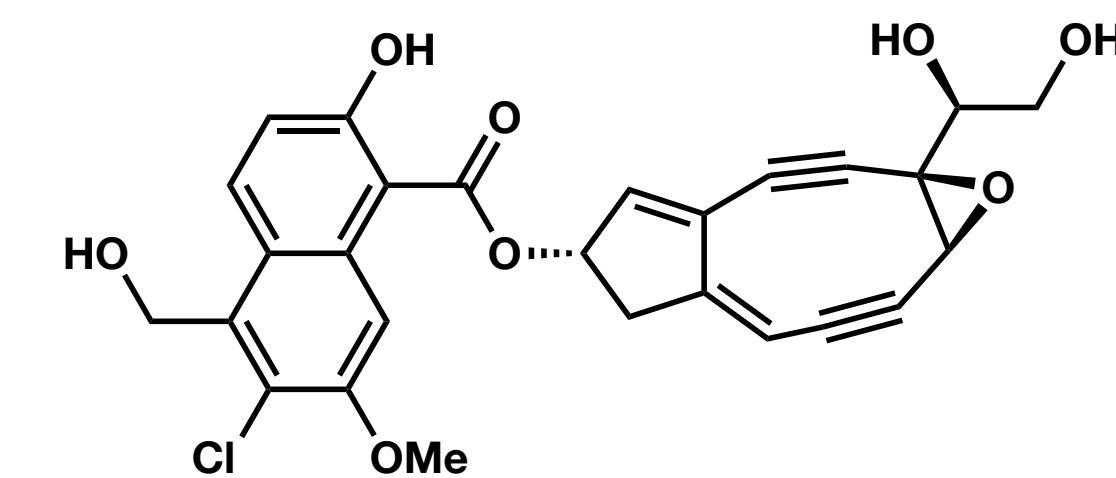
## General Reactivity

Nucleophilic attack triggers conformation change to initiate a **Bergman Cyclization**, forming diradicals that abstract hydrogens from DNA, resulting in  **$\beta$ -scission** of the DNA backbone

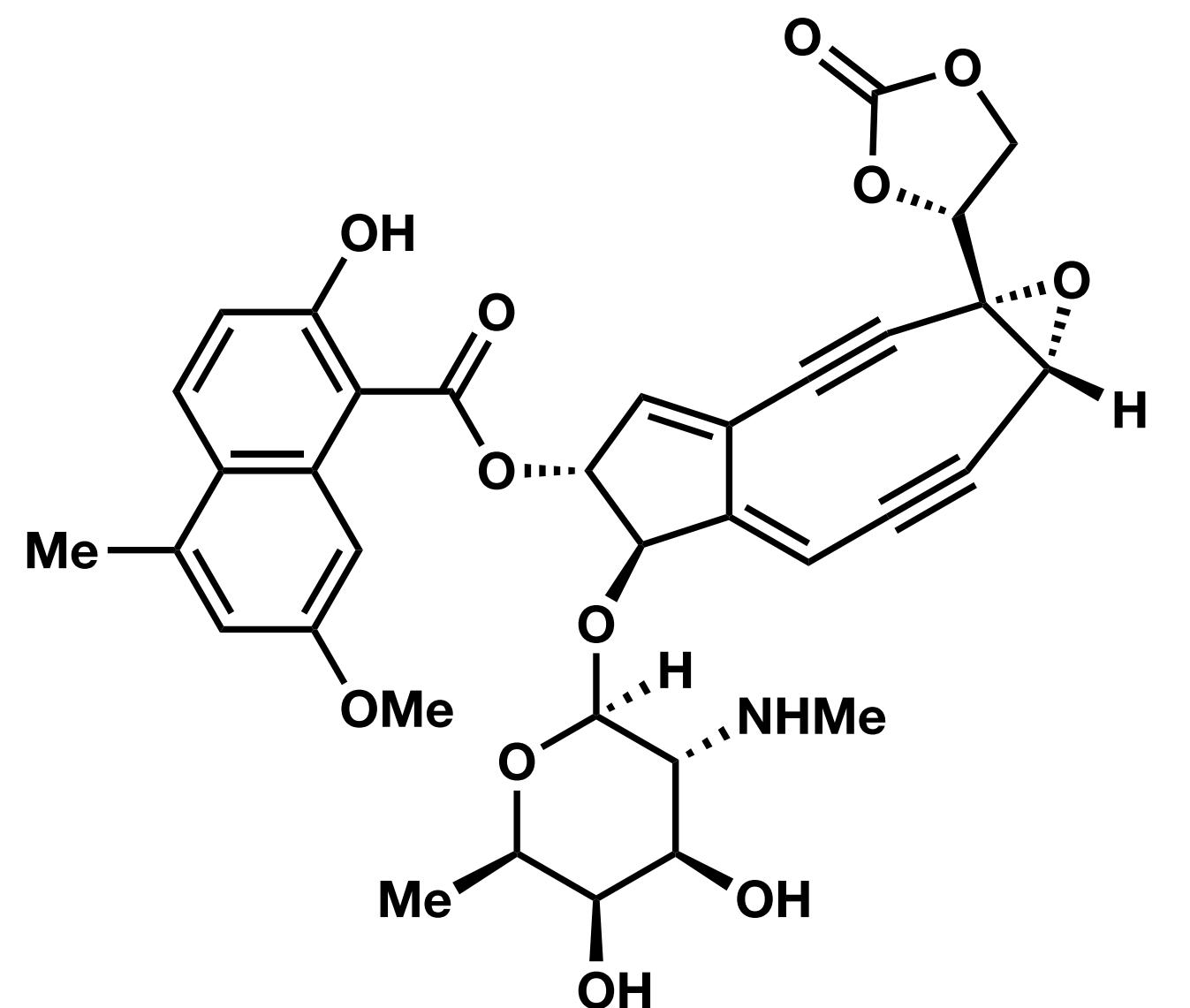
kedarcidin chromophore



N1999A2

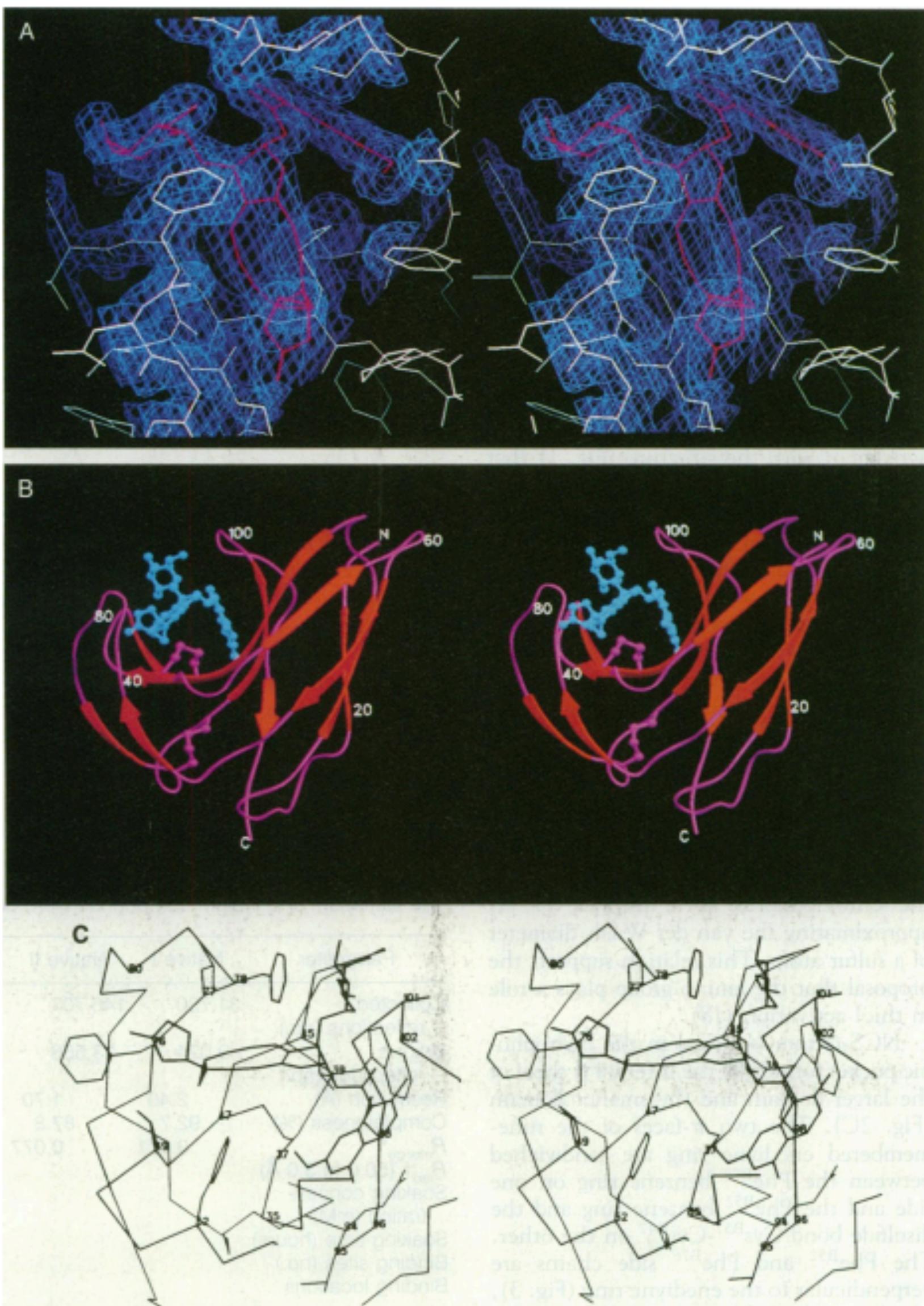


# Neocarzinostatin



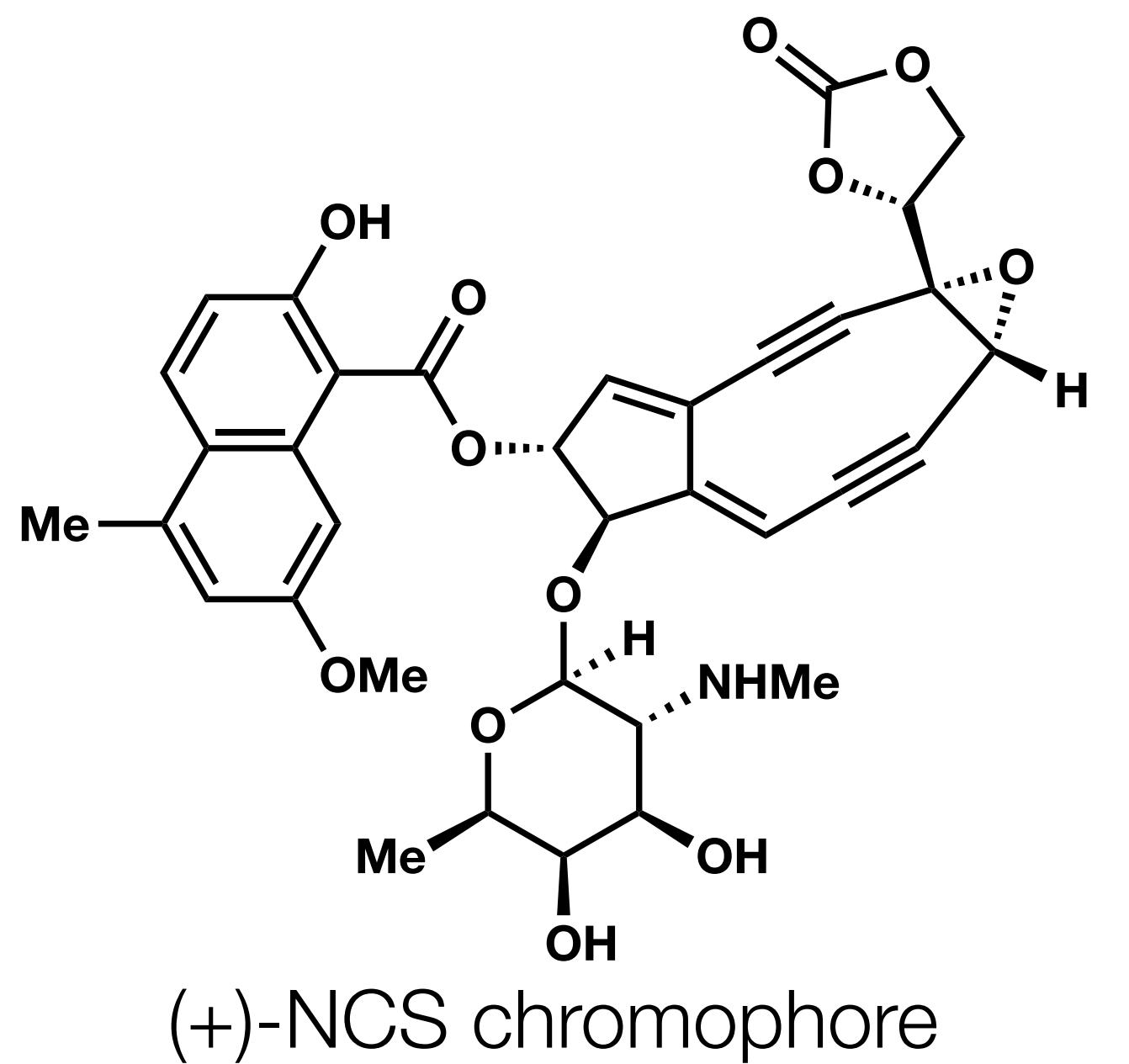
(+)-NCS chromophore

- Isolated from *Streptomyces carzinostaticus*

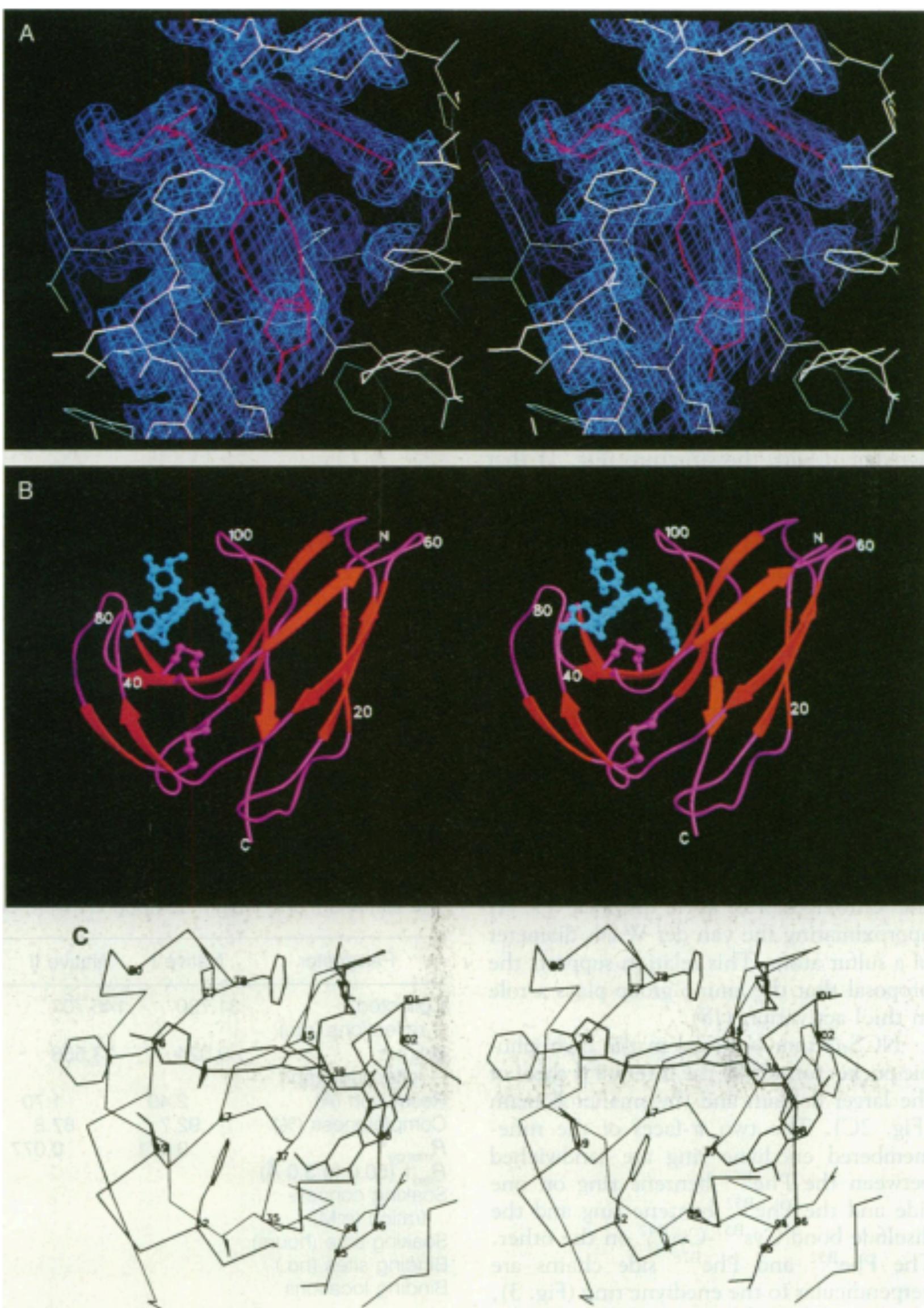


**Fig. 2.** (A) Stereoview of the electron density map (blue) of the region surrounding NCS-chrom (red) and neighboring protein (white). The map was calculated with  $2|F_o| - |F_c|$  coefficients for data between 5 to 1.8 Å resolution and contoured at  $1.0\sigma$ . (B) Stereoview of the polypeptide fold of holo-NCS. NCS-chrom and cysteine residues are represented by ball-and-stick models. (C) Stereoview of the protein environment of NCS-chrom. Residues within 4 Å of the NCS-chrom are labeled. Both (B) and (C) were drawn with the program SETOR (33).

# Neocarzinostatin

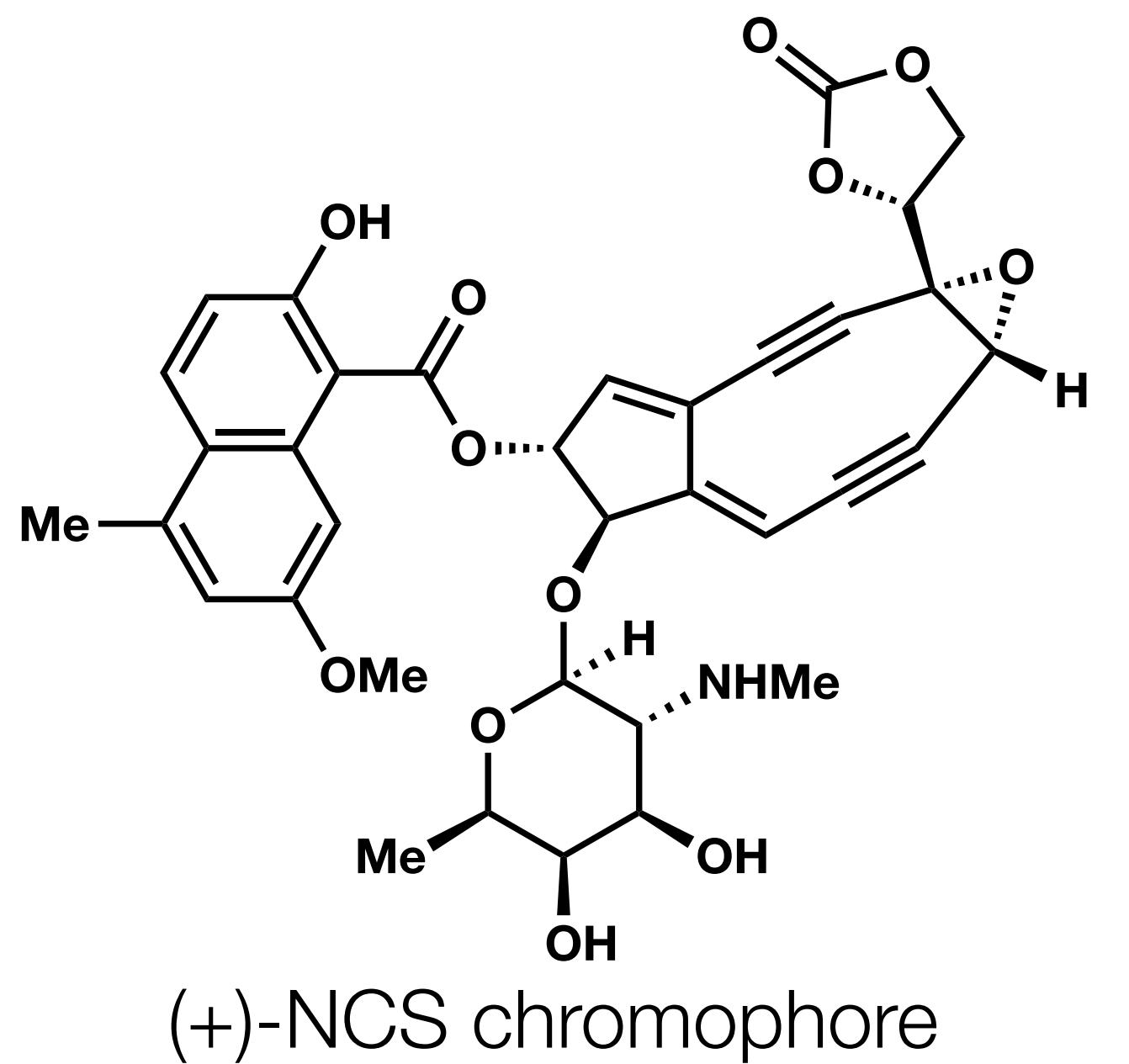


- Isolated from *Streptomyces carzinostaticus*
- Composed of a 113-amino acid protein component (apoNCS) and a chromophore component (NCS-chrom)

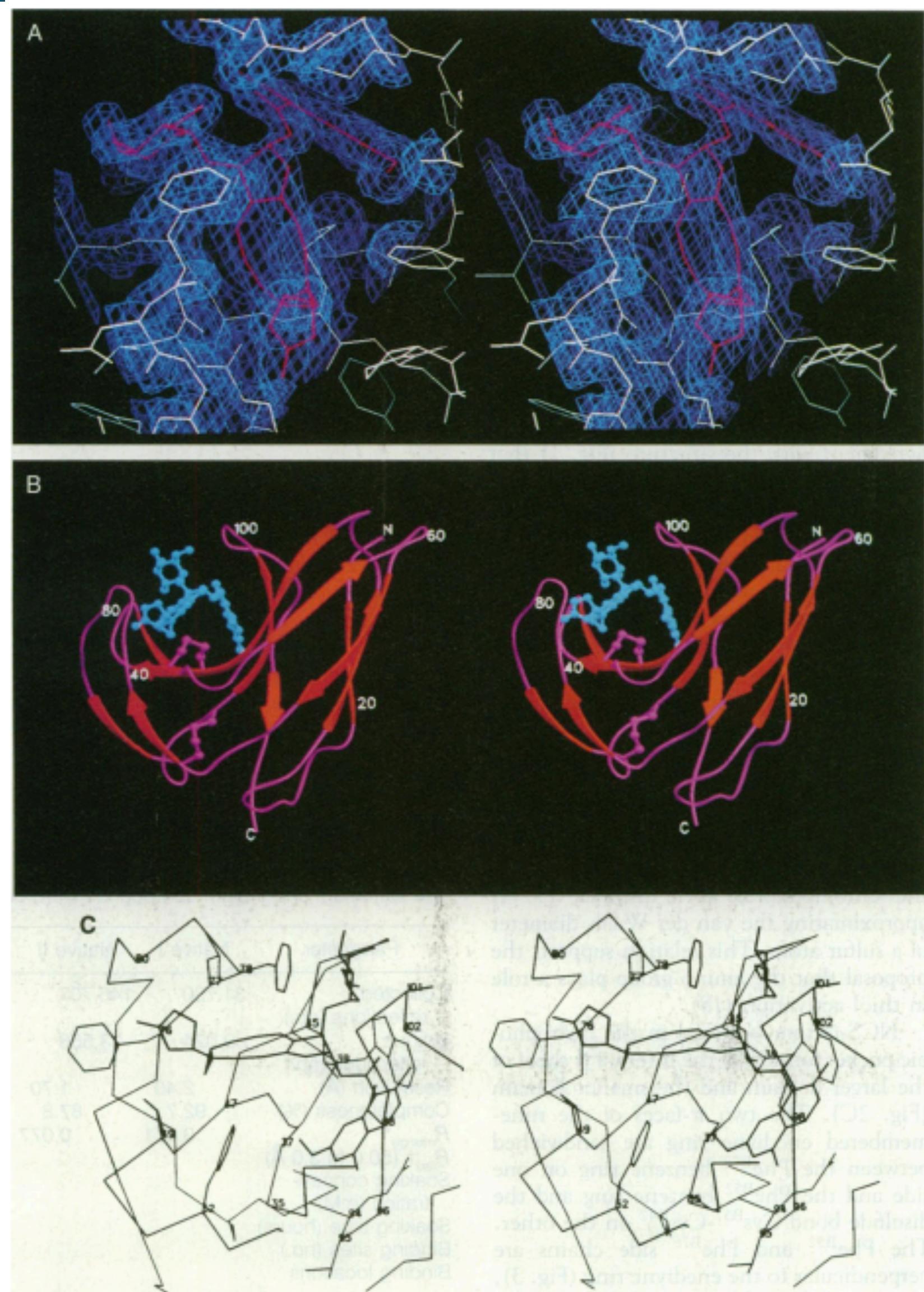


**Fig. 2.** (A) Stereoview of the electron density map (blue) of the region surrounding NCS-chrom (red) and neighboring protein (white). The map was calculated with  $2|F_o| - |F_c|$  coefficients for data between 5 to 1.8 Å resolution and contoured at  $1.0\sigma$ . (B) Stereoview of the polypeptide fold of holo-NCS. NCS-chrom and cysteine residues are represented by ball-and-stick models. (C) Stereoview of the protein environment of NCS-chrom. Residues within 4 Å of the NCS-chrom are labeled. Both (B) and (C) were drawn with the program SETOR (33).

# Neocarzinostatin

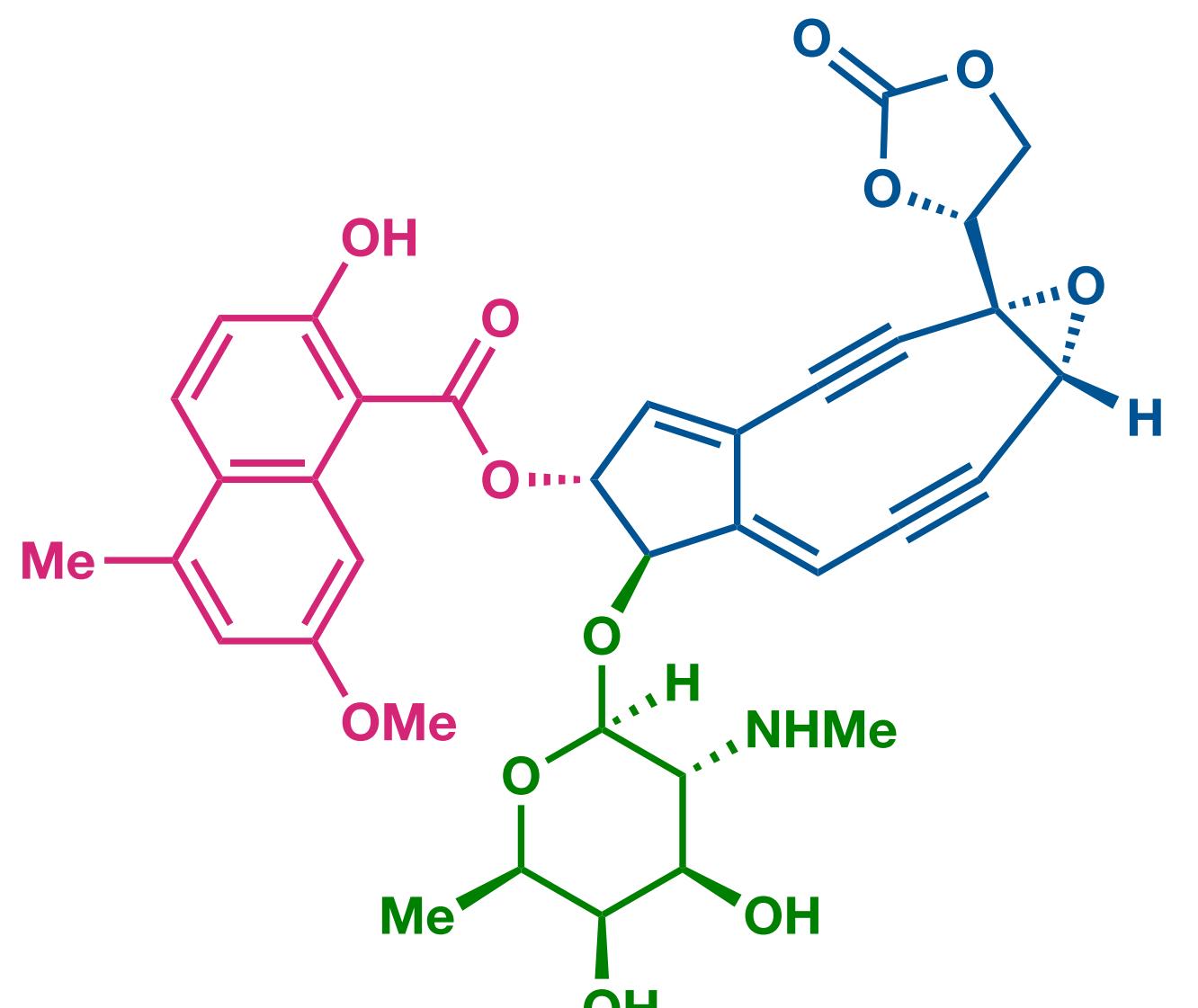


- Isolated from *Streptomyces carzinostaticus*
- Composed of a 113-amino acid protein component (apoNCS) and a chromophore component (NCS-chrom)
- Potent DNA cleaving agent, mechanism of action was heavily studied by Kappen, Goldberg, Saito, **Myers**, and others.



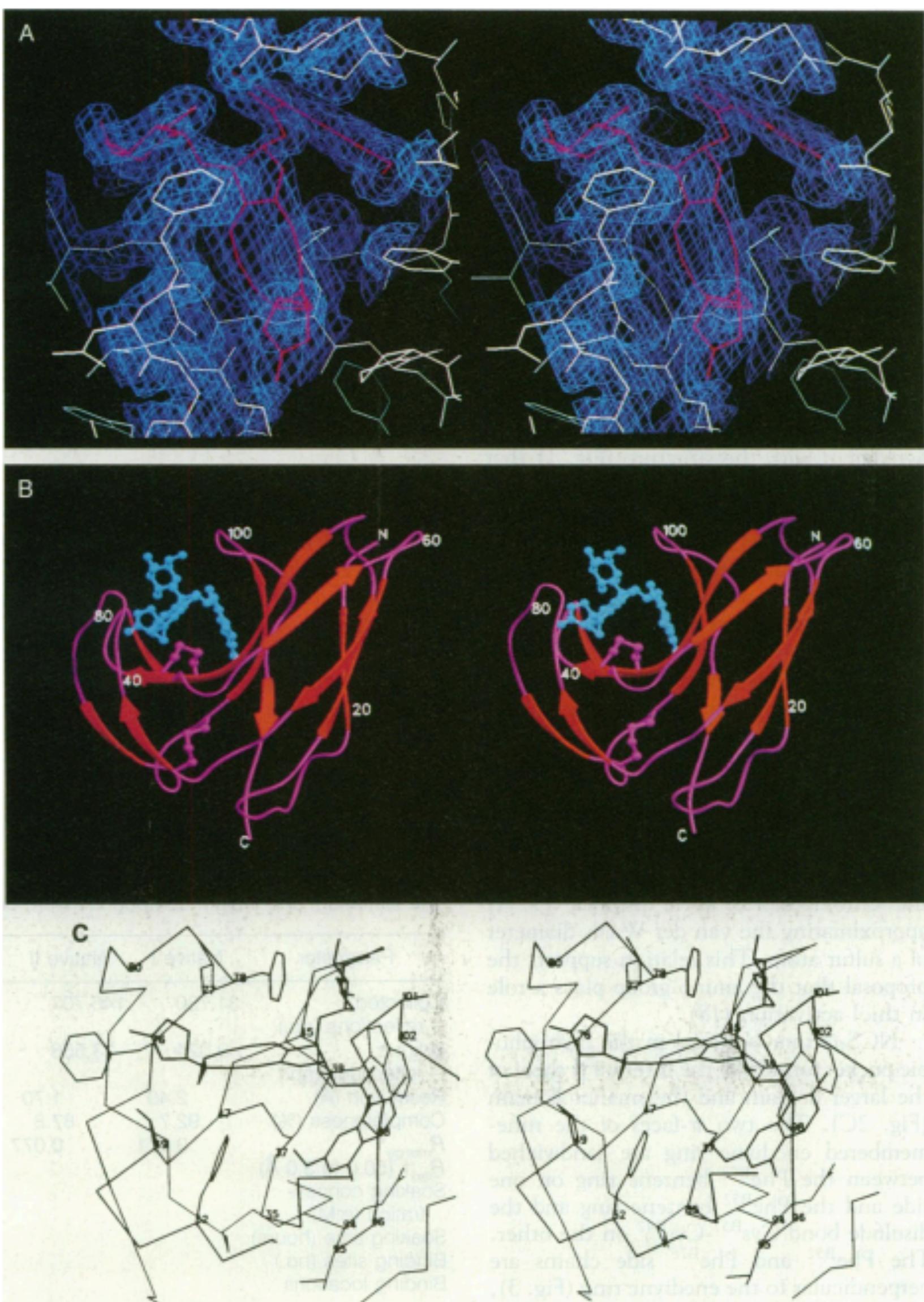
**Fig. 2.** (A) Stereoview of the electron density map (blue) of the region surrounding NCS-chrom (red) and neighboring protein (white). The map was calculated with  $2|F_o| - |F_c|$  coefficients for data between 5 to 1.8 Å resolution and contoured at  $1.0\sigma$ . (B) Stereoview of the polypeptide fold of holo-NCS. NCS-chrom and cysteine residues are represented by ball-and-stick models. (C) Stereoview of the protein environment of NCS-chrom. Residues within 4 Å of the NCS-chrom are labeled. Both (B) and (C) were drawn with the program SETOR (33).

# Neocarzinostatin



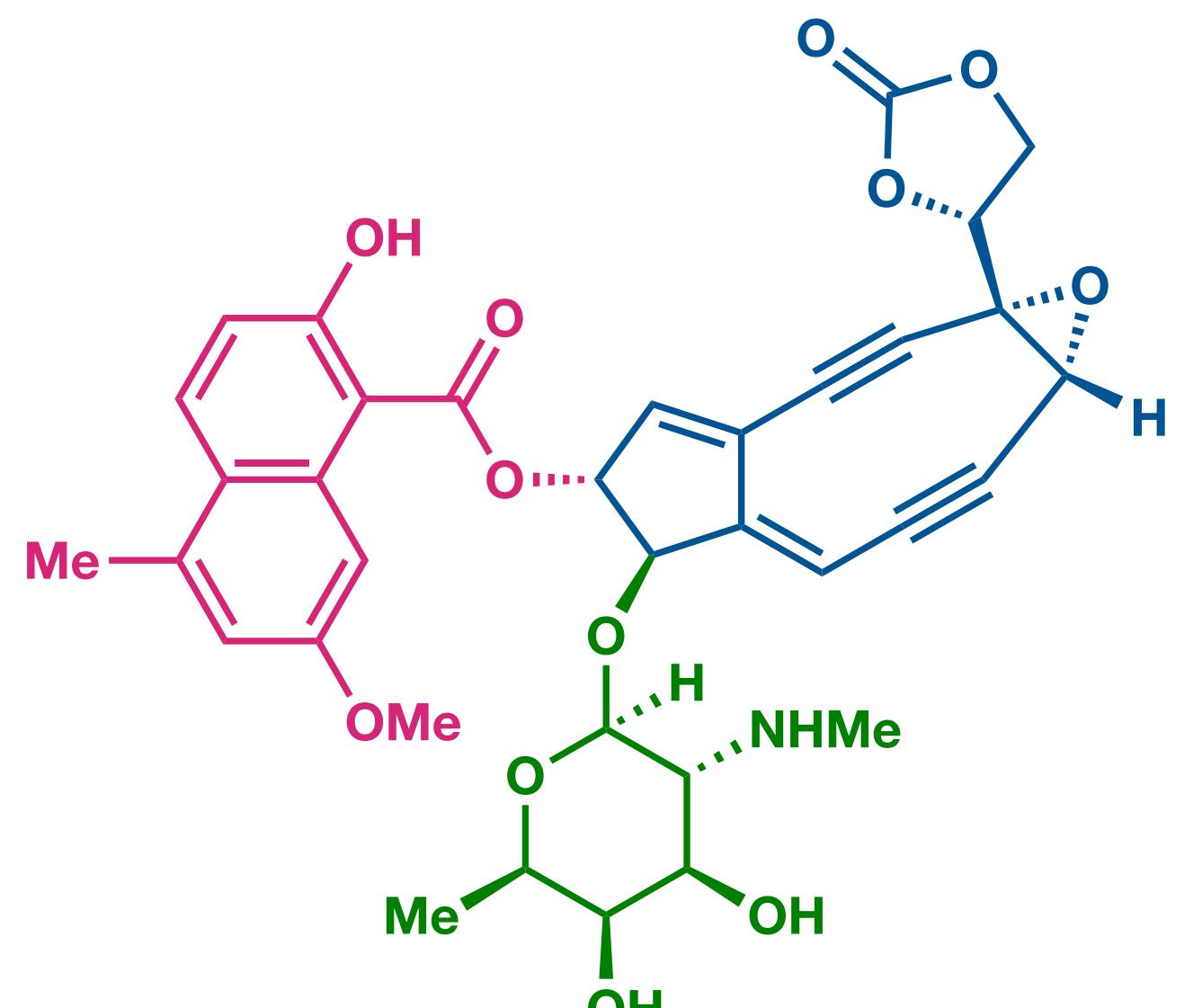
(+)-NCS chromophore

- Isolated from *Streptomyces carzinostaticus*
- Composed of a 113-amino acid protein component (apoNCS) and a chromophore component (NCS-chrom)
- Potent DNA cleaving agent, mechanism of action was heavily studied by Kappen, Goldberg, Saito, **Myers**, and others.
- Chromophore features a key **epoxy-enediyne core**, a **napthoic acid**, and an **aminosugar**



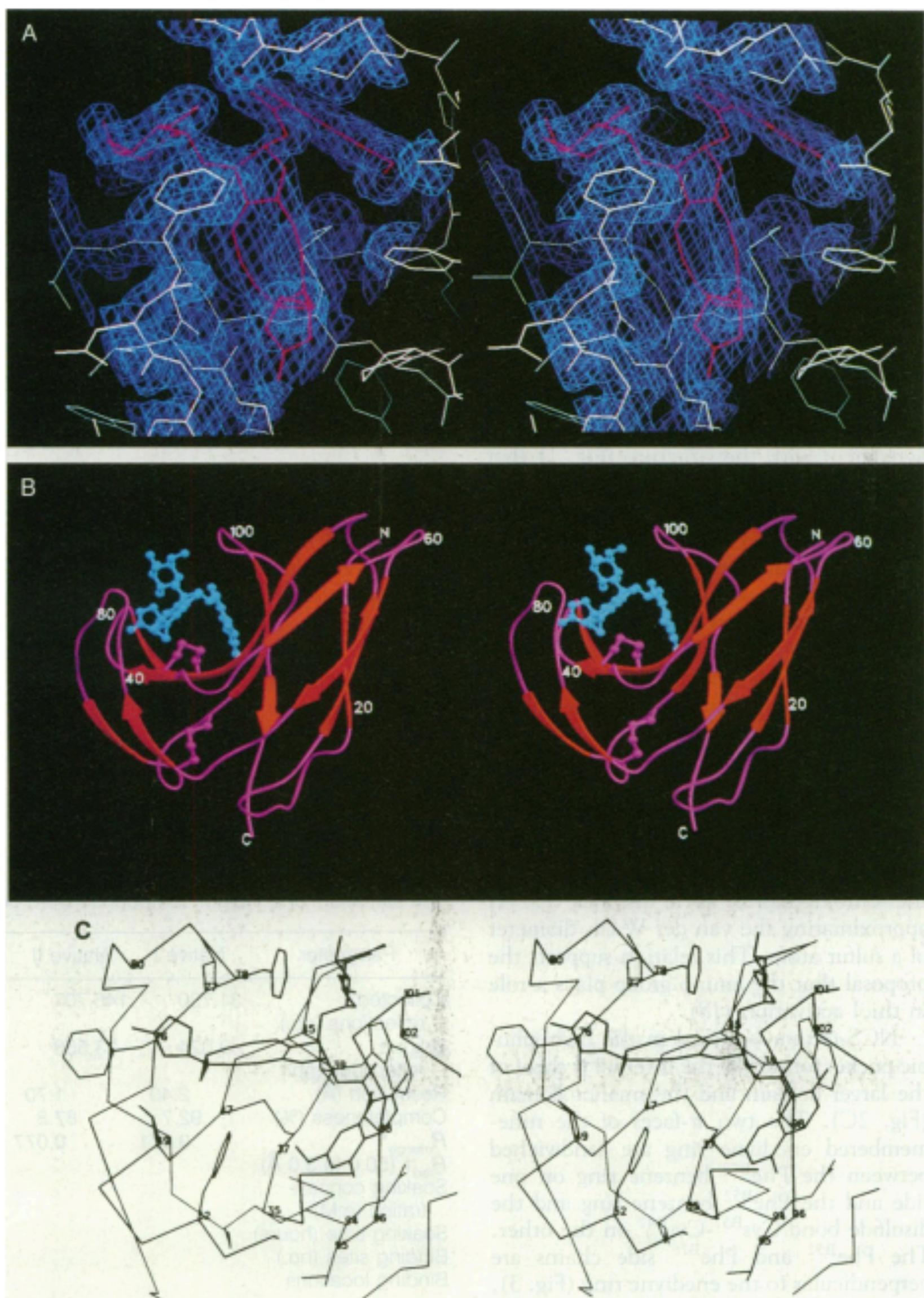
**Fig. 2.** (A) Stereoview of the electron density map (blue) of the region surrounding NCS-chrom (red) and neighboring protein (white). The map was calculated with  $2|F_o| - |F_c|$  coefficients for data between 5 to 1.8 Å resolution and contoured at  $1.0\sigma$ . (B) Stereoview of the polypeptide fold of holo-NCS. NCS-chrom and cysteine residues are represented by ball-and-stick models. (C) Stereoview of the protein environment of NCS-chrom. Residues within 4 Å of the NCS-chrom are labeled. Both (B) and (C) were drawn with the program SETOR (33).

# Neocarzinostatin



(+)-NCS chromophore

- Isolated from *Streptomyces carzinostaticus*
- Composed of a 113-amino acid protein component (apoNCS) and a chromophore component (NCS-chrom)
- Potent DNA cleaving agent, mechanism of action was heavily studied by Kappen, Goldberg, Saito, **Myers**, and others.
- Chromophore features a key **epoxy-enediyne core**, a **napthoic acid**, and an **aminosugar**
- Crystal structure of Holo NCS elucidated by Rees in 1993 by Rees (X-Ray, 1.8 Å resolution)



**Fig. 2.** (A) Stereoview of the electron density map (blue) of the region surrounding NCS-chrom (red) and neighboring protein (white). The map was calculated with  $2|F_o| - |F_c|$  coefficients for data between 5 to 1.8 Å resolution and contoured at  $1.0\sigma$ . (B) Stereoview of the polypeptide fold of holo-NCS. NCS-chrom and cysteine residues are represented by ball-and-stick models. (C) Stereoview of the protein environment of NCS-chrom. Residues within 4 Å of the NCS-chrom are labeled. Both (B) and (C) were drawn with the program SETOR (33).

# NCS - Mechanism of Action

## What was known prior to Myers' Independent Career

Mostly work by Kappen and Goldberg (1979-1980)

- Clear dependency on thiols for activation
- Reacting the chromophore with methyl thioglycolate leads to a compound that appears to be the adduct with two extra hydrogens
- Hypothesized that a radical intermediate abstracts the 5' hydrogen on the sugar backbone, leading to DNA cleavage
- DNA cleavage is not aerobically dependent
- Napthoate functionality was hypothesized to intercalate into the DNA minor groove

# NCS - Mechanism of Action

## What was known prior to Myers' Independent Career

Mostly work by Kappen and Goldberg (1979-1980)

- Clear dependency on thiols for activation
- Reacting the chromophore with methyl thioglycolate leads to a compound that appears to be the adduct with two extra hydrogens
- Hypothesized that a radical intermediate abstracts the 5' hydrogen on the sugar backbone, leading to DNA cleavage
- DNA cleavage is not aerobically dependent
- Napthoate functionality was hypothesized to intercalate into the DNA minor groove

## Myers' First Paper (1987)

PROPOSED STRUCTURE OF THE NEOCARZINOSTATIN CHROMOPHORE-METHYL THIOGLYCOLATE ADDUCT; A MECHANISM FOR THE NUCLEOPHILIC ACTIVATION OF NEOCARZINOSTATIN

Andrew G. Myers  
Contribution No. 7628 from the Arnold and Mabel Beckman Laboratory of Chemical Synthesis  
California Institute of Technology, Pasadena, California 91125

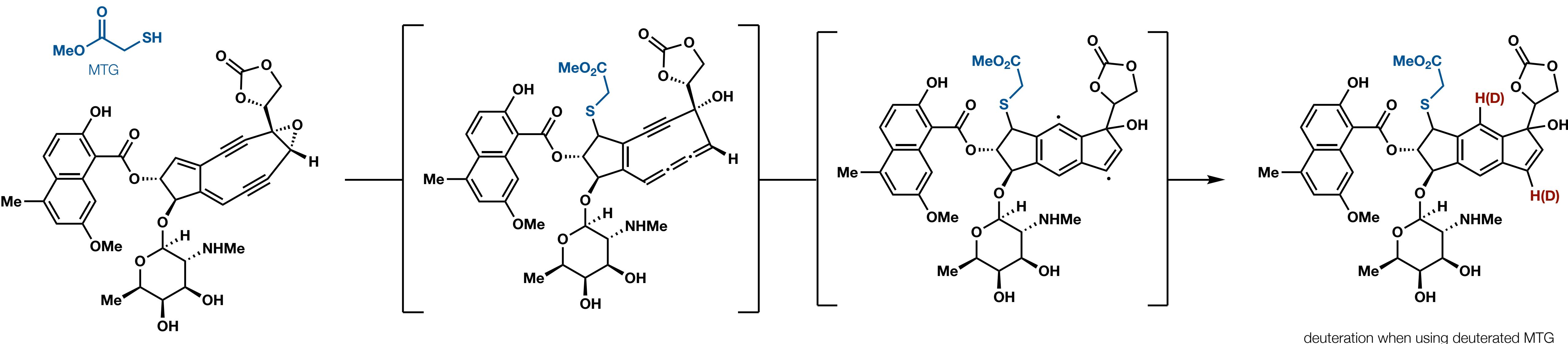
# NCS - Mechanism of Action

Myers' First Paper (1987)

PROPOSED STRUCTURE OF THE NEOCARZINOSTATIN CHROMOPHORE-METHYL THIOGLYCOLATE ADDUCT; A MECHANISM FOR THE NUCLEOPHILIC ACTIVATION OF NEOCARZINOSTATIN

Andrew G. Myers

Contribution No. 7628 from the Arnold and Mabel Beckman Laboratory of Chemical Synthesis  
California Institute of Technology, Pasadena, California 91125



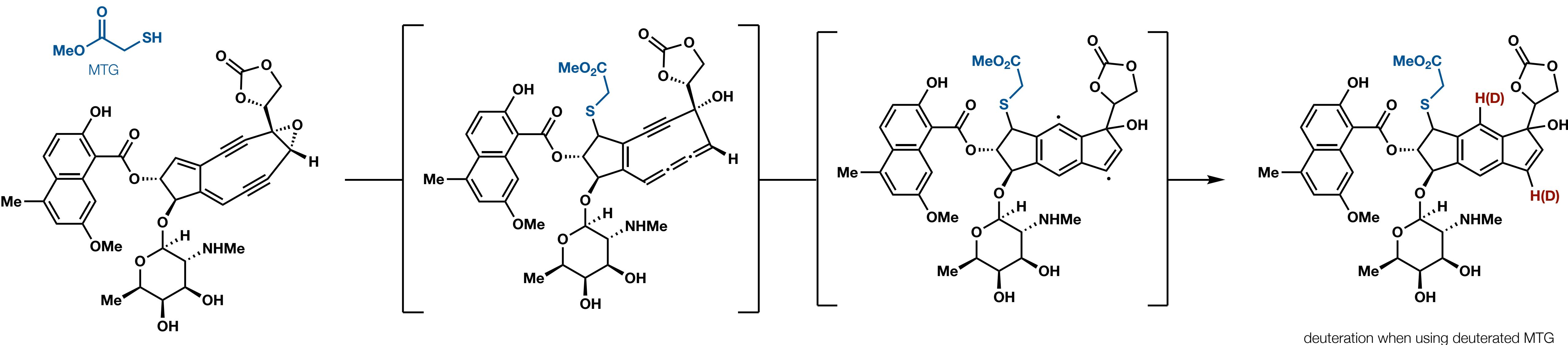
# NCS - Mechanism of Action

Myers' First Paper (1987)

PROPOSED STRUCTURE OF THE NEOCARZINOSTATIN CHROMOPHORE-METHYL THIOGLYCOLATE ADDUCT; A MECHANISM FOR THE NUCLEOPHILIC ACTIVATION OF NEOCARZINOSTATIN

Andrew G. Myers

Contribution No. 7628 from the Arnold and Mabel Beckman Laboratory of Chemical Synthesis  
California Institute of Technology, Pasadena, California 91125

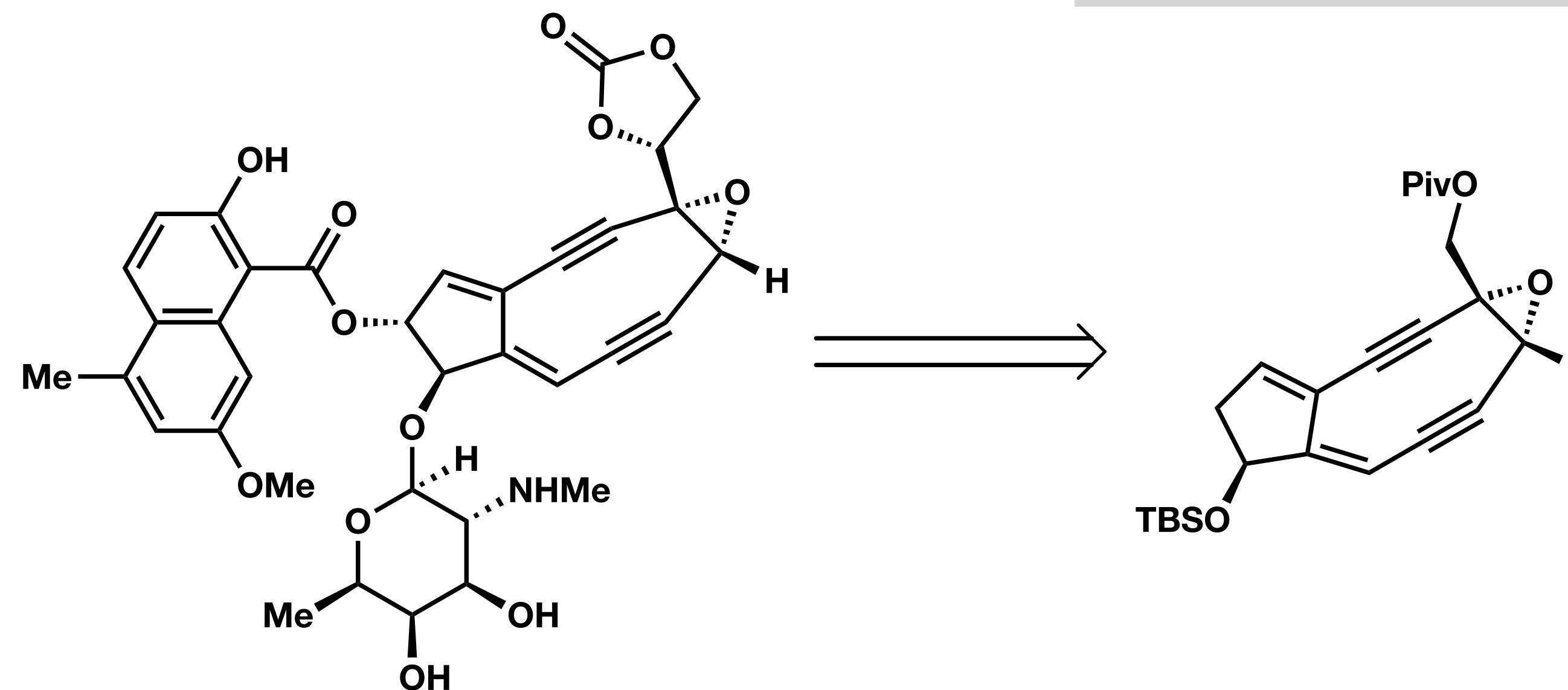


*Subsequent studies also confirmed the epoxide stereochemistry*

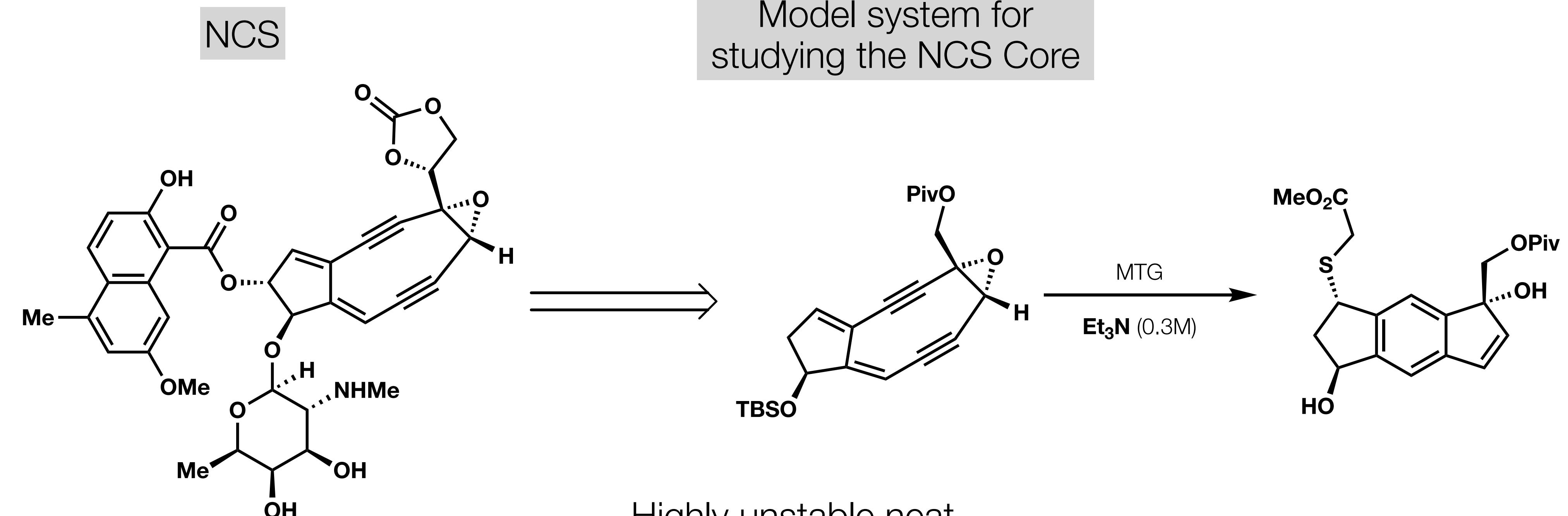
# NCS - Role of Aminoglycoside

NCS

Model system for  
studying the NCS Core

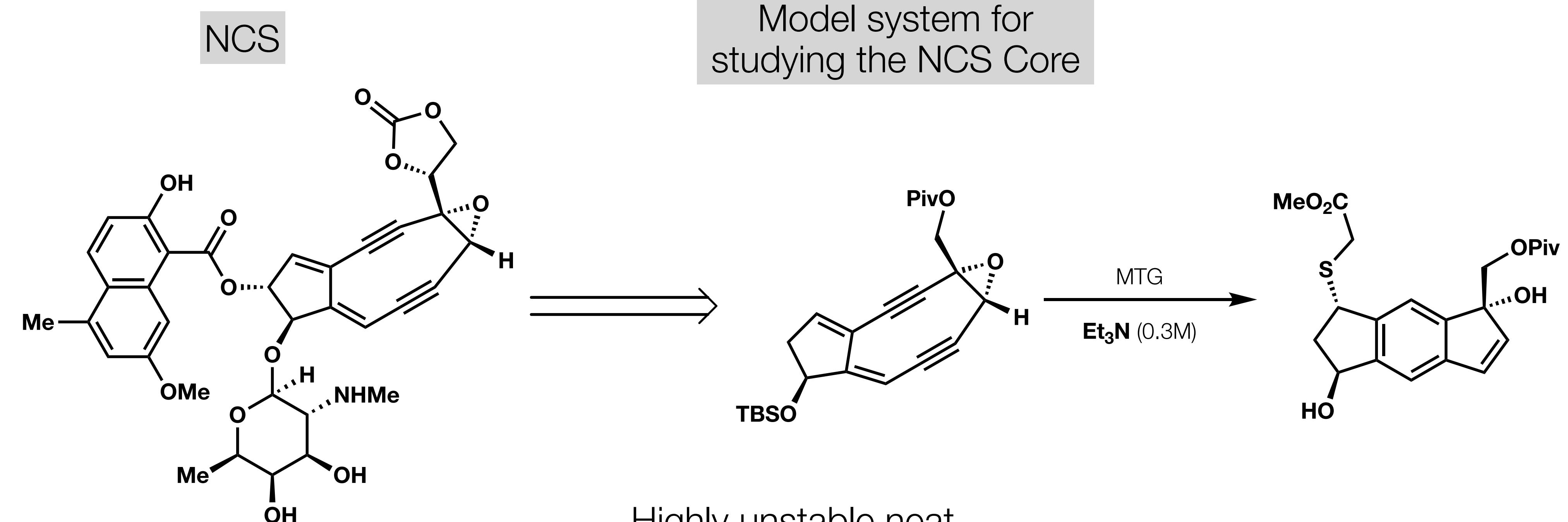


# NCS - Role of Aminoglycoside



- Highly unstable neat
- Reacts readily with MTG at -70°C
- But completely inert to MTG (up to 60 °C) in the absence of external base

# NCS - Role of Aminoglycoside

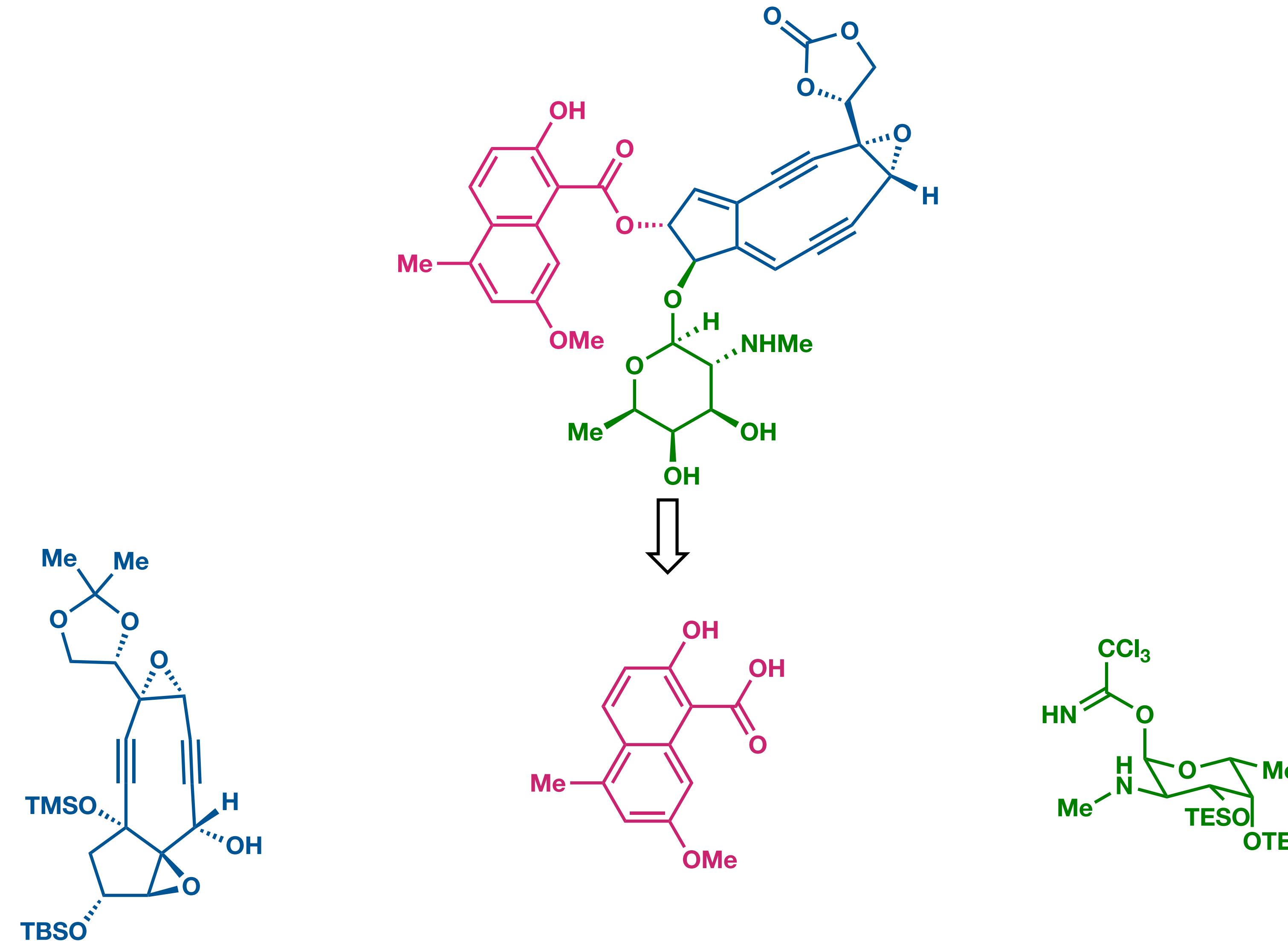


- Highly unstable neat
- Reacts readily with MTG at -70°C
- But completely inert to MTG (up to 60 °C) in the absence of external base

Strong evidence that the amino sugar serves as an internal base to facilitate thiol addition

Analogous roles of aminoglycosides observed in other natural products (i.e. calichemicin)

# Synthesis of NCS

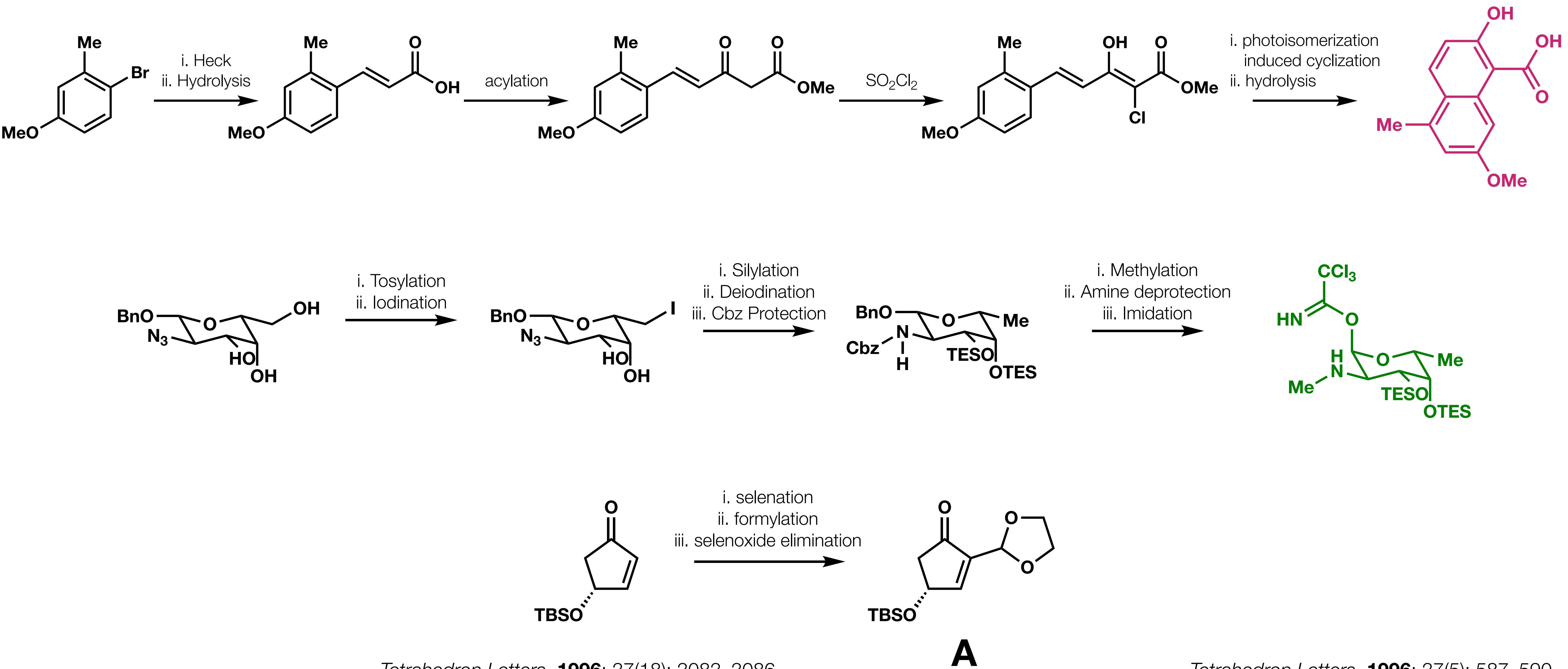


JACS **1991**; 113(2): 694–695.

JACS **1996**: 118(41): 10006–10007.

Tetrahedron Letters. **1996**; 37(5): 587–590. JACS **1998**; 120(21): 5319–5320.

# NCS - Fragment Syntheses

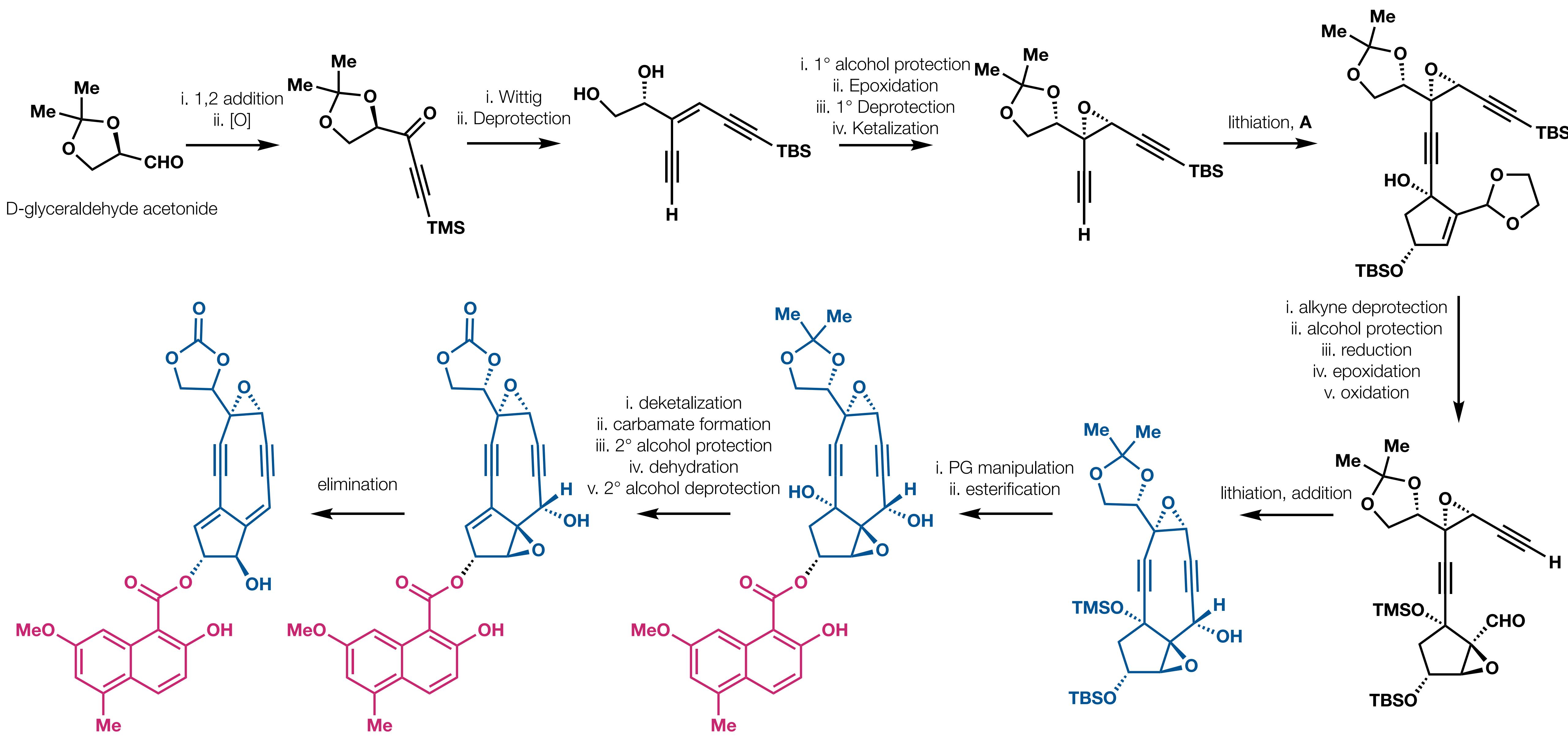


Tetrahedron Letters. **1996**; 37(18): 3083–3086.

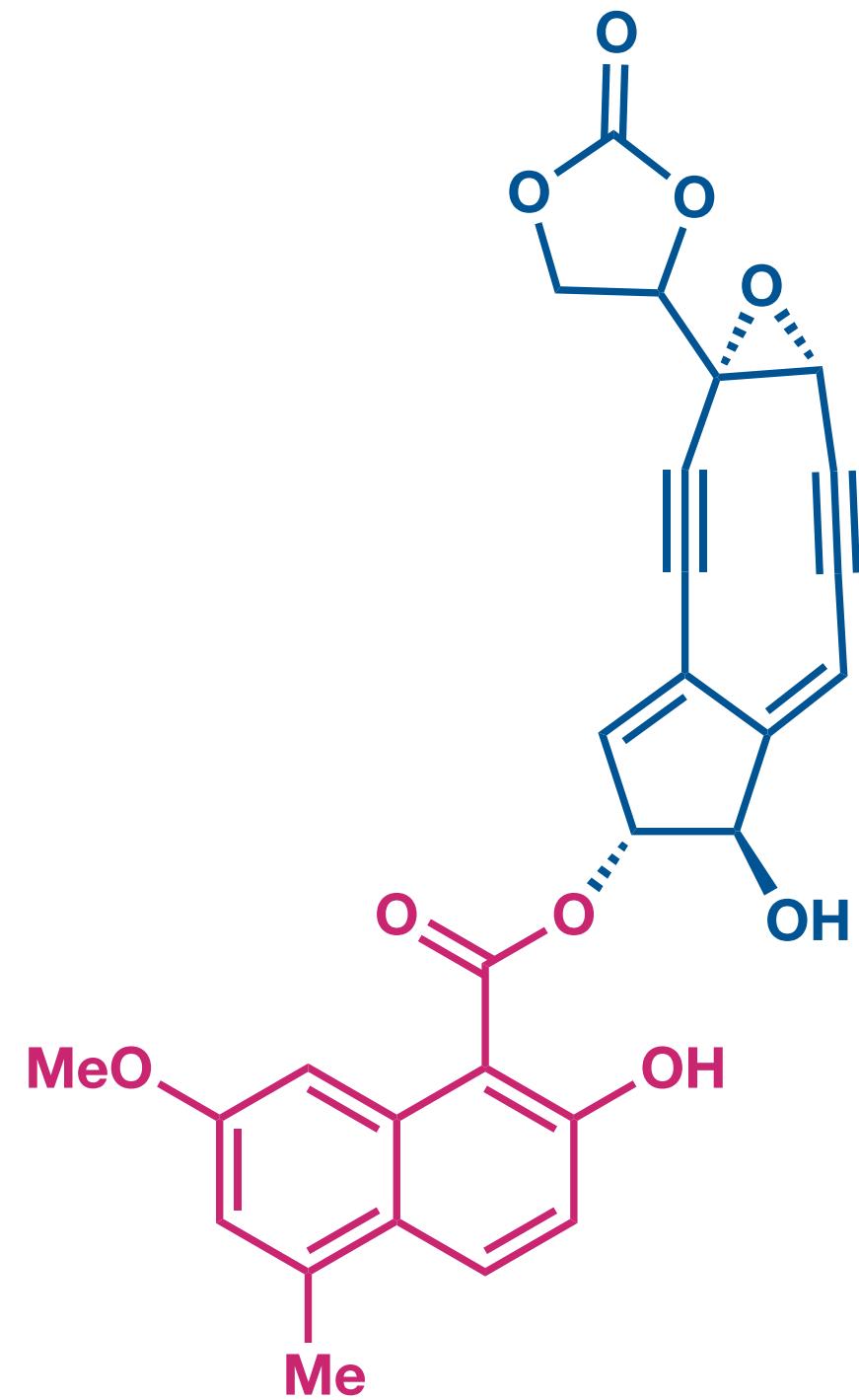
Tetrahedron Letters. **1996**; 37(5): 587–590.

JACS **1998**; 120(21): 5319–5320.

# Synthesis of NCS-chrom Aglycon



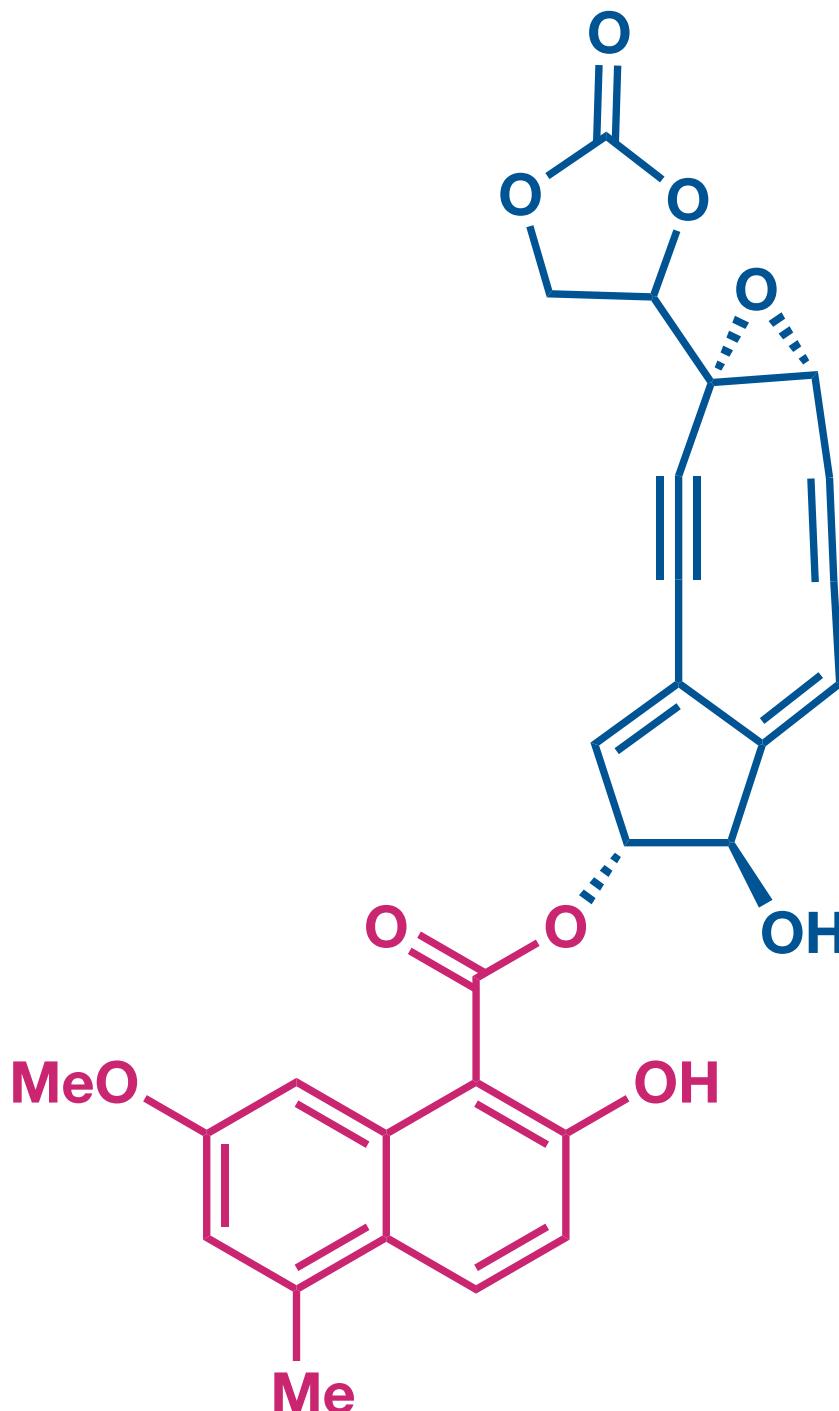
# NCS-chrom Aglycon



NCS-chrom aglycon

- First synthesis
- Highly Unstable
- Fresh batches were prepared and purified before biological studies

# NCS-chrom Aglycon



NCS-chrom aglycon

- First synthesis
- Highly Unstable
- Fresh batches were prepared and purified before biological studies

## Comparison of aglycon to NCS-chrom in DNA Cleavage Assays

Key takeaways:

A glycon does not react with thiol nucleophiles in the absence of base and DNA (consistent with model substrate studies)

When incubated with DNA and apo-NCS, 10x the concentration of the aglycon was needed to promote cleavage

MTG induced cleavage reactions with NCS-chrom are less efficient in the absence of apo-NCS

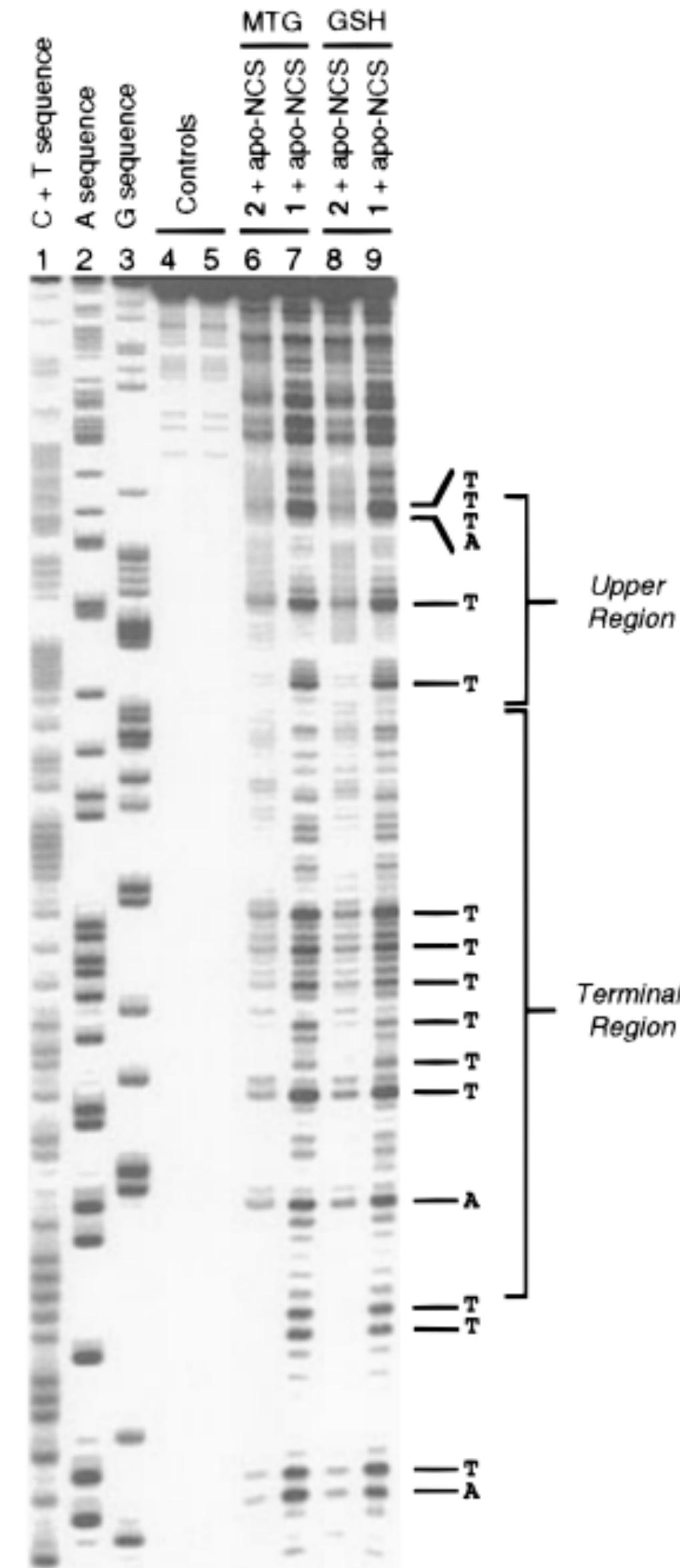
Supporting evidence that apo-NCS still binds to the aglycon

Sequence specificity of cleavage is the same (mainly AT pairs)

Kinetic studies reveal the RDS is likely thiol addition, which occurs as a ternary complex of NCS, thiol nucleophile, and DNA

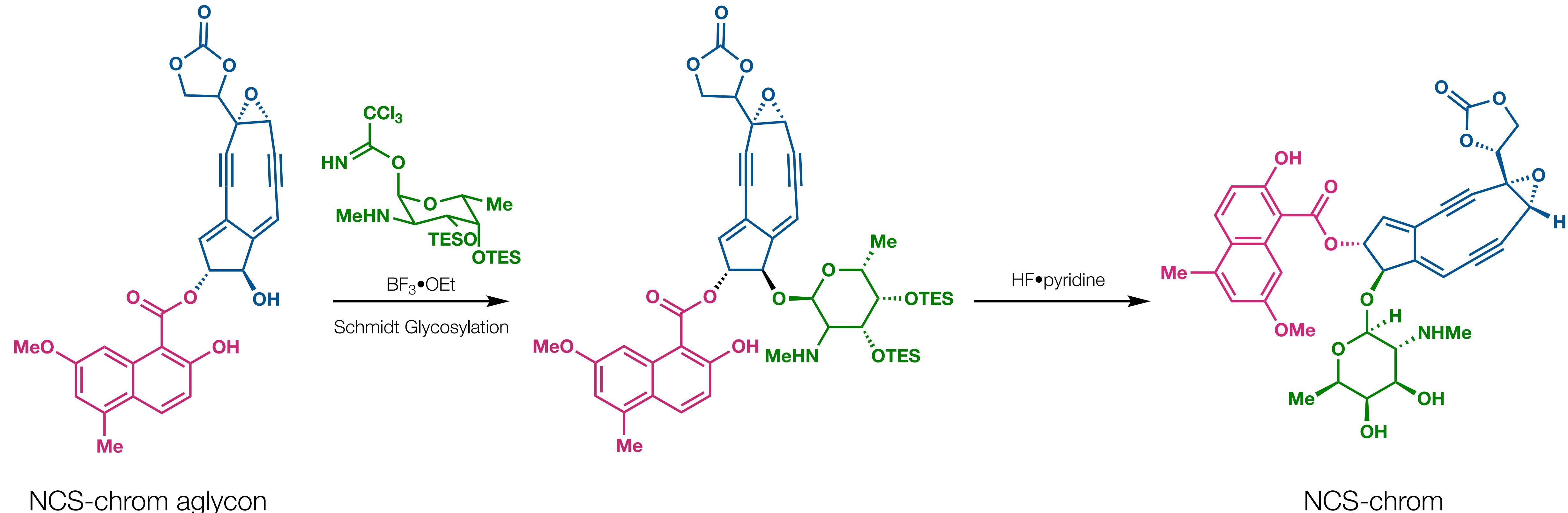
## Role of sugar is hypothesized to

- Stabilize NCS-chrom (as a steric group to slow down the approach of free radicals)
- Not be responsible for DNA cleavage
- Serve as an internal base to catalyze thiol attack



# Total Synthesis of NCS-chrom

Completion of NCS total synthesis: glycosylation

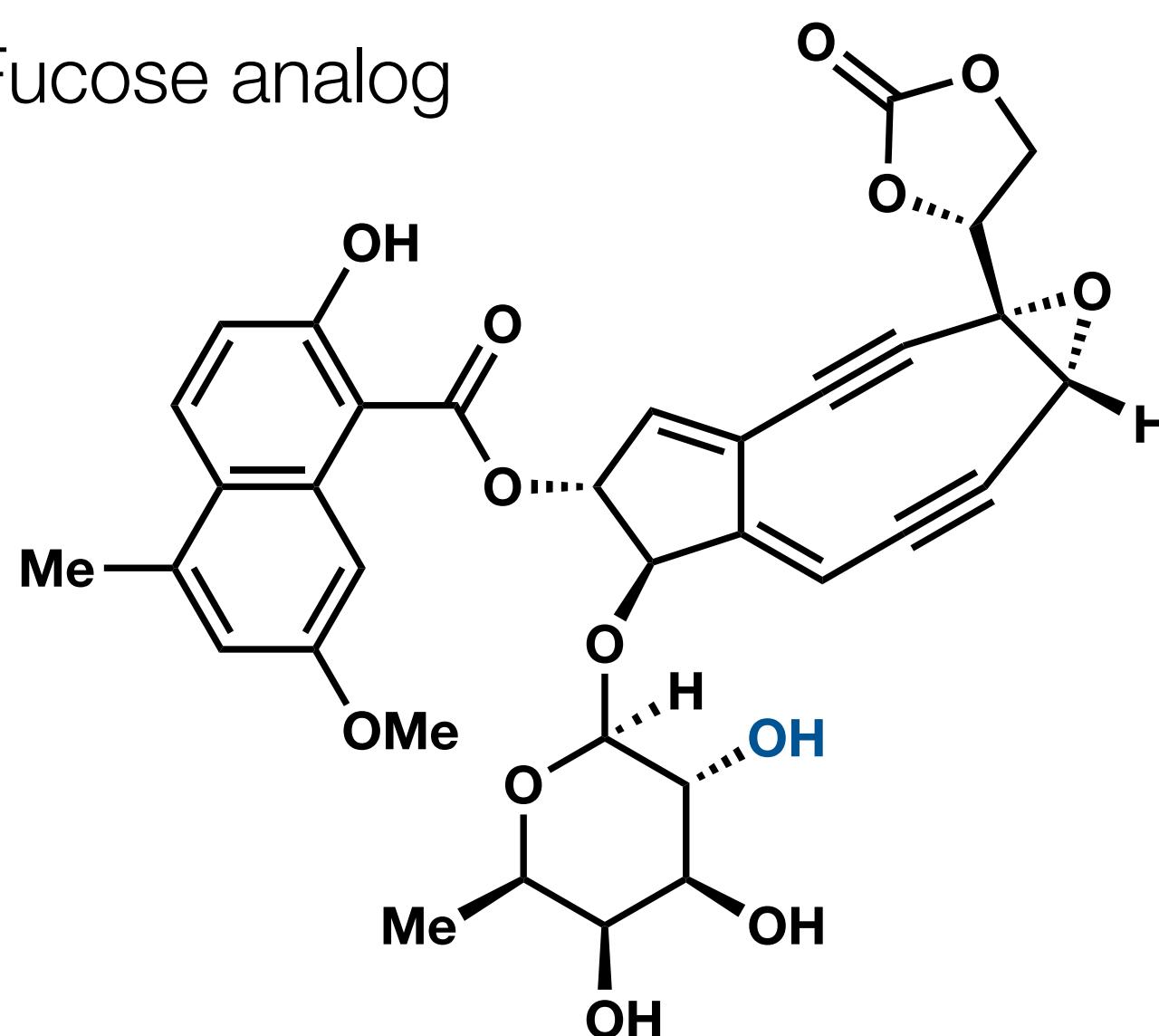


NCS-chrom aglycon

NCS-chrom

# Comparison to Fucose Analog

Fucose analog



NCS-chrom

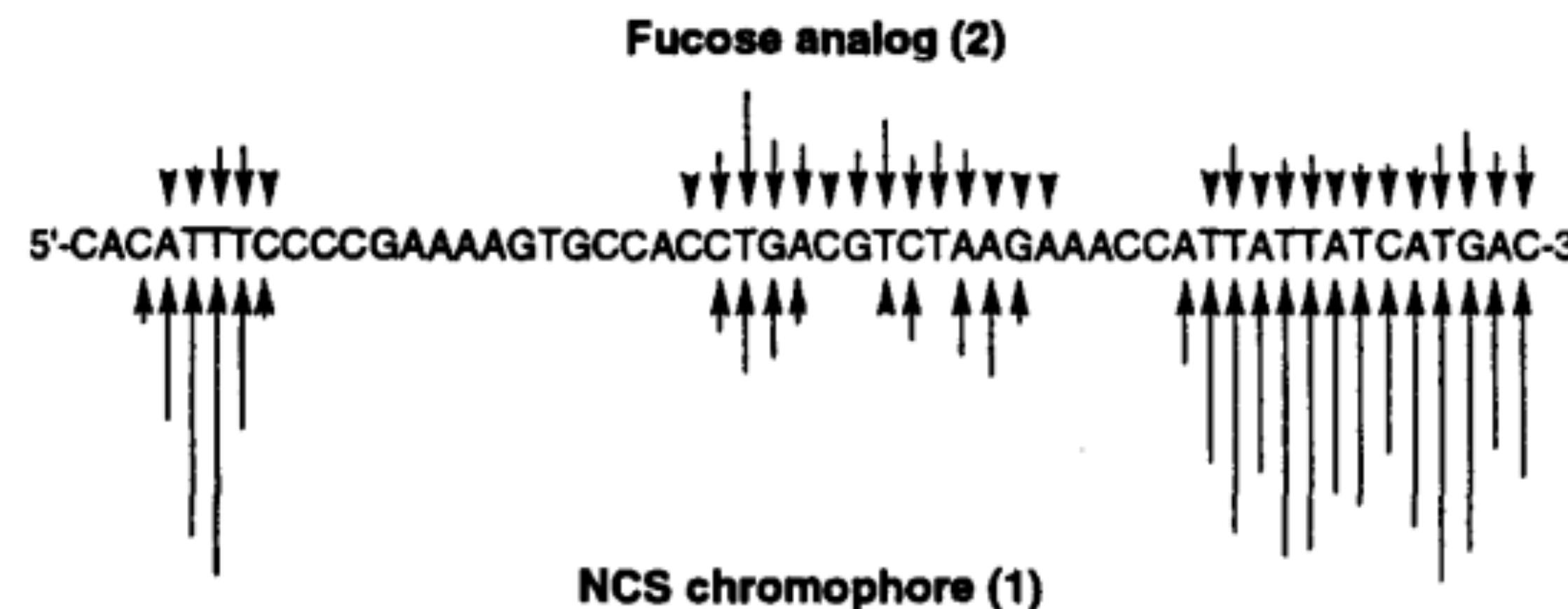
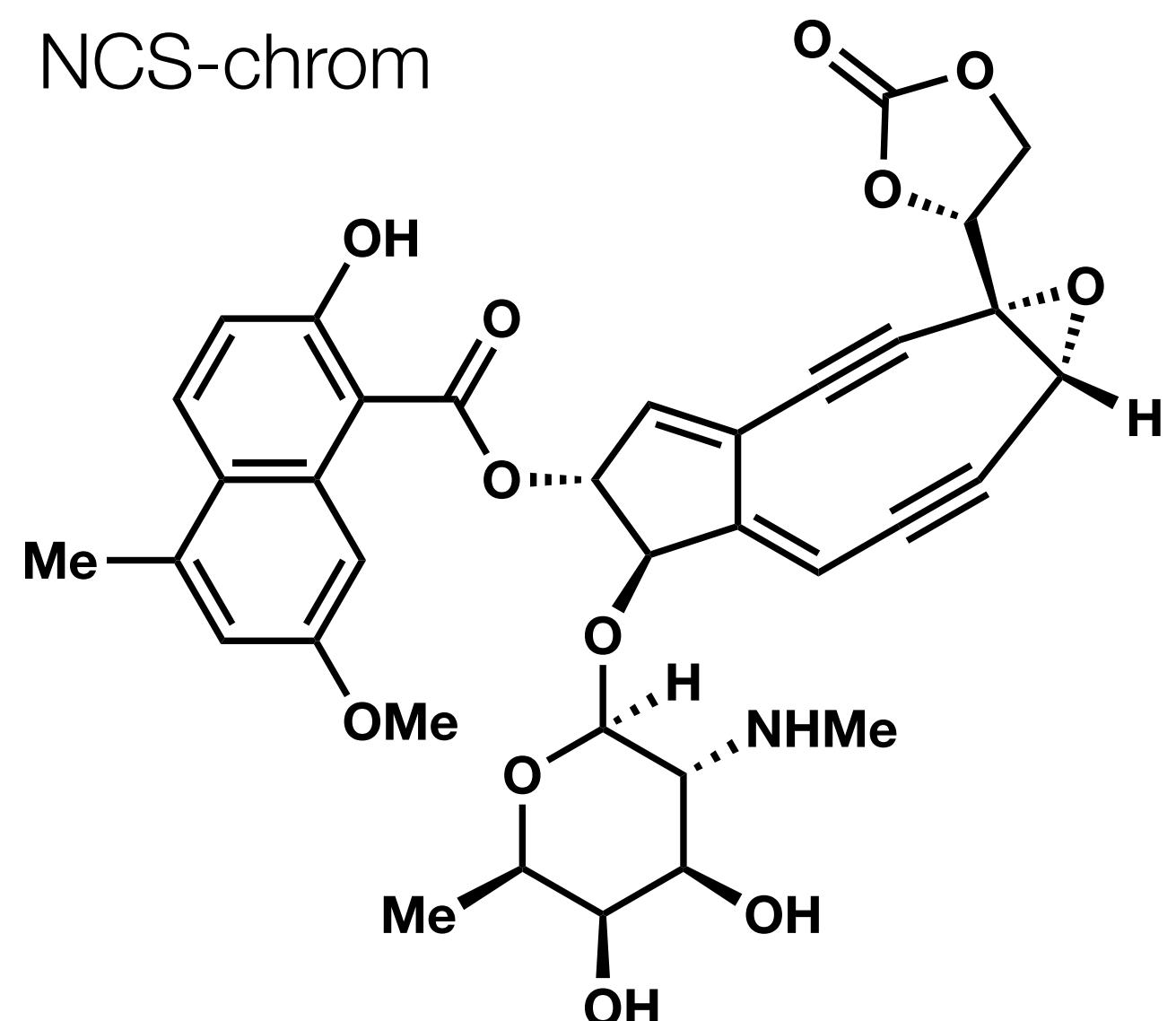
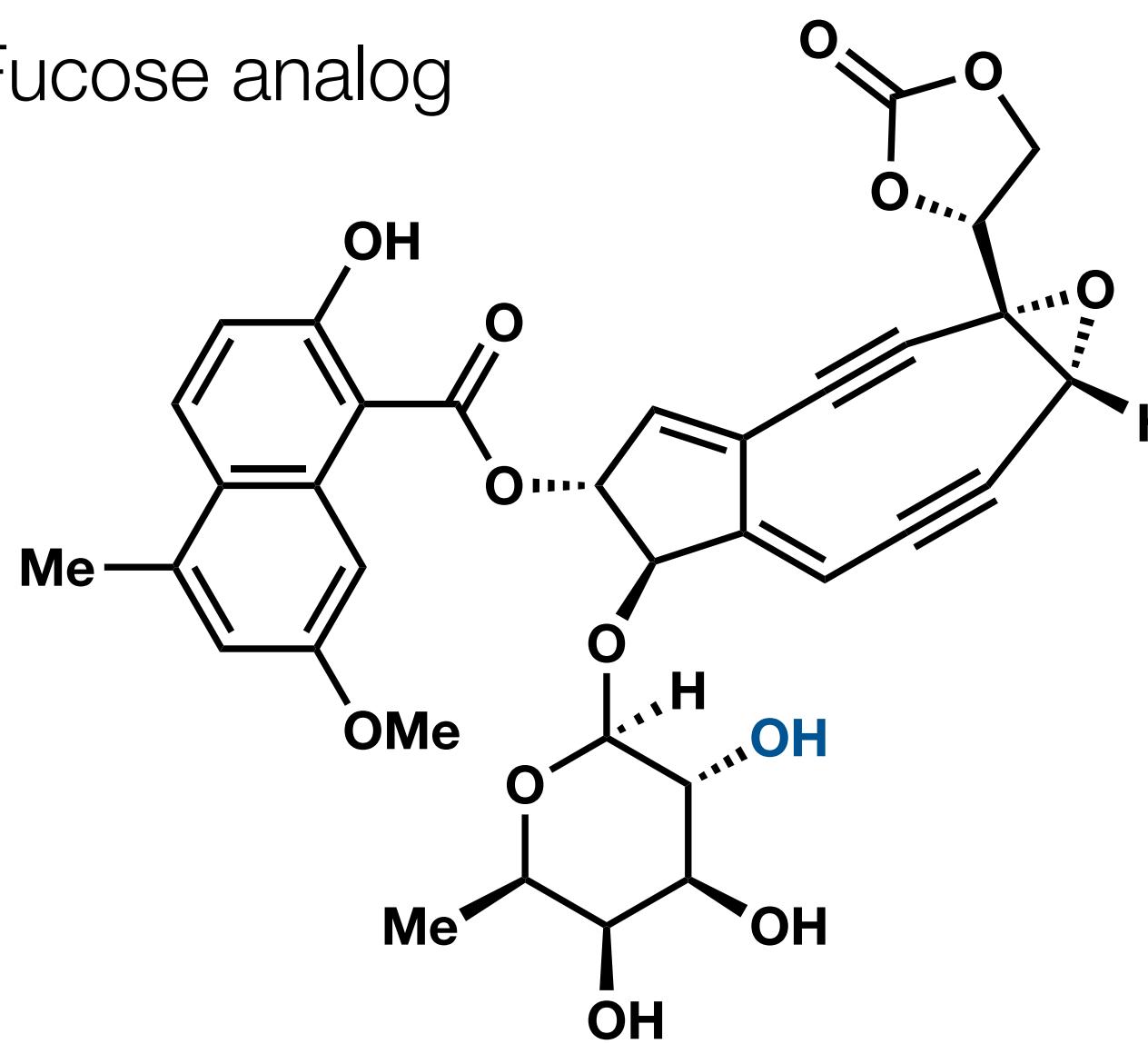


Figure 1. Histogram of DNA cleavage, upper: **2** (0.05 mM), calf thymus DNA (1 mM), 3'-<sup>32</sup>P labeled DNA (see text, 50 kcpm), NaCl (20 mM), Tris-HCl (50 mM, final pH 7.5), MTG (3 mM), at 2°C; lower: **1** at 2.5-fold lower concentration (0.02 mM). DNA cleavage is normalized to the concentration of cleaving agent. After 30 min, each reaction was quenched; DNA cleavage products were assayed by denaturing 8% polyacrylamide gel electrophoresis using storage phosphor autoradiography for quantitative analysis

# Comparison to Fucose Analog

Fucose analog



Fucose analog (2)

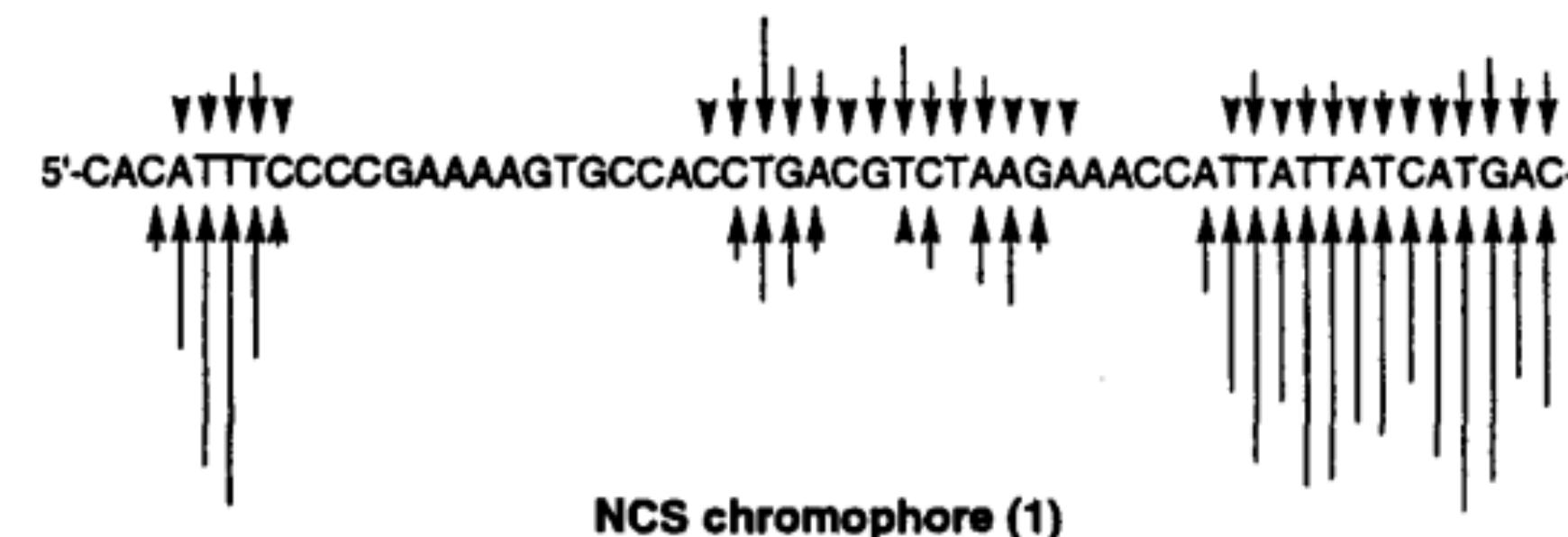
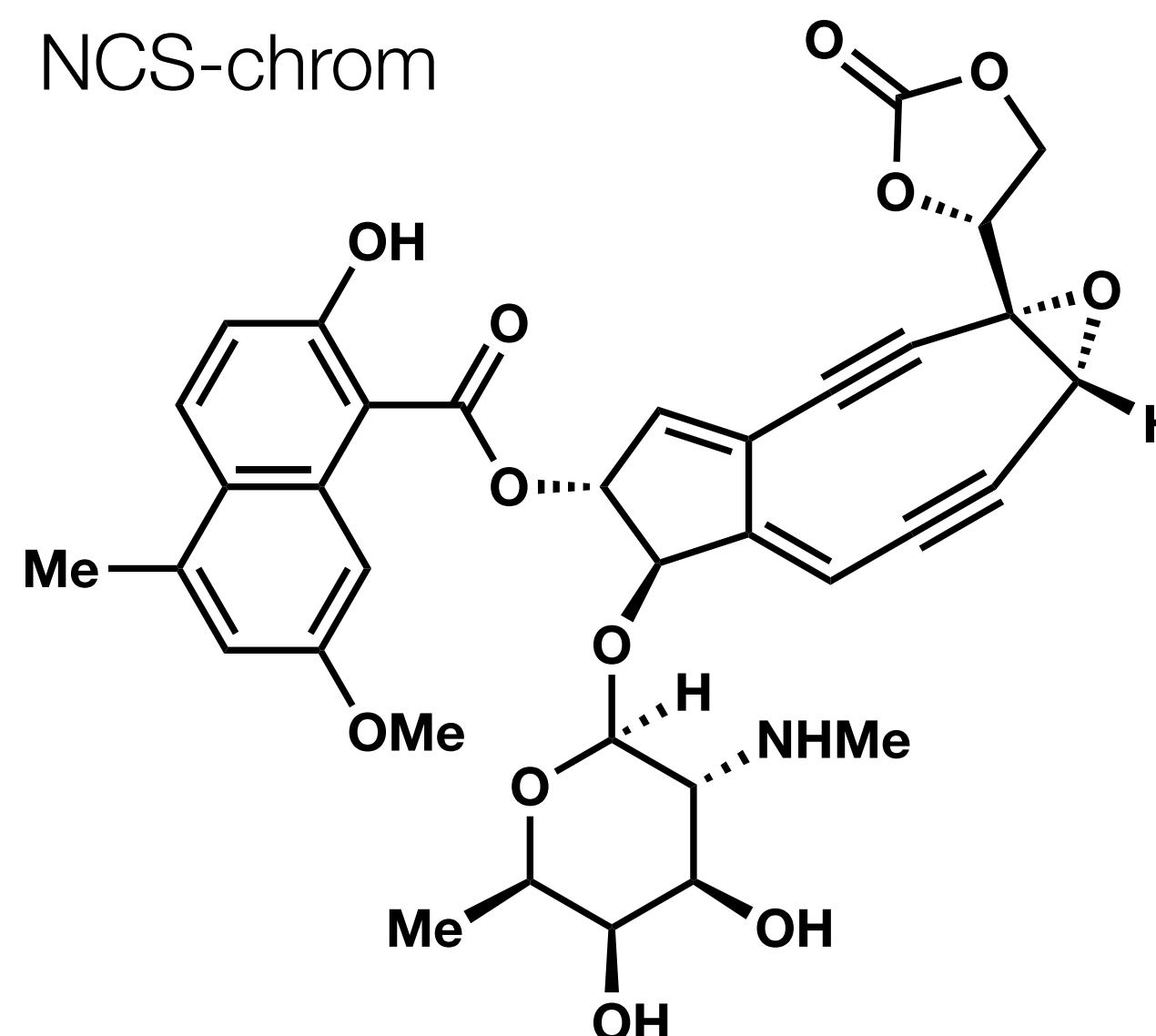


Figure 1. Histogram of DNA cleavage, upper: **2** (0.05 mM), calf thymus DNA (1 mM, 3'-<sup>32</sup>P labeled DNA (see text, 50 kcpm), NaCl (20 mM), Tris-HCl (50 mM, final pH 7.5), MTG (3 mM), at 2°C; lower: **1** at 2.5-fold lower concentration (0.02 mM). DNA cleavage is normalized to the concentration of cleaving agent. After 30 min, each reaction was quenched; DNA cleavage products were assayed by denaturing 8% polyacrylamide gel electrophoresis using storage phosphor autoradiography for quantitative analysis

NCS-chrom



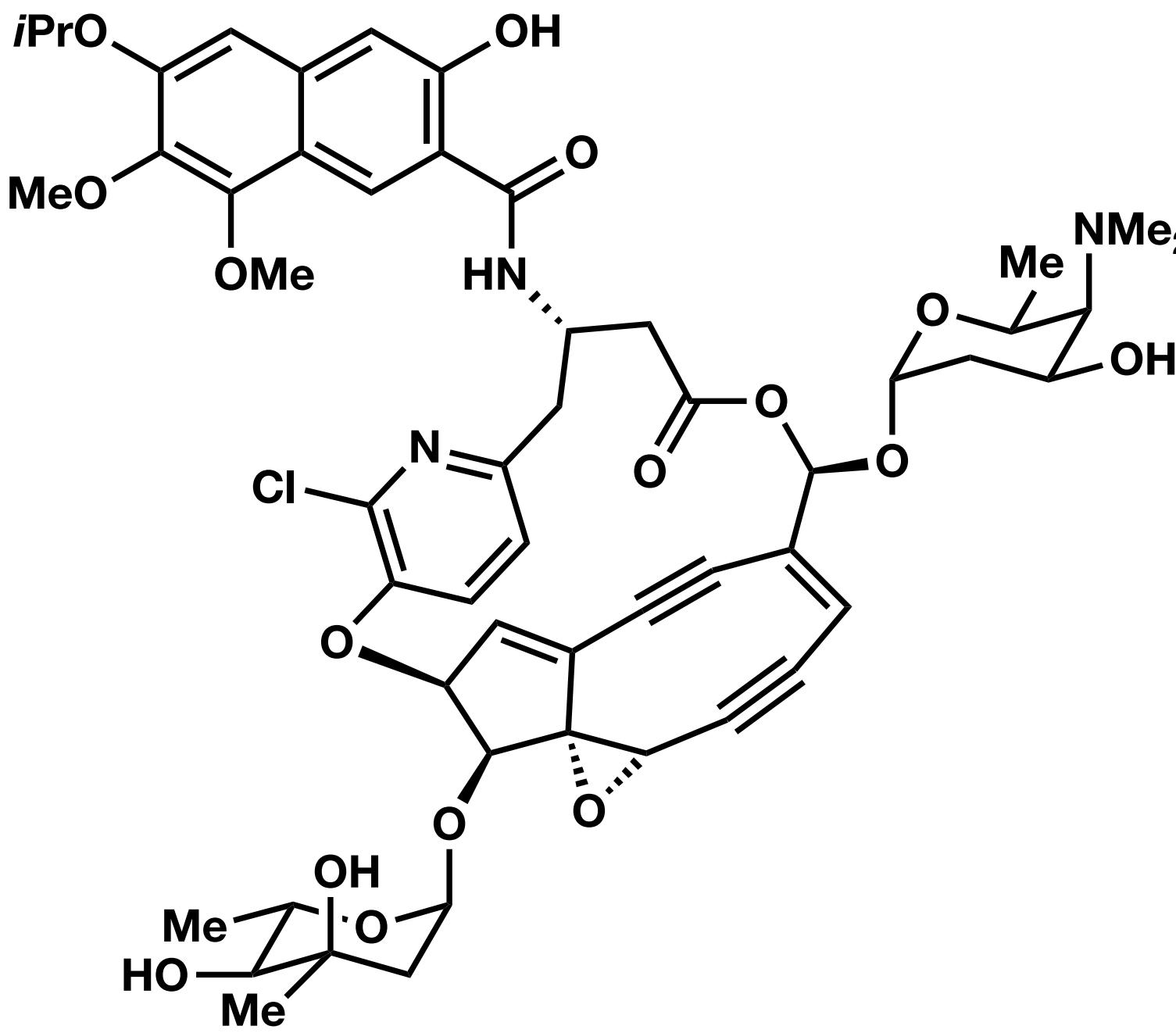
## Key Takeaways

Kinetic profiles for DNA cleavage showed that the fructose analog is 3.3 (with MTG) or 4.6 (with GSH) fold slower with excess thiol

The two compounds have similar patterns of DNA cleavage

Pre-incubating the fucose analog with apo-NCS protects it from thiol activation, likely a consequence of tight binding

# Kedarcidin

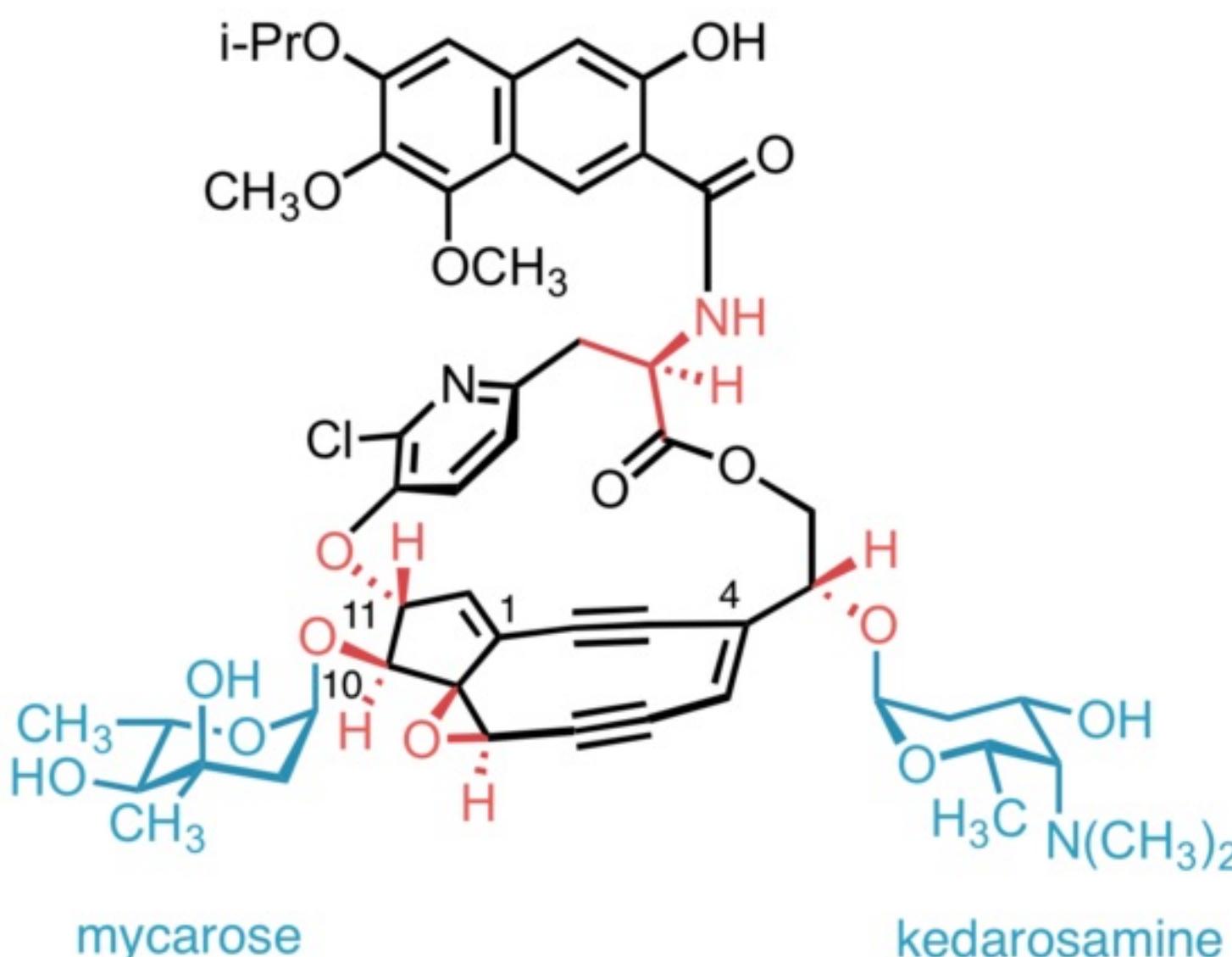


First discovered by BMS in 1992 through fermentation of an Actinomycete strain

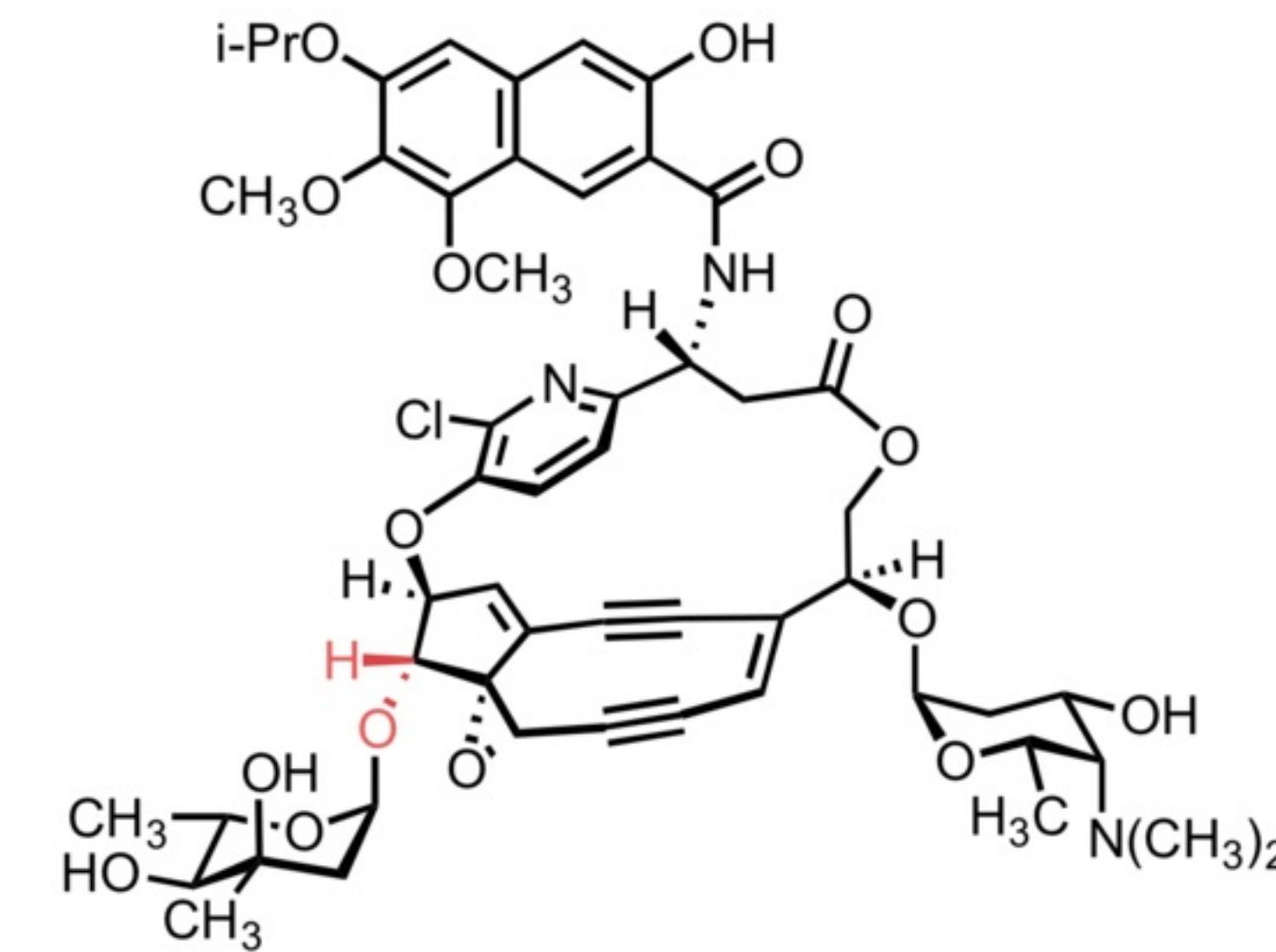
Like NCS, is composed of a enediyne chromophore and an apoprotein for stabilization

# Kedarcidin

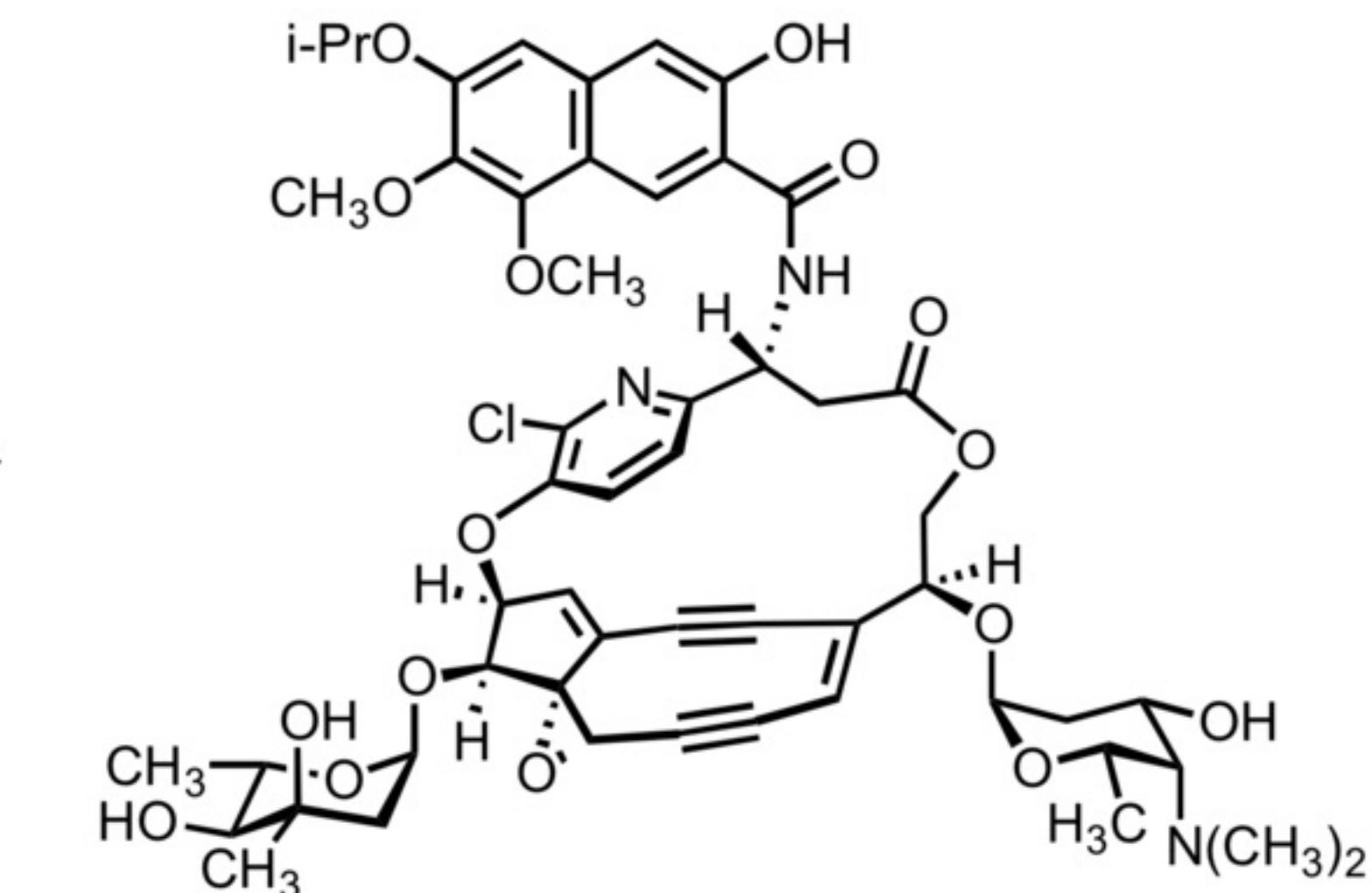
## Structural Revisions Throughout the Years



Originally proposed structure, 1993  
Leet, J. E. et al.

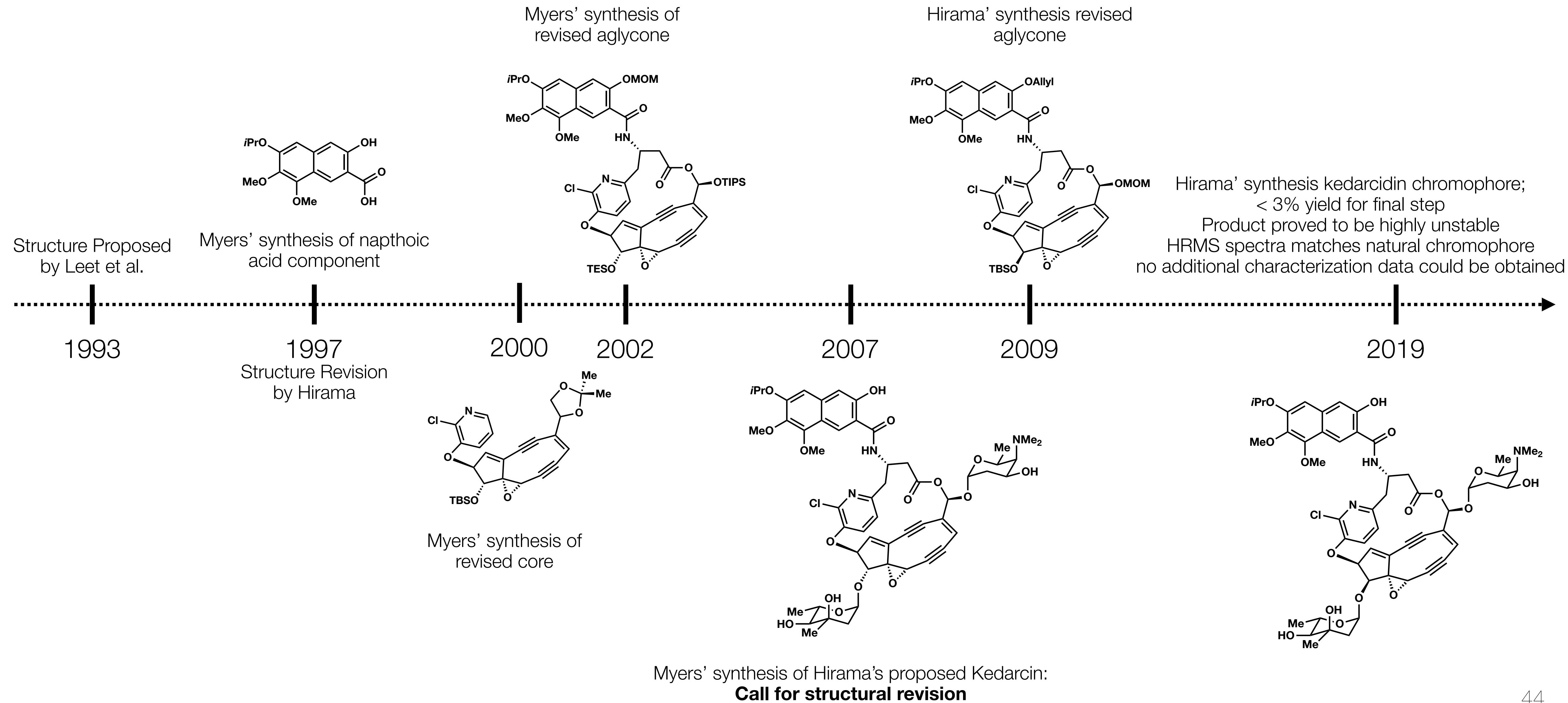


Revised structure, 1997  
Hirama, M. et al.

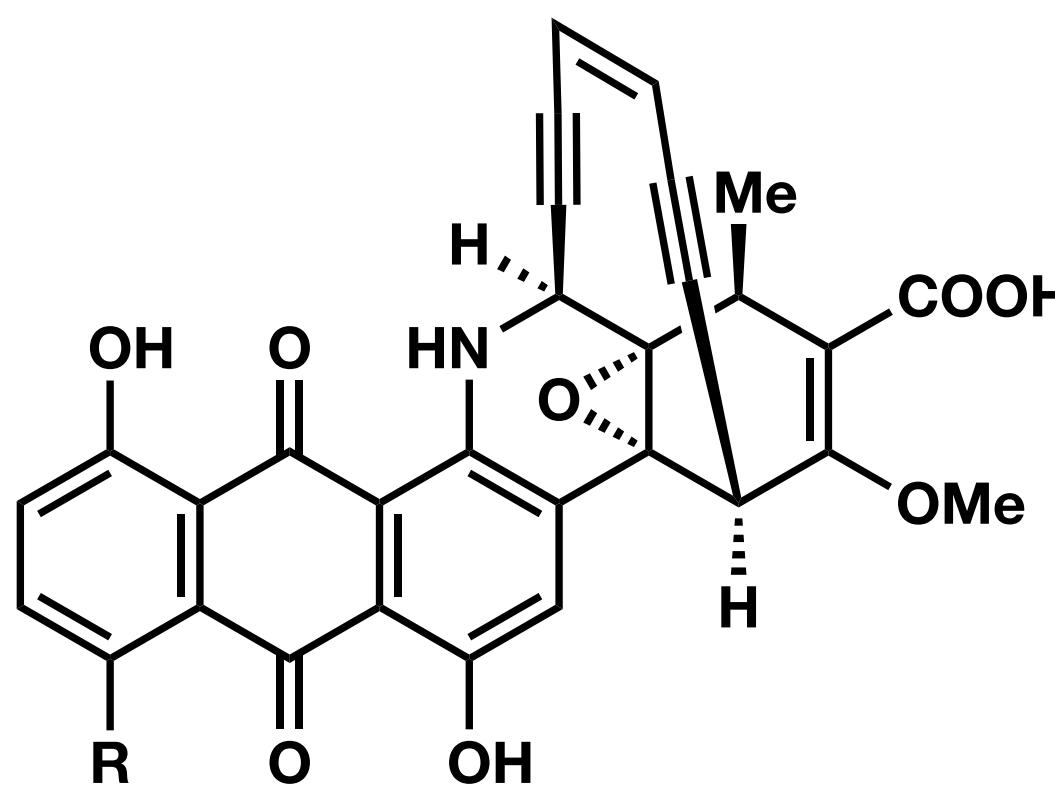


Kedarcidin chromophore  
(Revised structure, 2007)

# Kedarcidin on a Timeline



# Dynemicin A

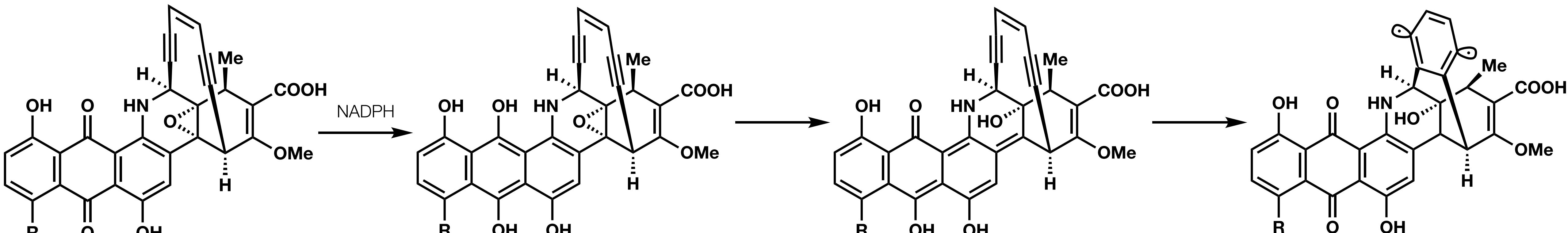


**R = OH: dynemicin A**

**R = H: deoxydynemicin A**

Isolated from *Micromonospora chersina*

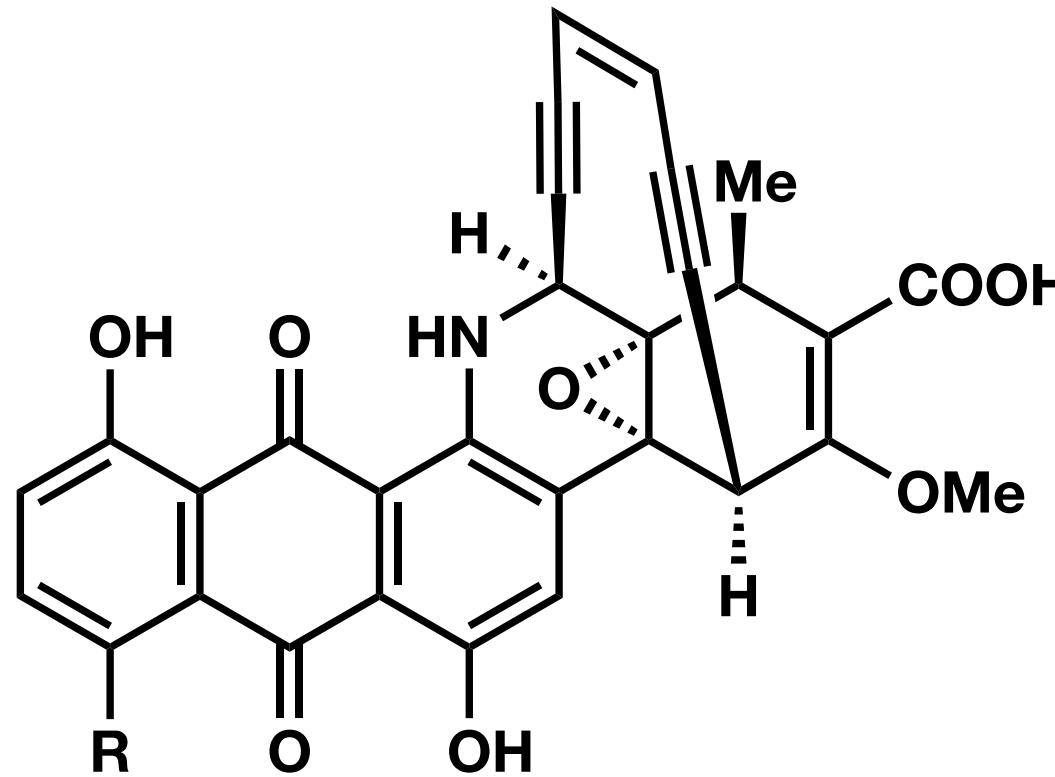
# Dynemicin A



R = OH: dynemicin A

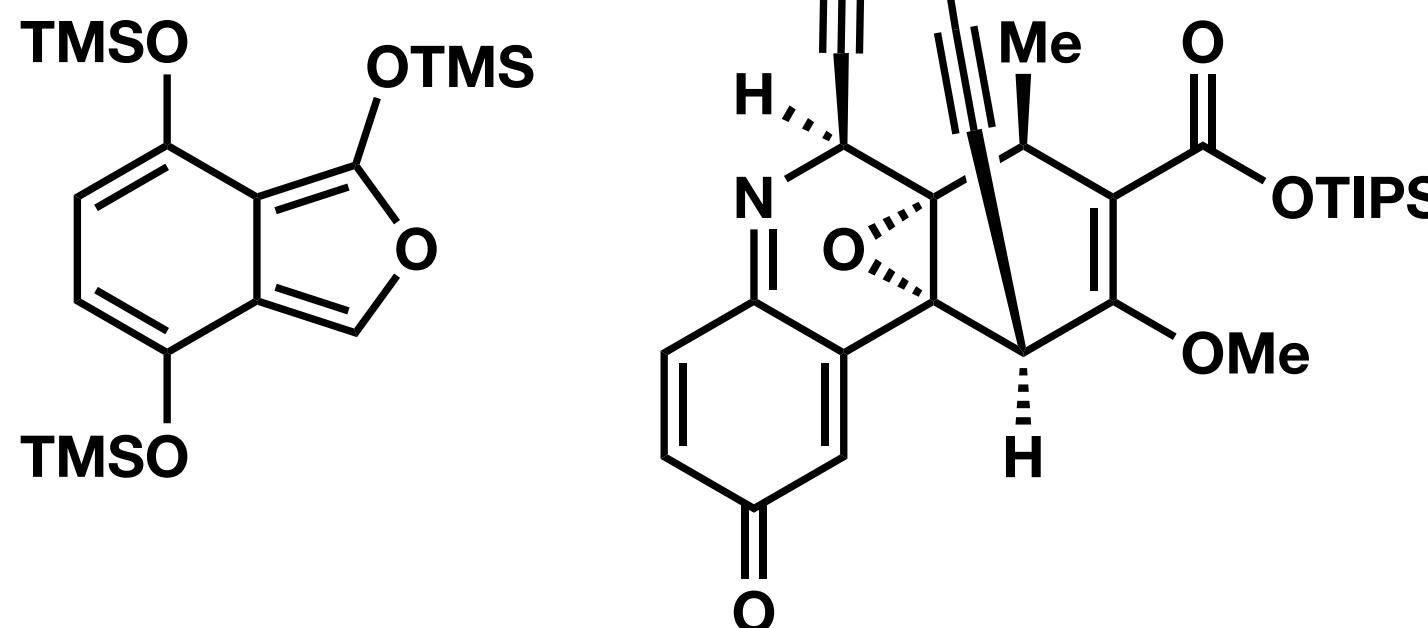
R = H: deoxydynemicin A

# Dynemicin A

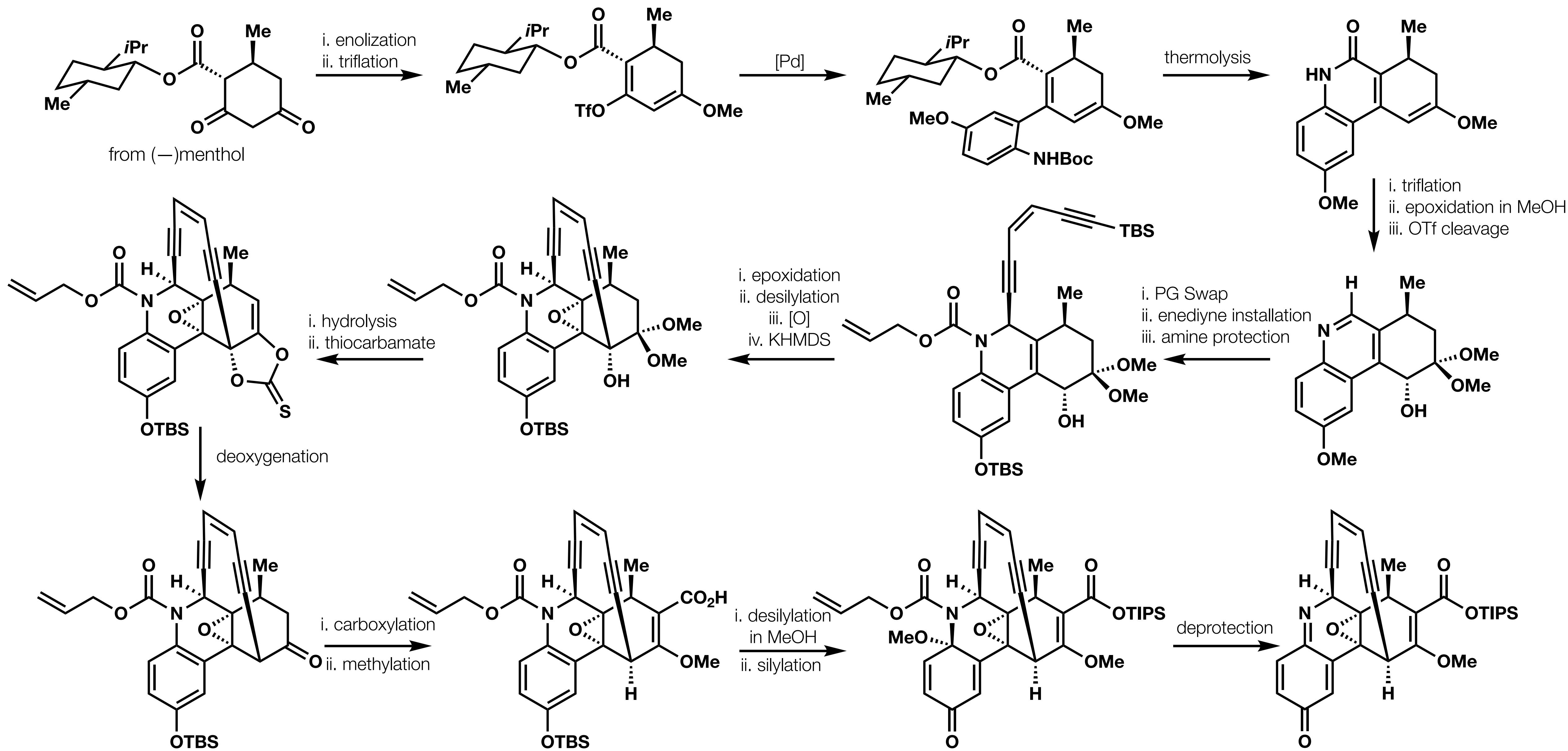


R = OH: dynemicin A

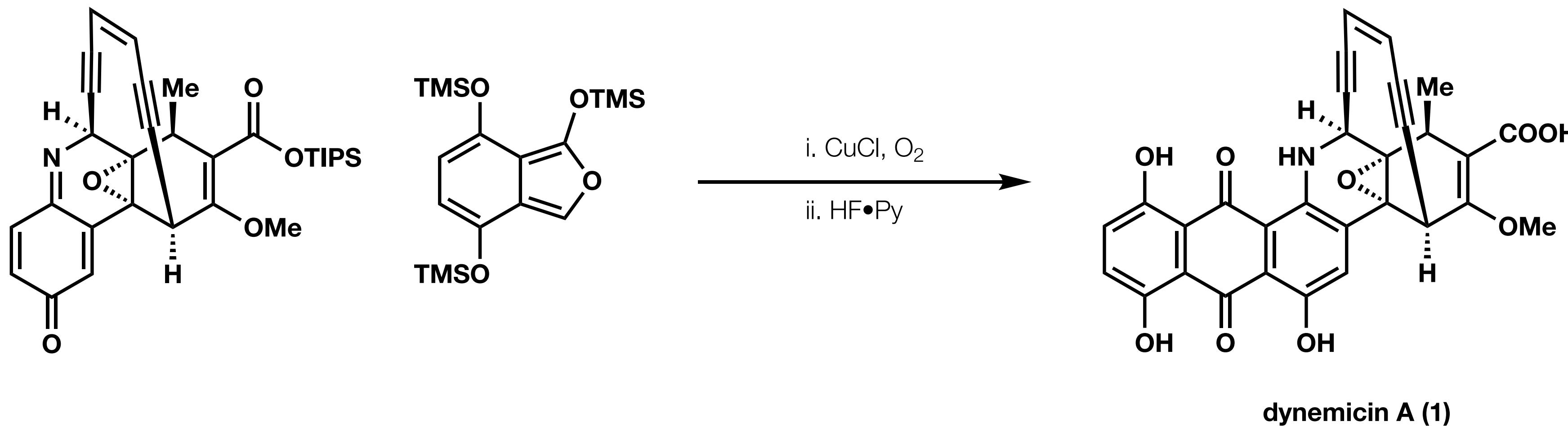
R = H: deoxydynemicin A



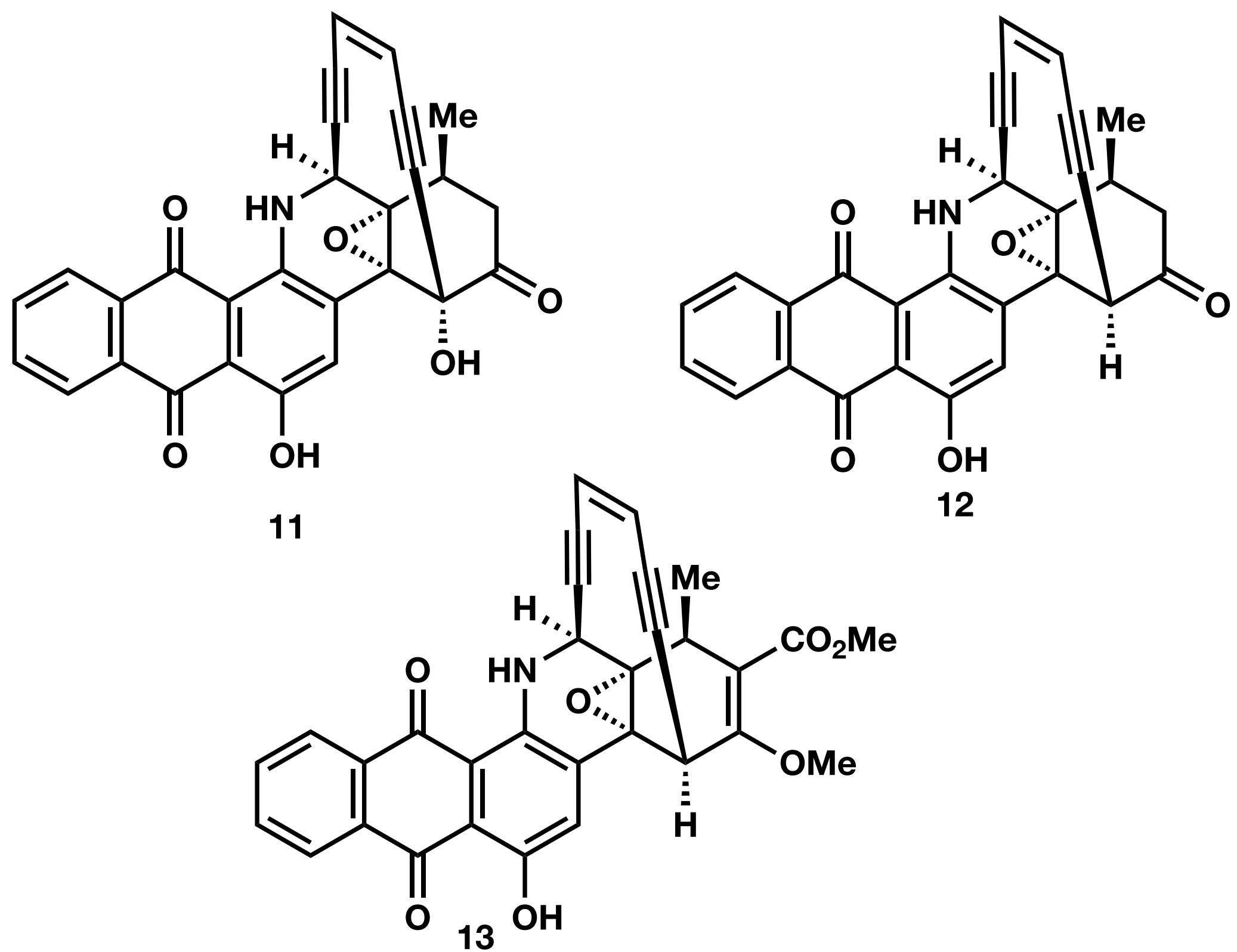
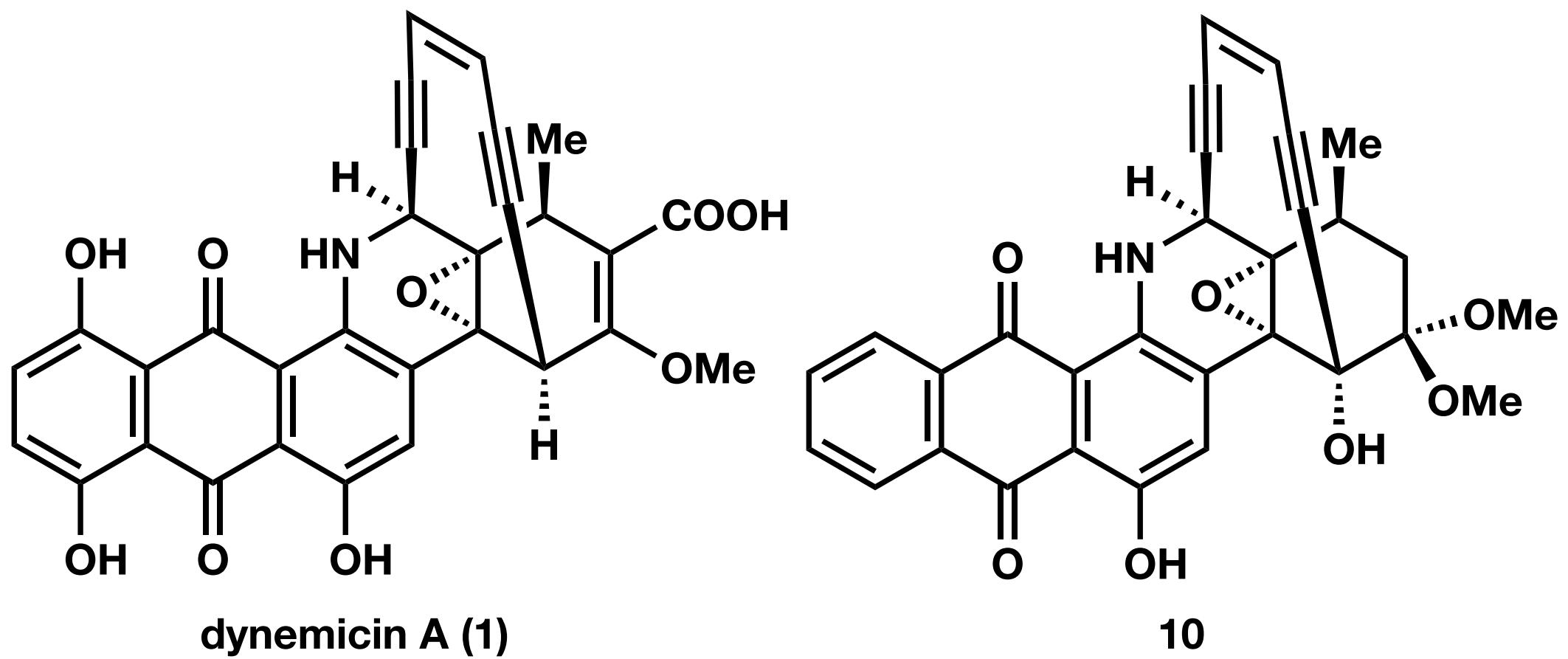
# Dynemicin A



# Dynemicin A



# Dynemicin A



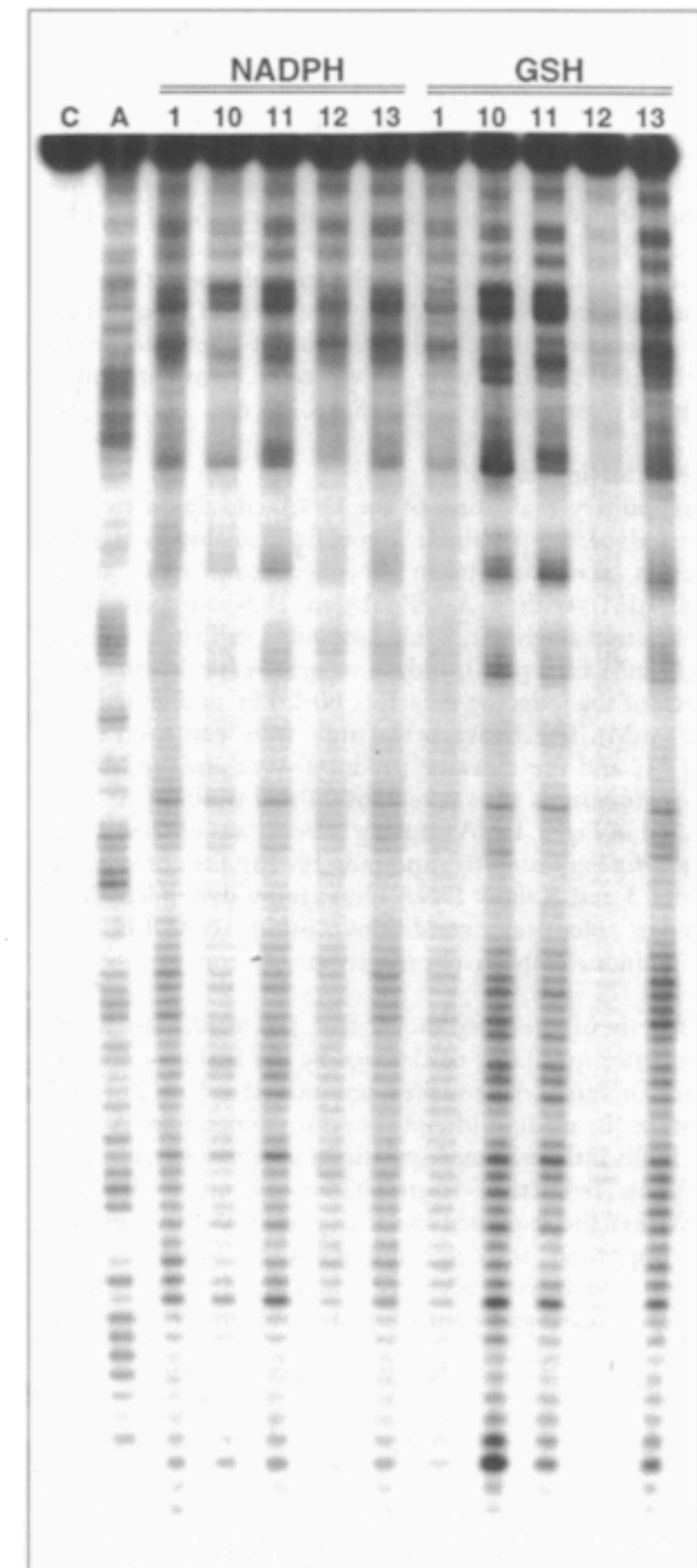
Comparison of dynemicin to synthetic  
analogs  
in DNA Cleavage Assays

## Key Takeaways

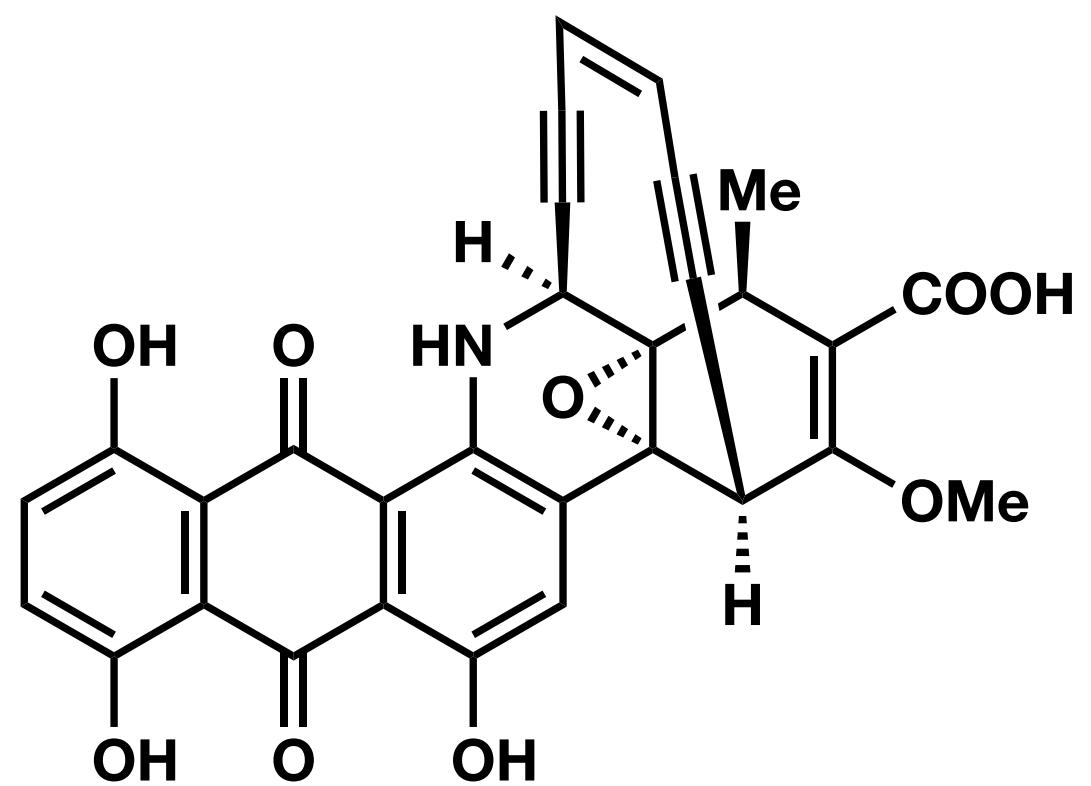
Little sequence specificity for cleavage  
observed

Different modes of activation result in  
differences in cleavage efficiencies  
between analogs

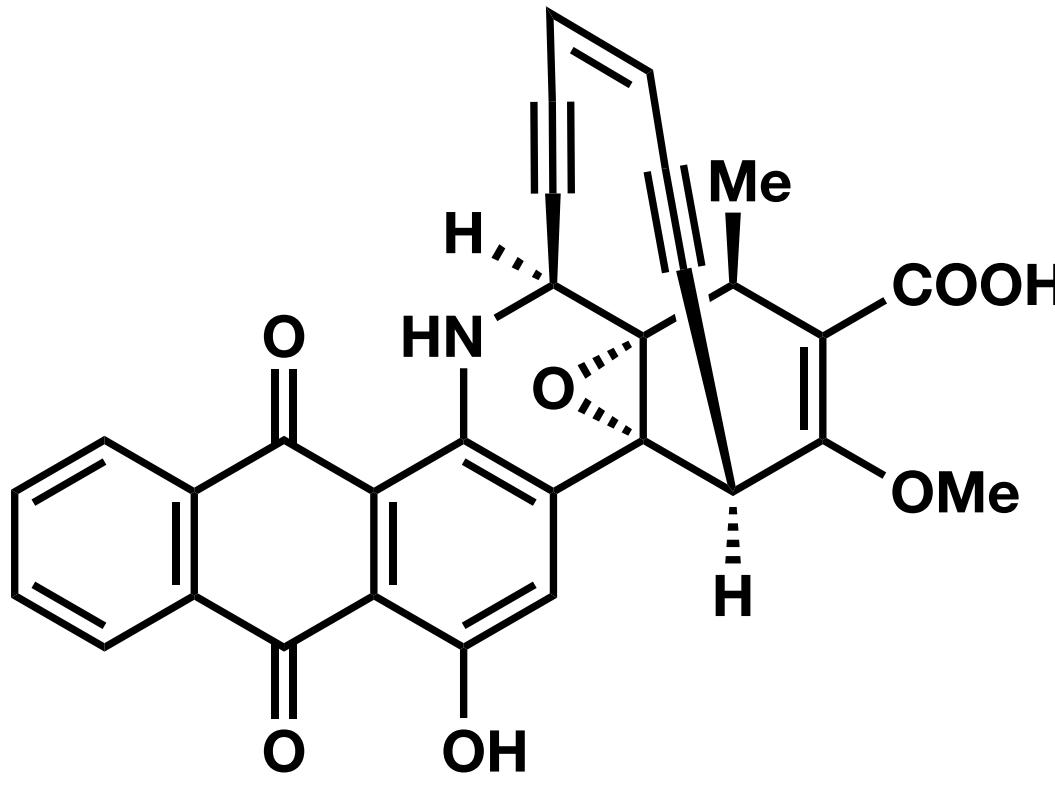
Mechanistically, 10 behaves as a  
perfect analog of 1



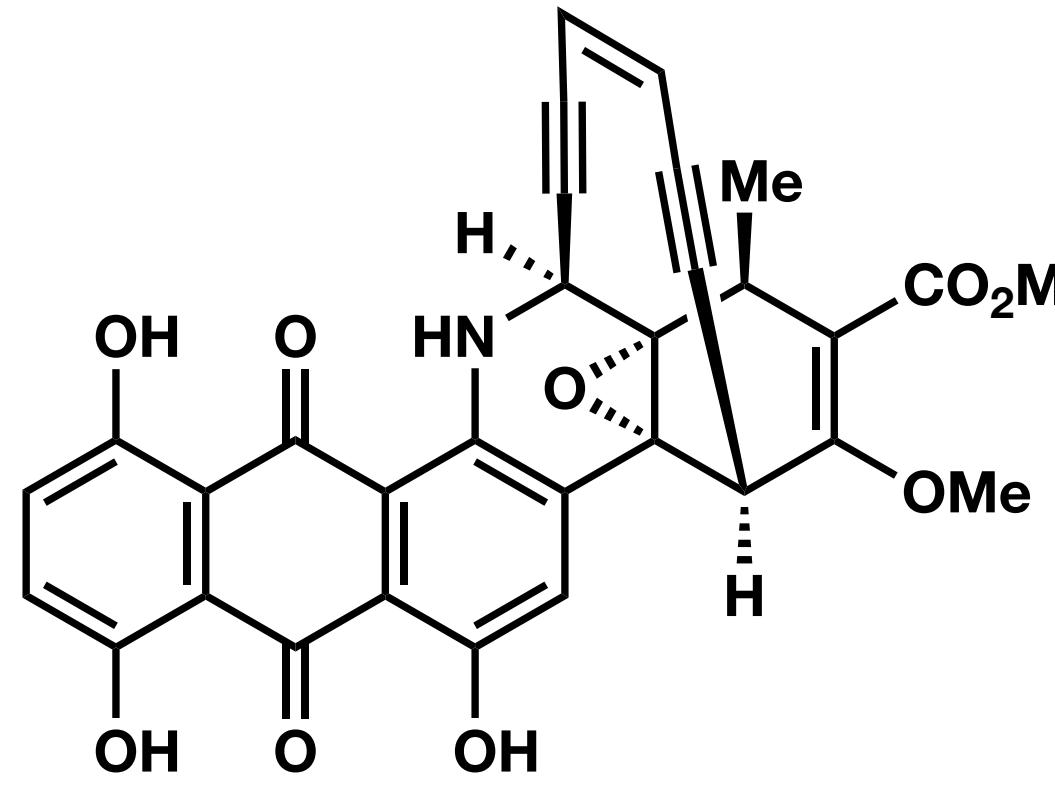
# Dynemicin A



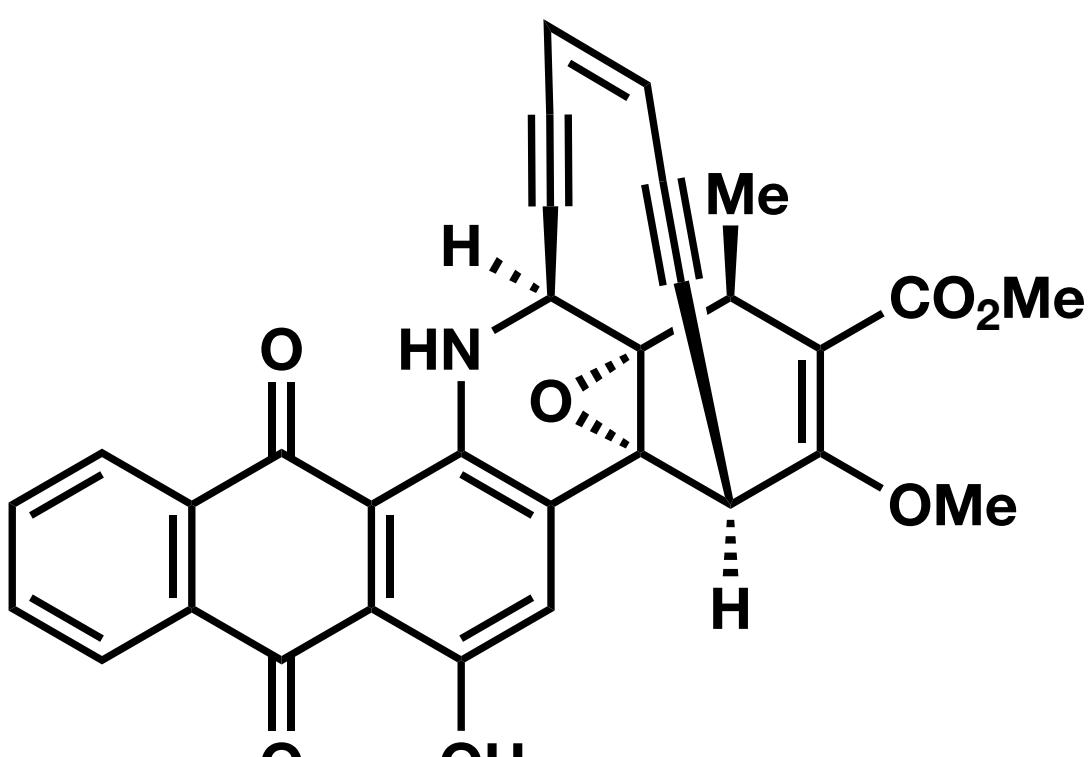
$$K_B = (5 \pm 2) \times 10^4 \text{ M}^{-1}$$



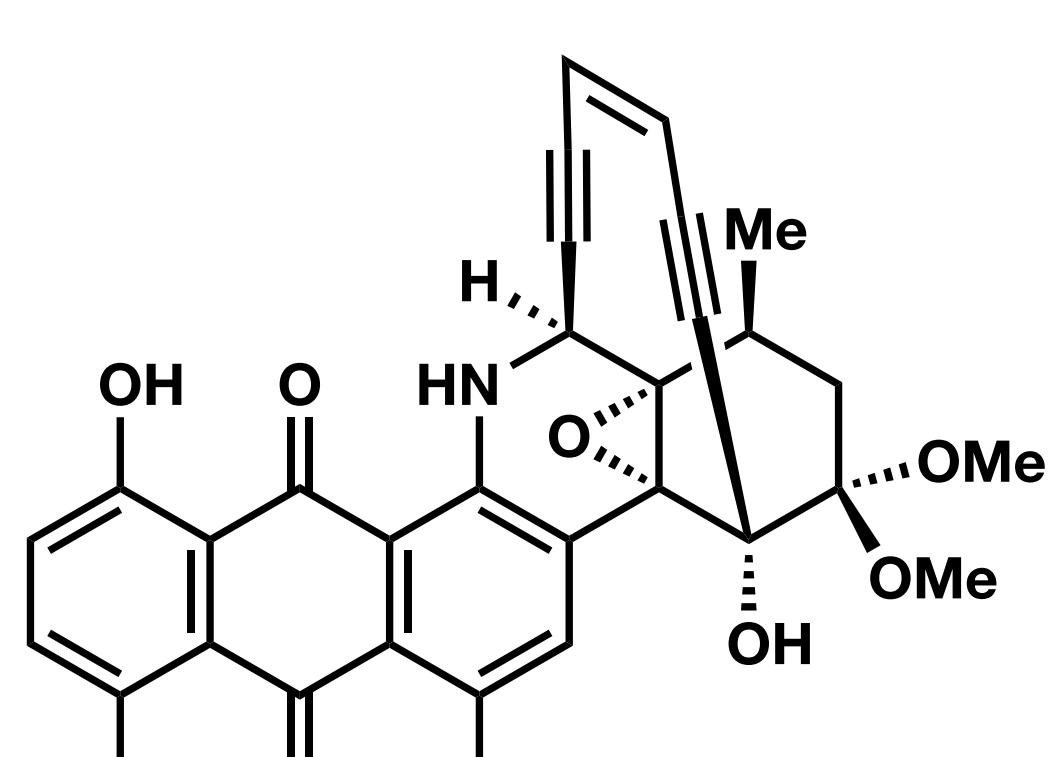
$$K_B = (6 \pm 1) \times 10^2 \text{ M}^{-1}$$



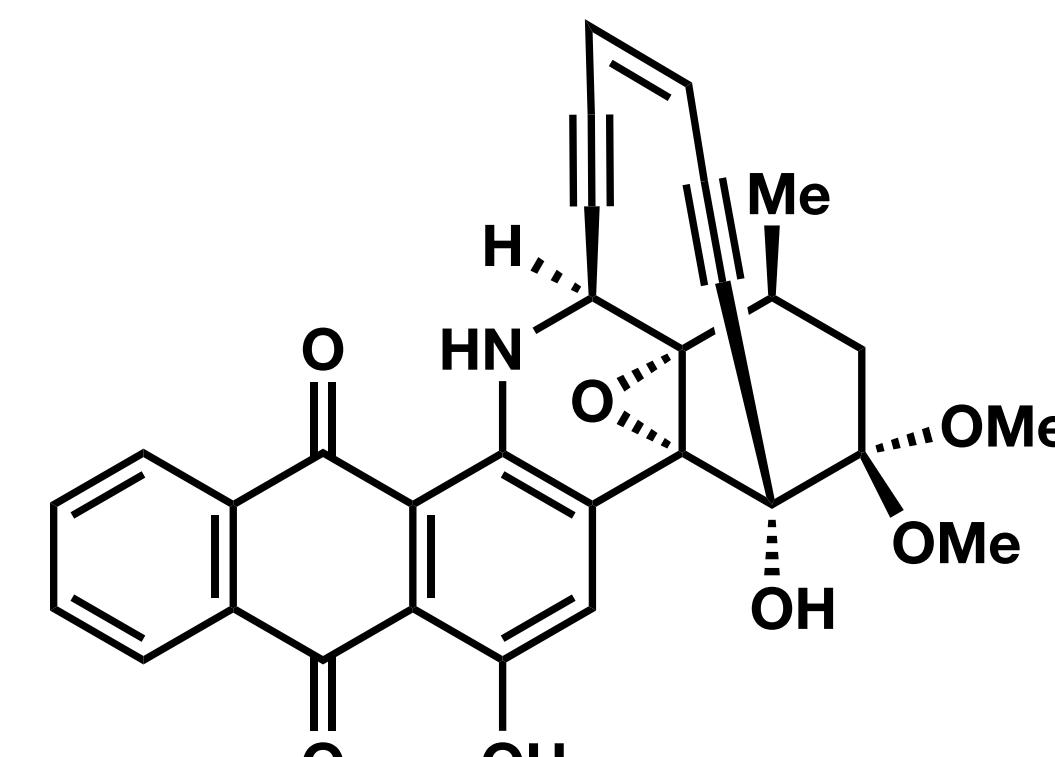
$$K_B = (8 \pm 2) \times 10^6 \text{ M}^{-1}$$



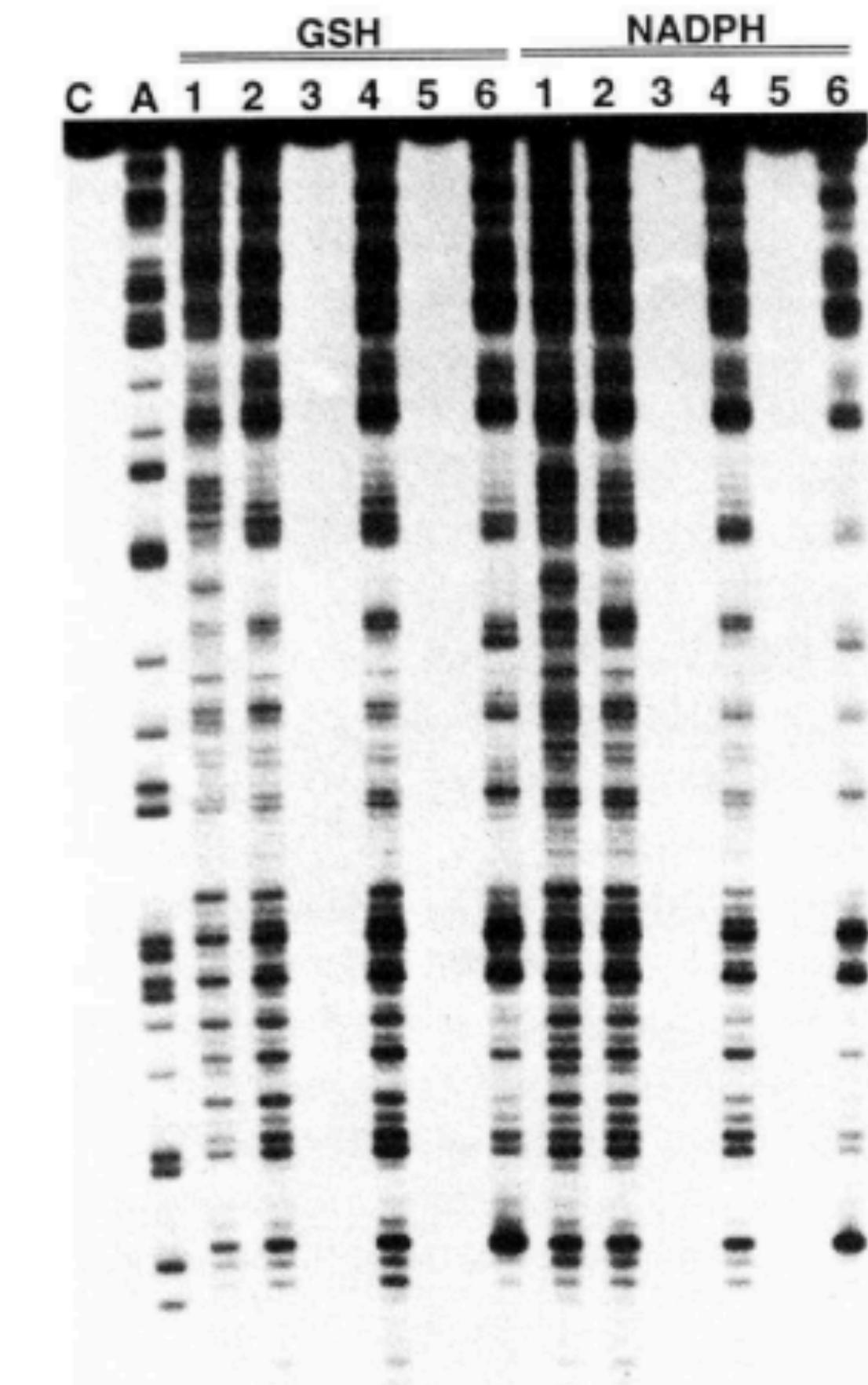
$$K_B = (5 \pm 2) \times 10^4 \text{ M}^{-1}$$



$$K_B = (4 \pm 2) \times 10^6 \text{ M}^{-1}$$

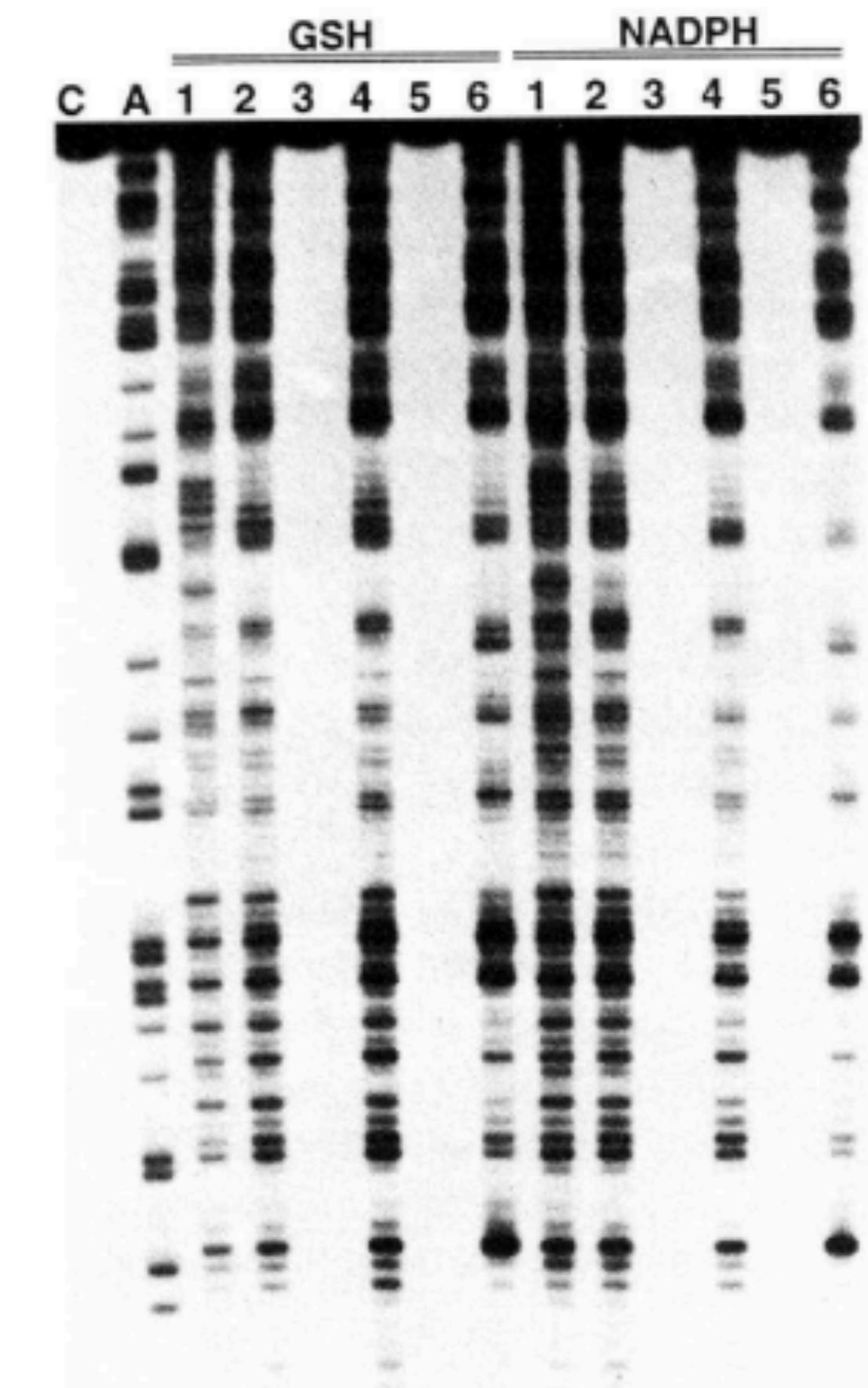
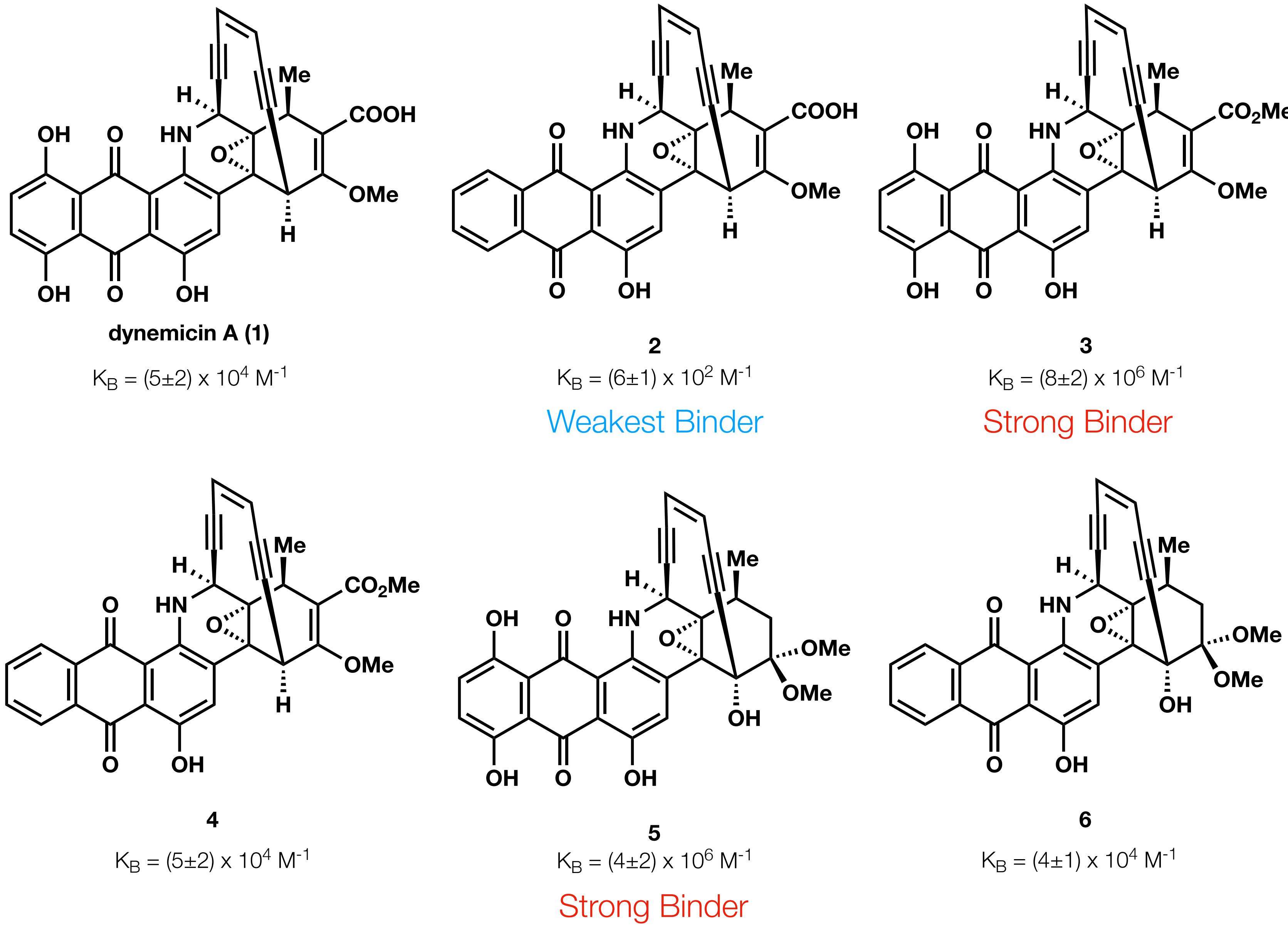


$$K_B = (4 \pm 1) \times 10^4 \text{ M}^{-1}$$



**Figure 2.** Cleavage of a 5'-<sup>32</sup>P-labeled 193-base-pair restriction fragment of pBR322 (*Eco*RI/*Ssp*I digests) by **1–6** and GSH or NADPH [calf thymus DNA (1.0 mM bp), restriction fragment (~10<sup>5</sup> cpm), tris-HCl buffer (30 mM, pH 7.5), sodium chloride (50 mM), dynemicin A or synthetic anthraquinone (0.05 mM), 37 °C, 12 h]. Reactions initiated by addition of GSH (20 mM) or NADPH (20 mM), as indicated. Lane C: 193-bp restriction fragment alone. Lane A: products from an adenine-specific cleavage reaction (Iverson, B. L.; Dervan, P. B. *Nucleic Acids Res.* **1987**, *15*, 7823).

# Dynemicin A



**Figure 2.** Cleavage of a 5'-<sup>32</sup>P-labeled 193-base-pair restriction fragment of pBR322 (*Eco*RI/*Ssp*I digests) by **1–6** and GSH or NADPH [calf thymus DNA (1.0 mM bp), restriction fragment (~10<sup>5</sup> cpm), tris-HCl buffer (30 mM, pH 7.5), sodium chloride (50 mM), dynemicin A or synthetic anthraquinone (0.05 mM), 37 °C, 12 h]. Reactions initiated by addition of GSH (20 mM) or NADPH (20 mM), as indicated. Lane C: 193-bp restriction fragment alone. Lane A: products from an adenine-specific cleavage reaction (Iverson, B. L.; Dervan, P. B. *Nucleic Acids Res.* **1987**, *15*, 7823).

# Dynemicin A

## Takeaways

If the DNA-cleaving agent binds too tightly to DNA, activation is prohibitively slow

Weakest binder should be most reactive

dynemicin A (**1**)

Validated experimentally, where **2** is ~50-fold more reactive towards GSH than **1**

However, at lower concentrations of DNA, **2** has decreased efficiency of cleaving DNA compared to **1**

Thus, the natural product, **1**, seems to strike an optimal balance (evolutionary?) between

i. Rate of Reaction with Nucleophile - where weak binding is advantageous

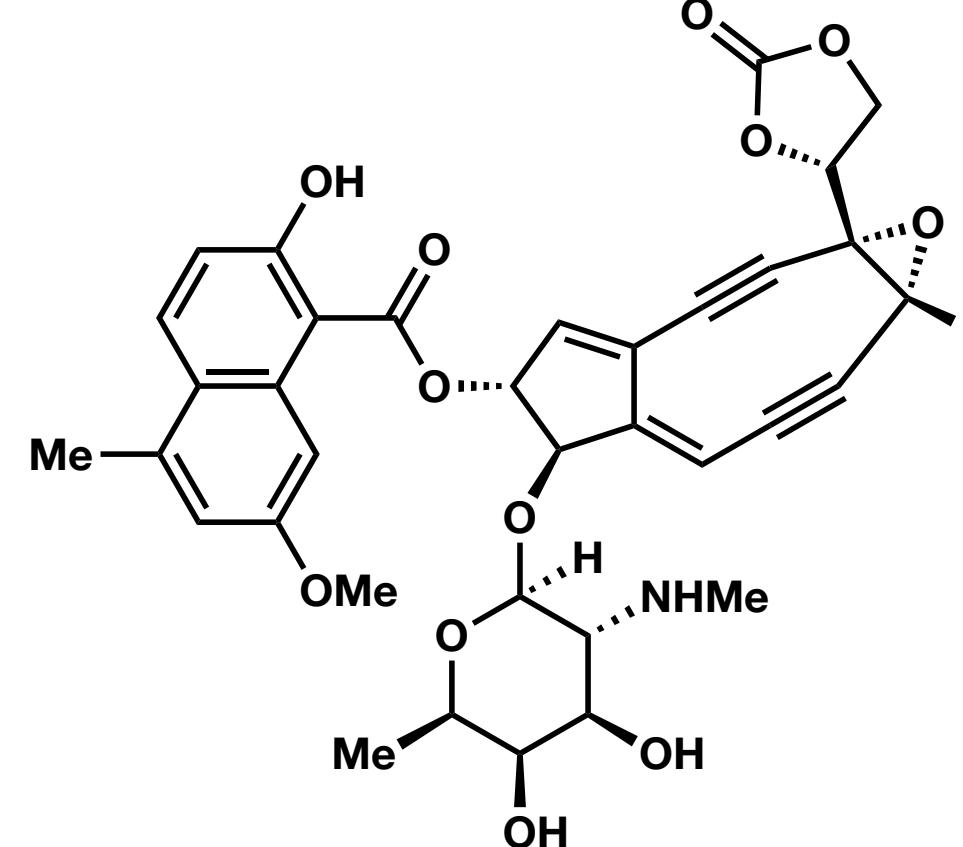
and

ii. Cleavage - where tight binding is advantageous

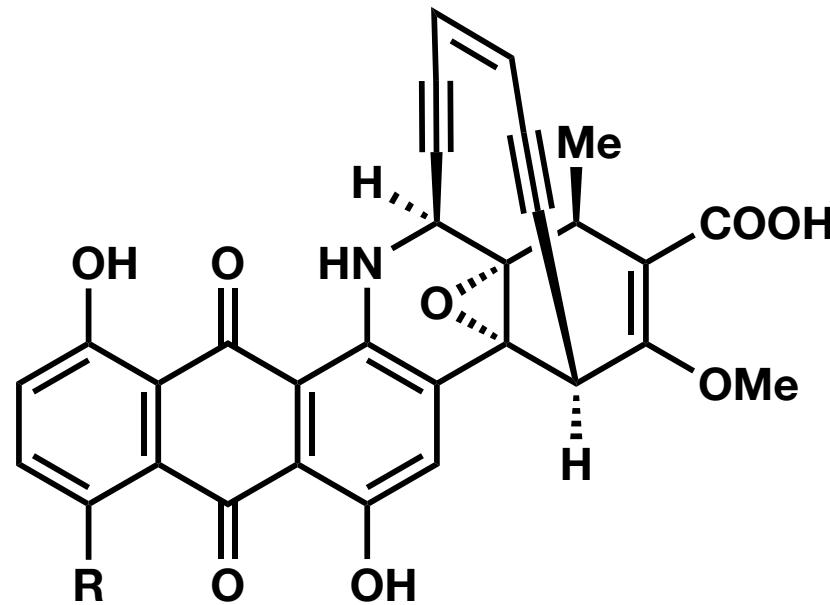
Figure 2. Cleavage of a 5'-<sup>32</sup>P-labeled 193-base-pair restriction fragment of pBR322 (*Eco*RI/*Ssp*I digests) by 1–6 and GSH or NADPH [calf thymus DNA (1.0 mM bp), restriction fragment (~10<sup>5</sup> cpm), tris-HCl buffer (30 mM, pH 7.5), sodium chloride (50 mM), dynemicin A or synthetic anthraquinone (0.05 mM), 37 °C, 12 h]. Reactions initiated by addition of GSH (20 mM) or NADPH (20 mM), as indicated. Lane C: 193-bp restriction fragment alone. Lane A: products from an adenine-specific cleavage reaction (Iverson, B. L.; Dervan, P. B. *Nucleic Acids Res.* 1987, 15, 7823).

# Ene-diyynes

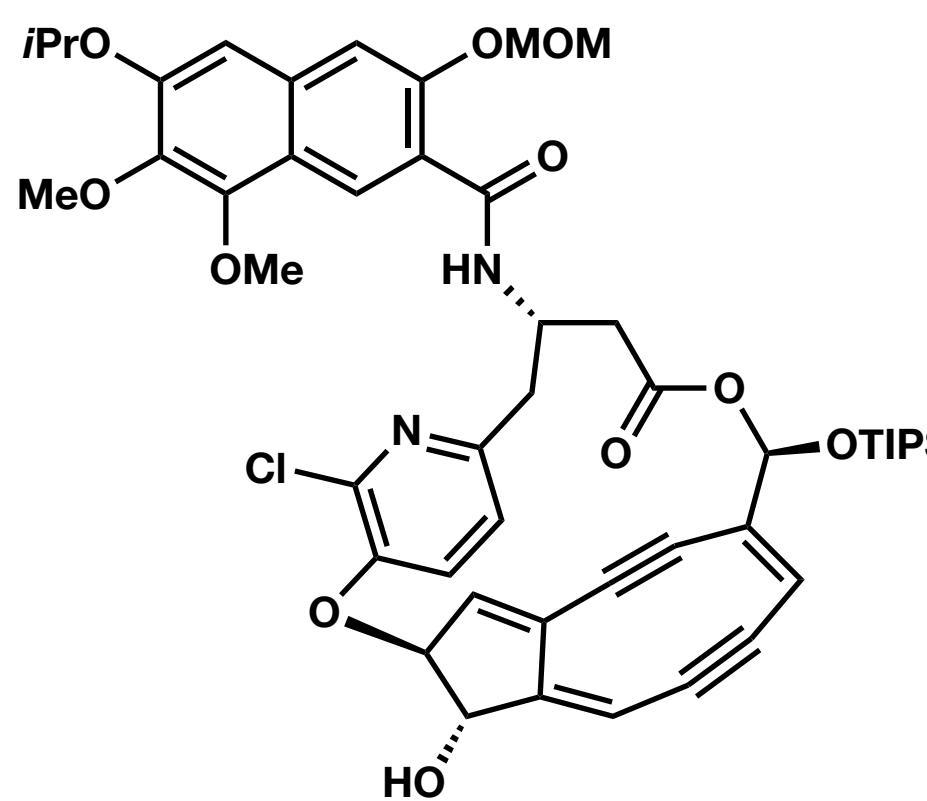
neocarzinostatin chromophore



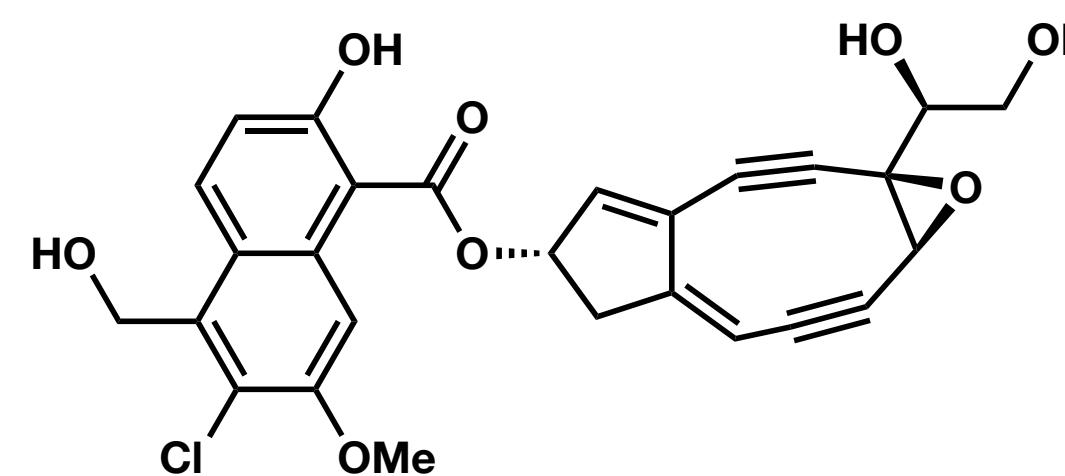
dynemicin A



kedarcidin chromophore



N1999A2



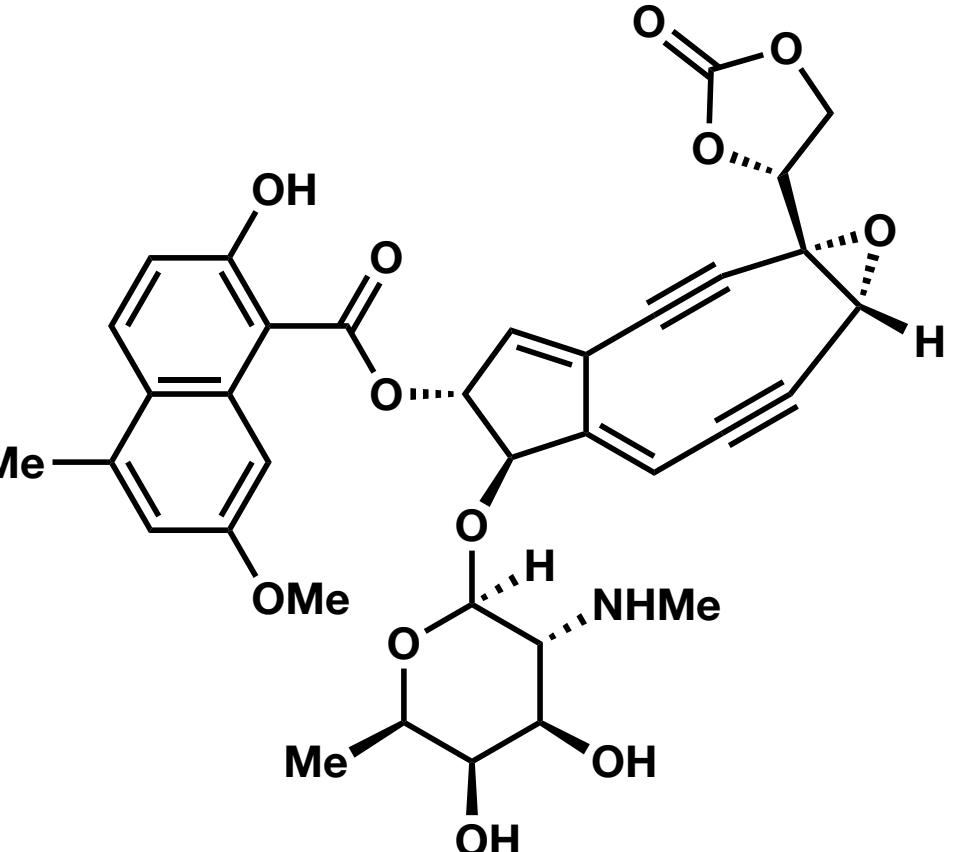
# Ene-diyne

## Overarching Research Theme

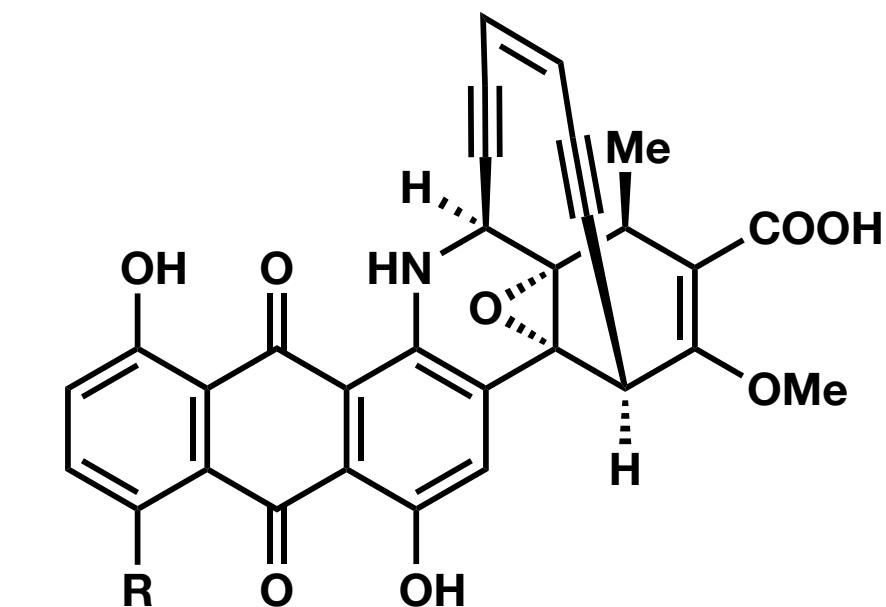
Utilize convergent syntheses of complex molecules for

- 1) the interrogation of biological mechanisms
- 2) structural elucidation
- 3) Unsuccessful (so far) in synthesizing unnatural analogs for improvement of bioactivity

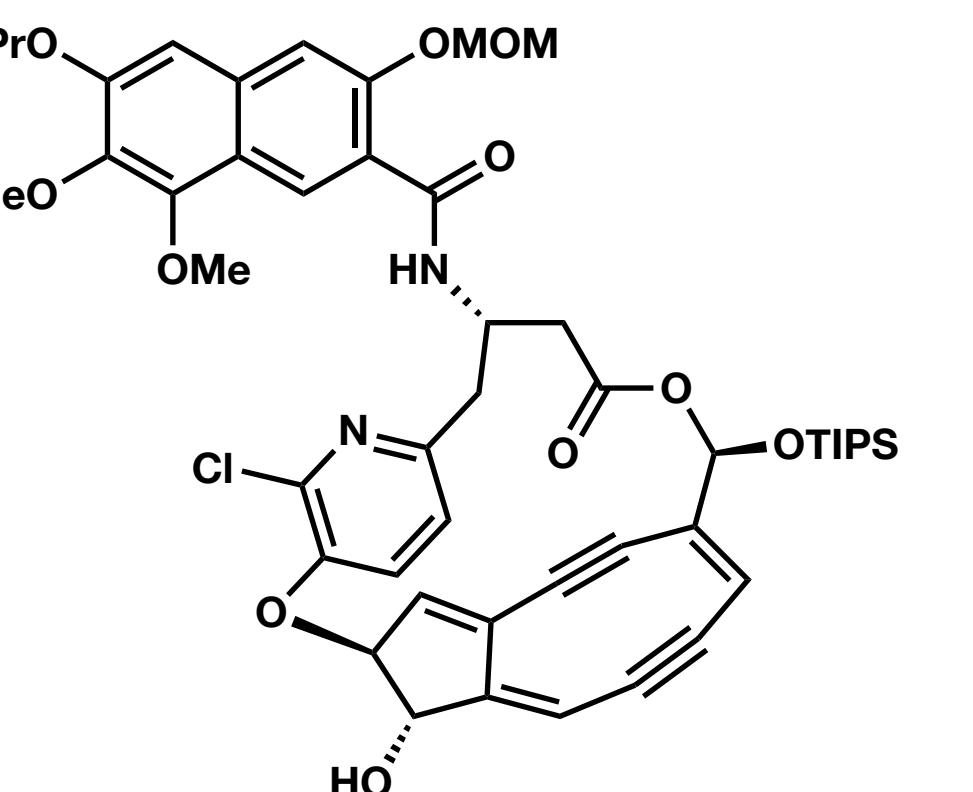
neocarzinostatin chromophore



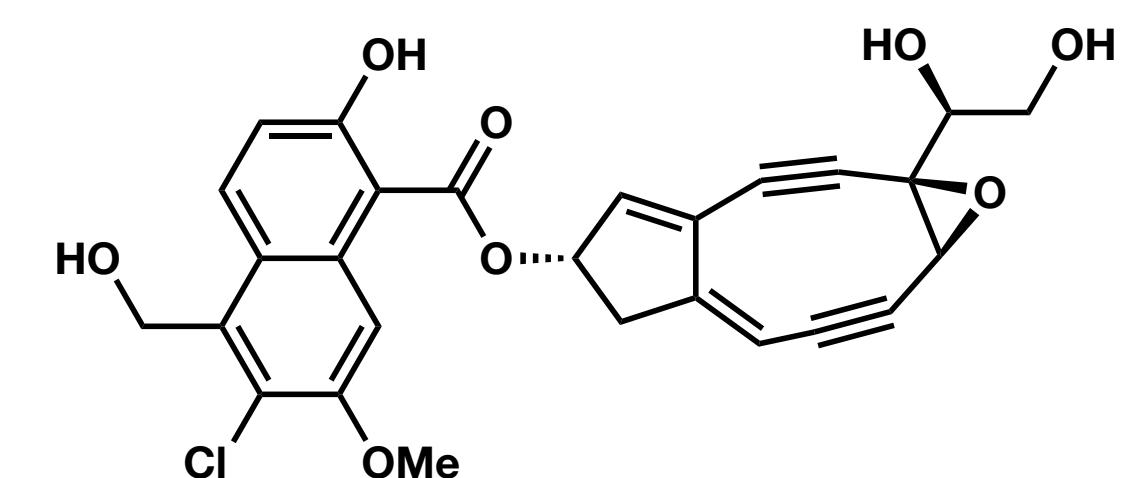
dynemicin A



kedarcidin chromophore

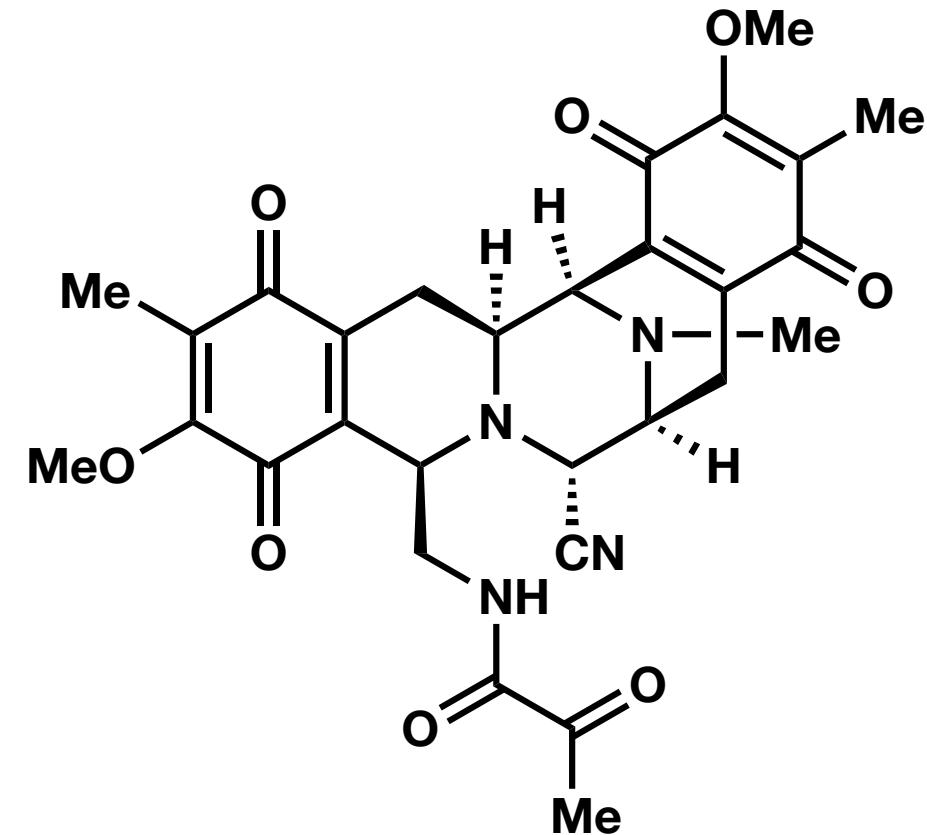


N1999A2

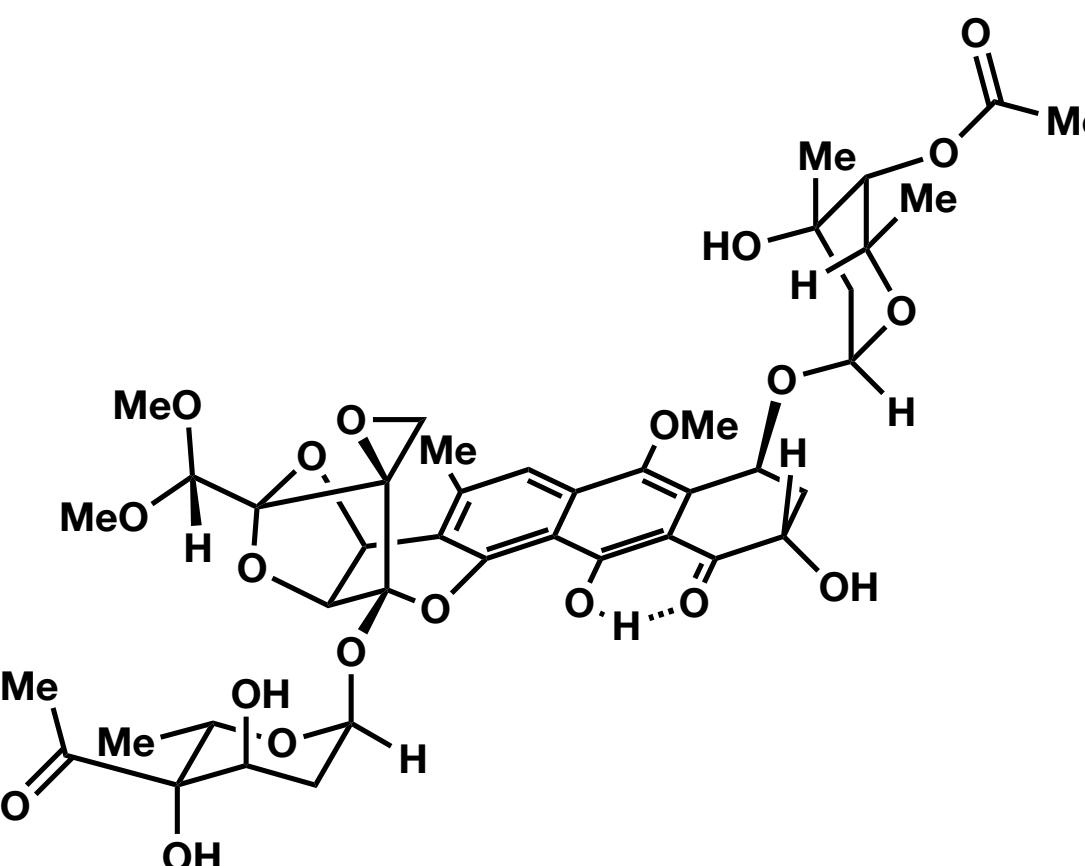


# DNA Alkylators

(-)-saframycin



trioxacarcins



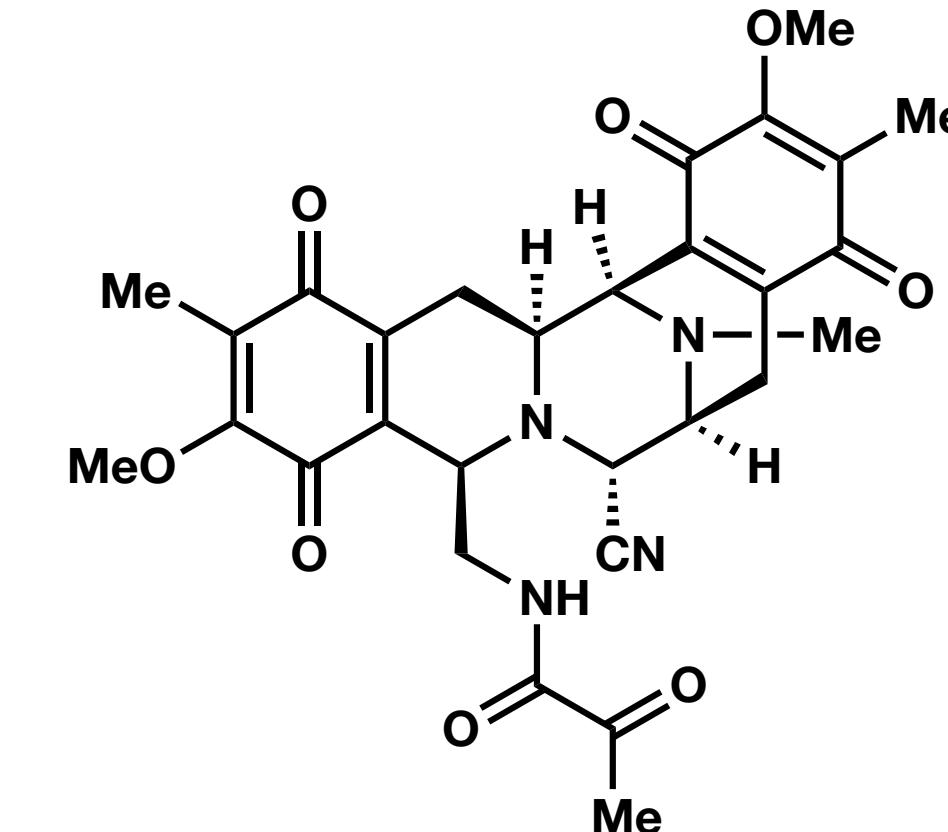
# DNA Alkylators

## Mechanism of Action

Nucleophiles or reductants trigger aromatization of quinone, facilitating intercalation

Protons promote iminium formation, which can be attacked by N2 of Guanine, resulting in DNA alkylation

(-)-saframycin

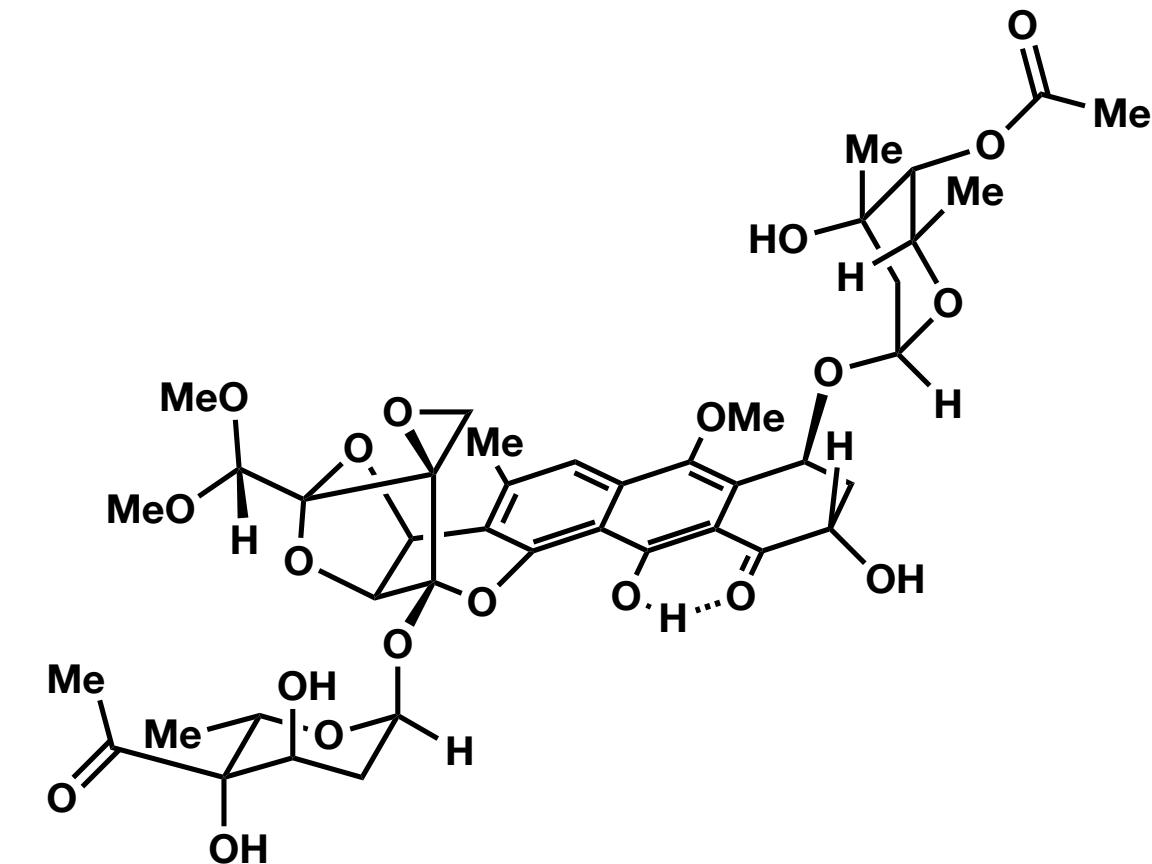


## Mechanism of Action

Crescent shape facilitates binding to the minor groove of DNA (GC-rich sequences)

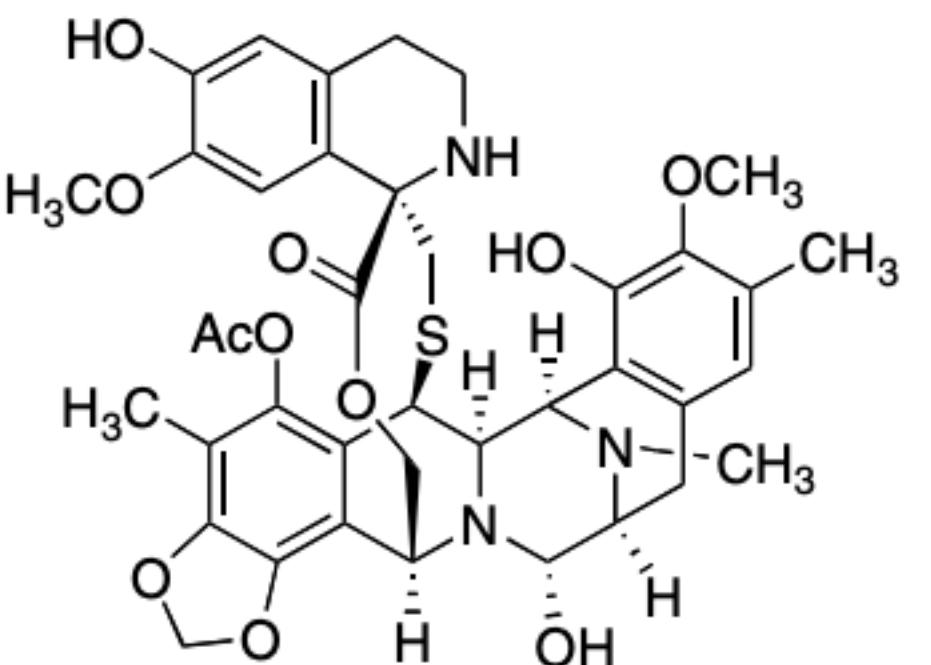
Spiroepoxide is susceptible to attack by N7 of Guanine, resulting in DNA alkylation

trioxacarcins



# DNA Alkylators

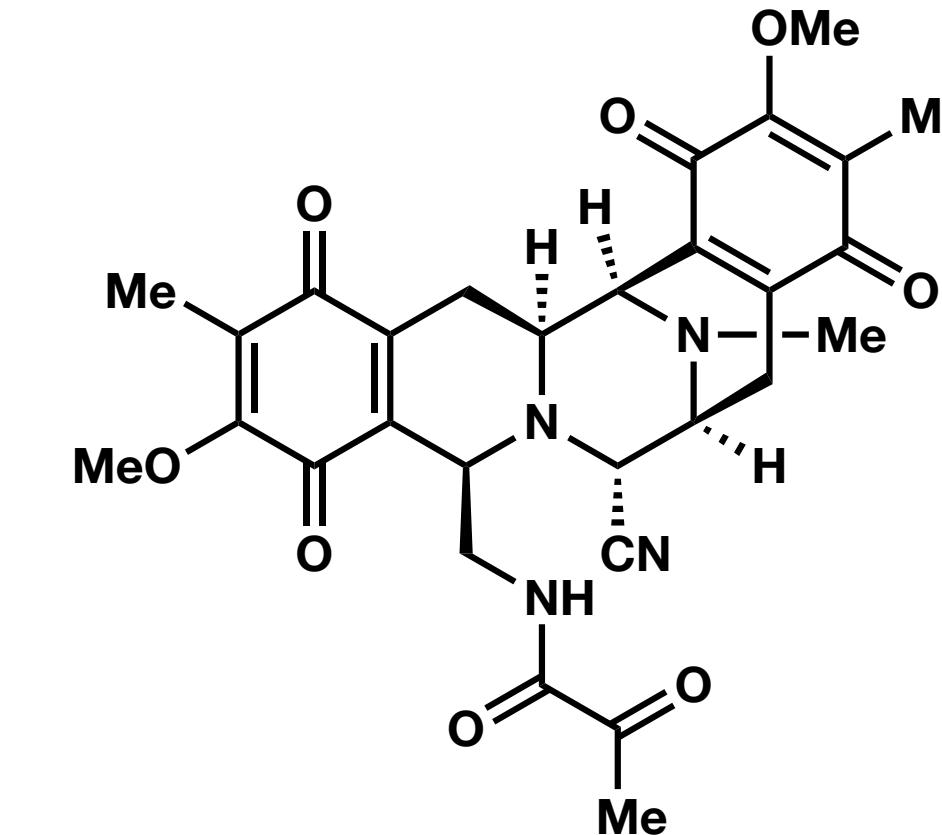
Related Natural Product: ET-743  
(Yondelis, FDA-approved in 2015)



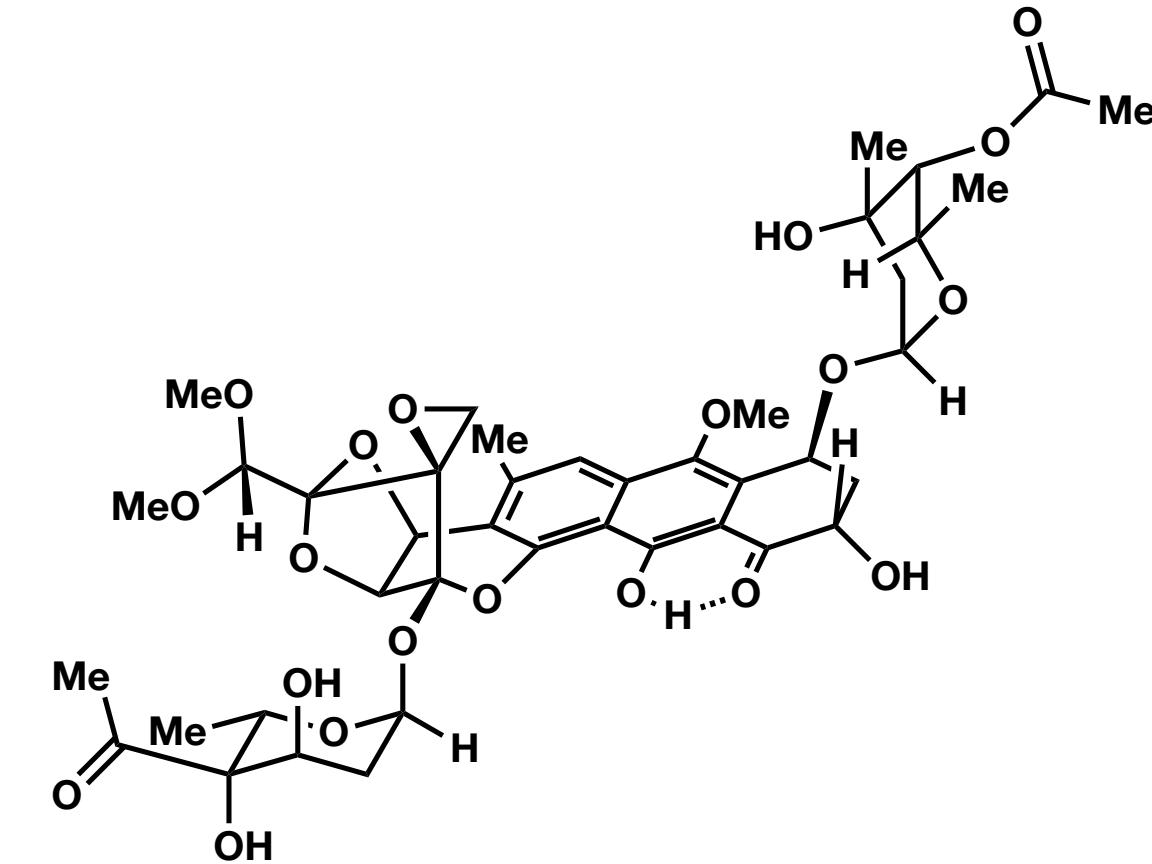
**ET-743 (2)**

For the treatment of advanced soft tissue sarcoma such as liposarcoma and leiomyosarcoma

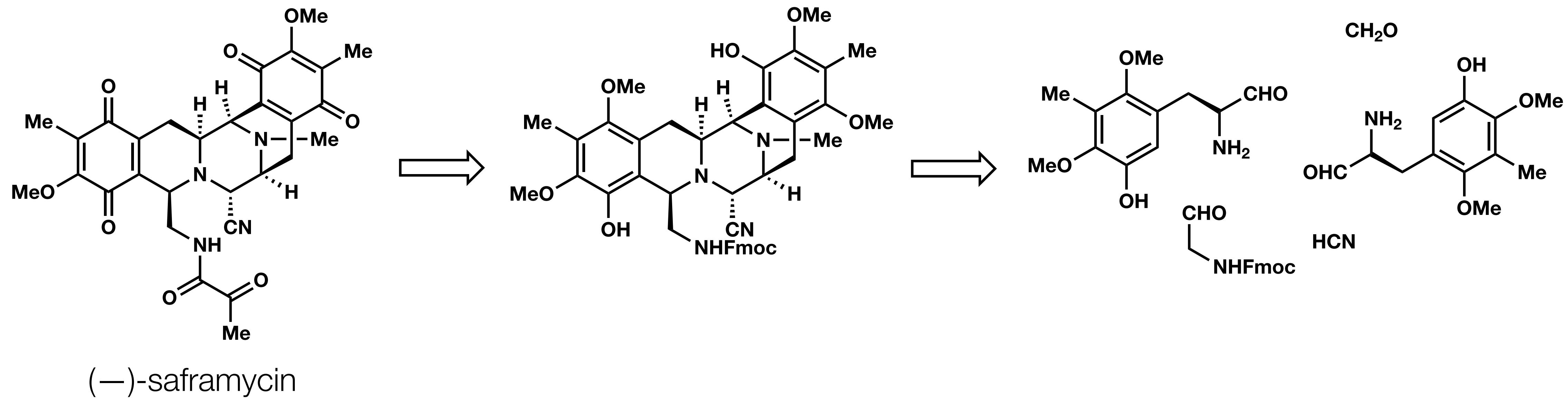
(-)-saframycin



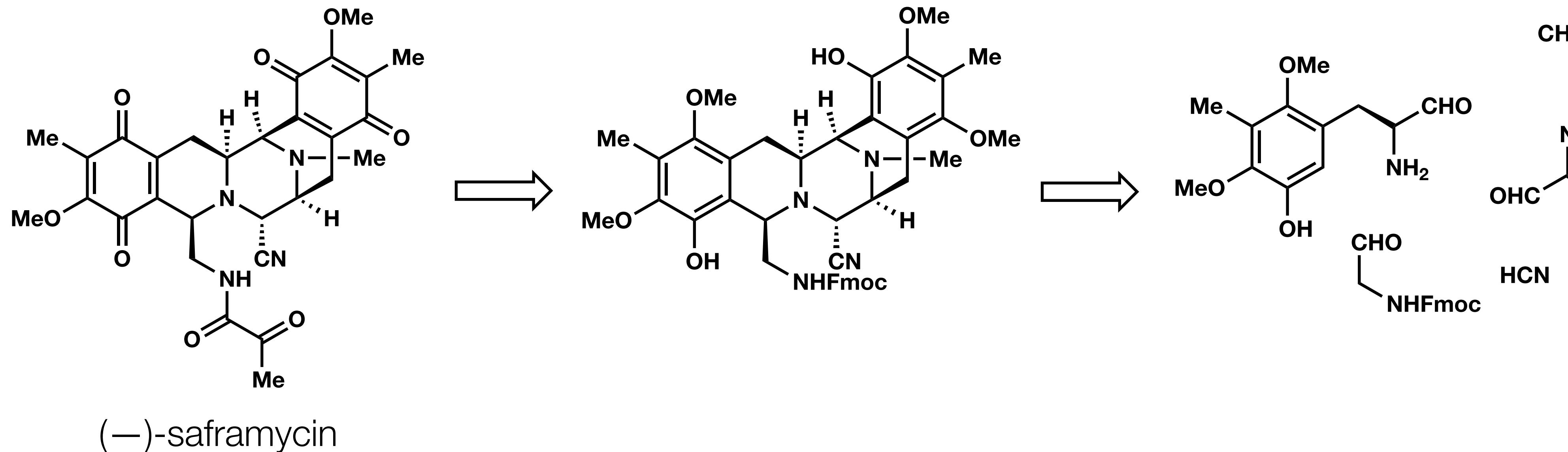
trioxacarcins



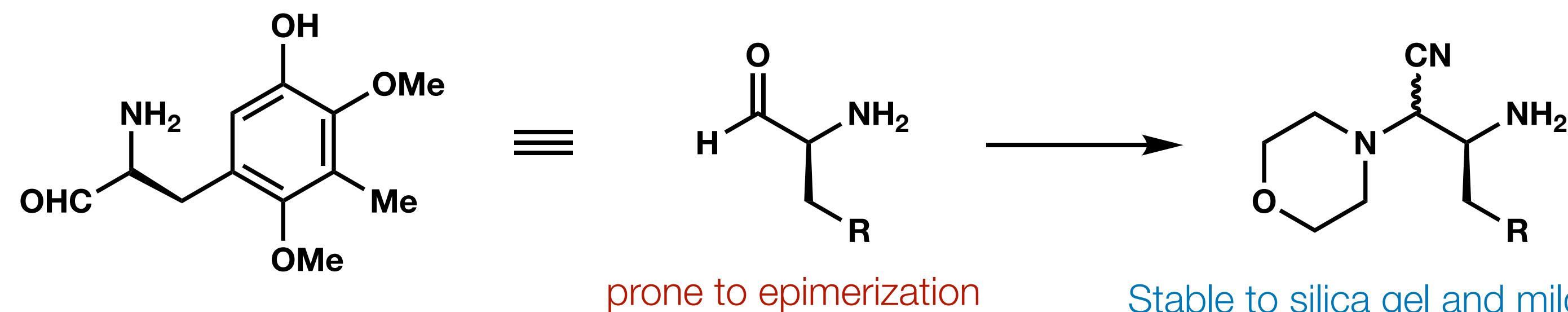
# Synthesis of Saframycin



# Synthesis of Saframycin



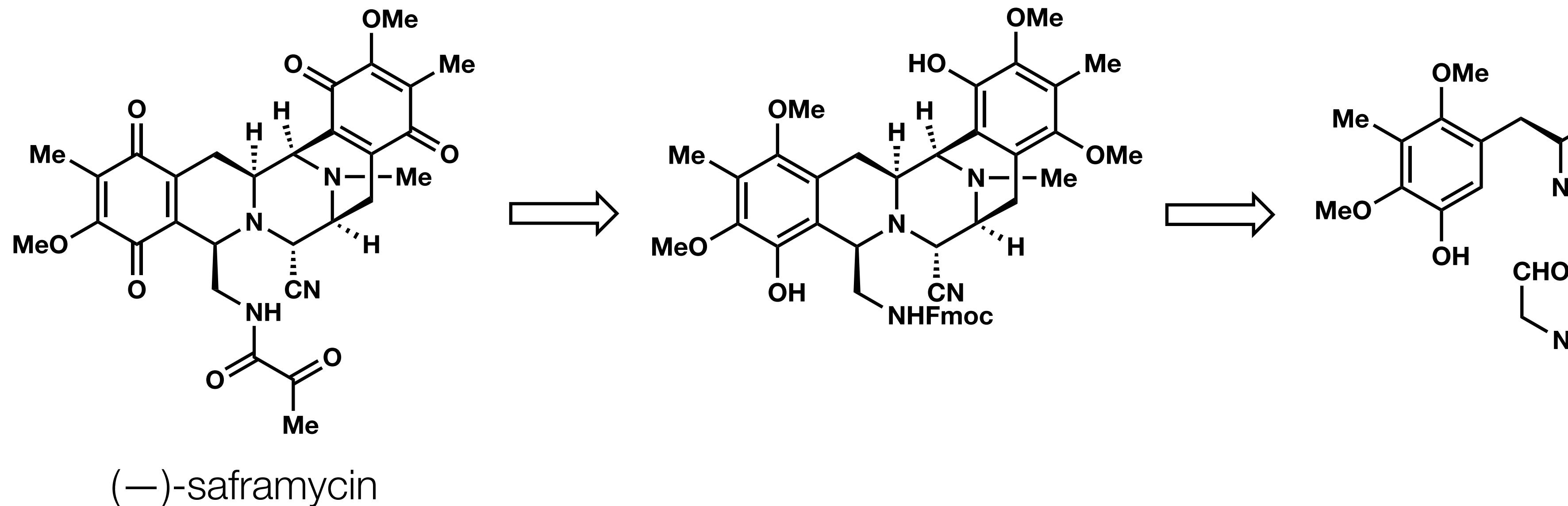
Key Method: “C-protected”  $\alpha$ -amino aldehydes



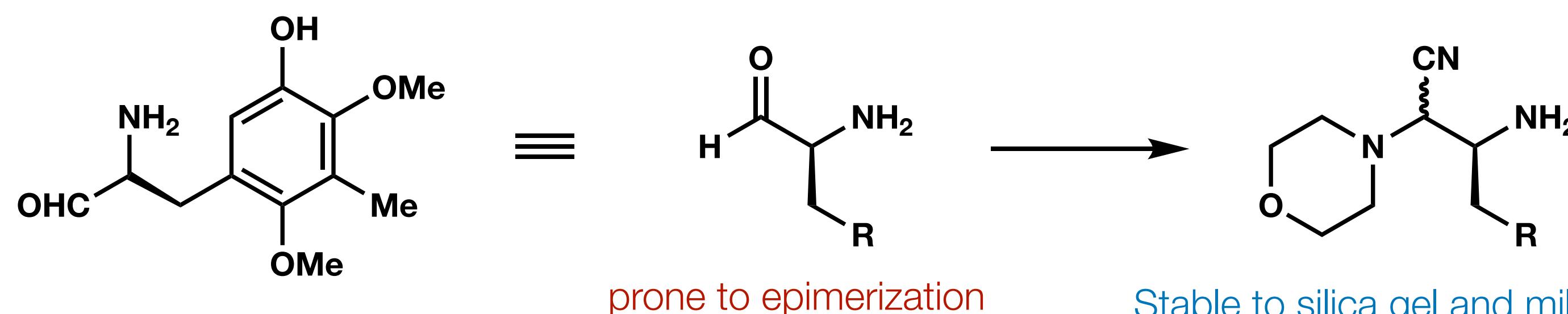
See: *J. Am. Chem. Soc.*  
**2000**, 122, 3236-3237

Stable to silica gel and mild acids  
Minimal epimerization observed during  
formation and subsequent  
manipulations (i.e. mannich, reduction,  
pictet-spenger)

# Synthesis of Saframycin



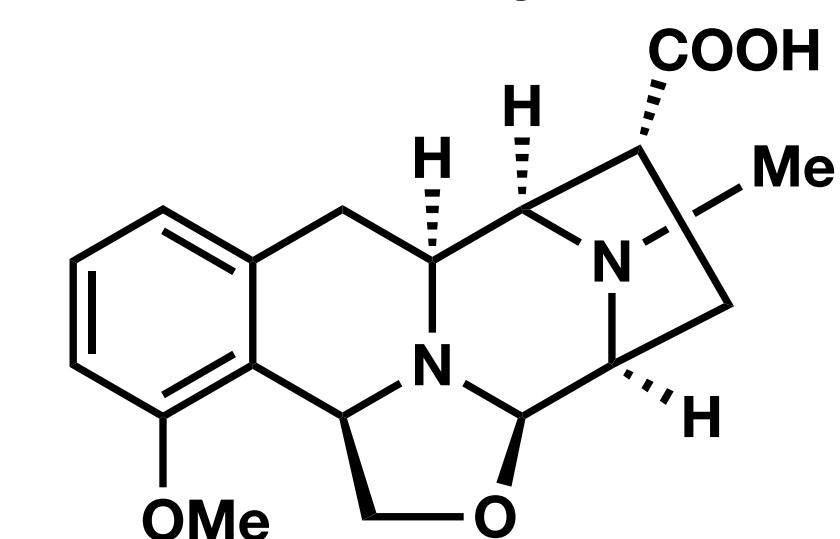
Key Method: “C-protected”  $\alpha$ -amino aldehydes



See: *J. Am. Chem. Soc.*  
2000, 122, 3236-3237

Stable to silica gel and mild acids  
Minimal epimerization observed during  
formation and subsequent  
manipulations (i.e. mannich, reduction,  
pictet-spenger)

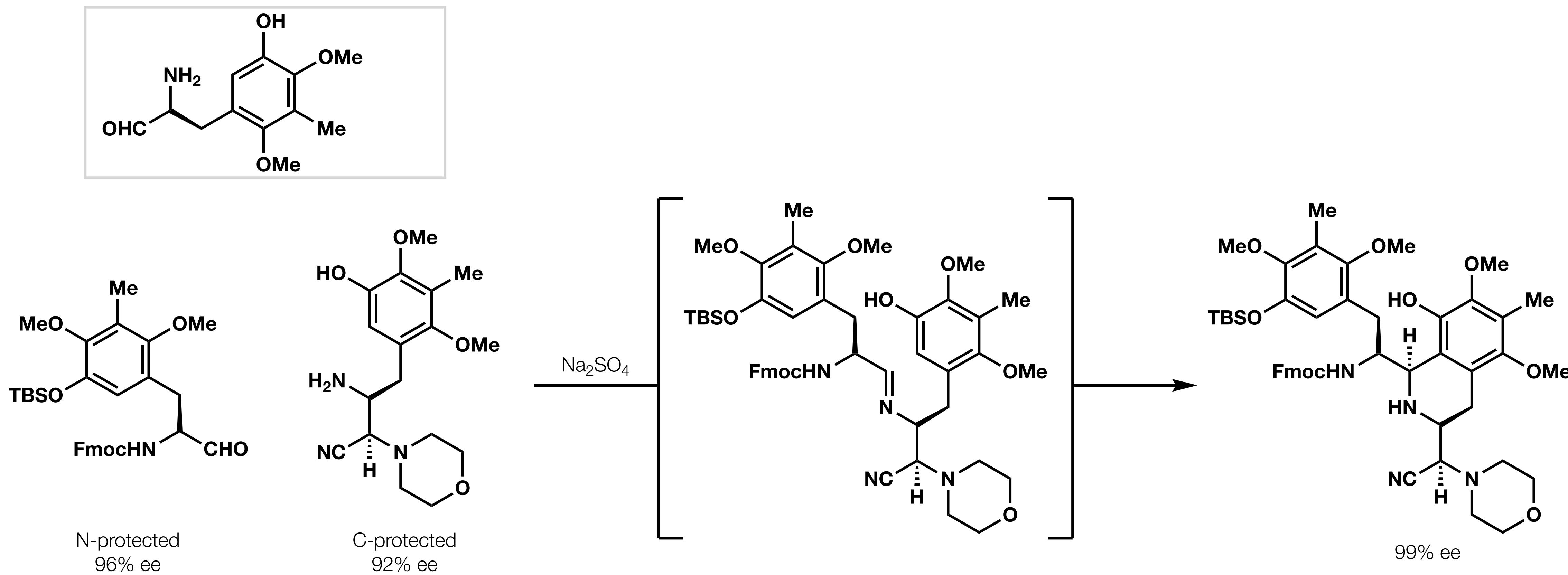
Similar strategy was applied  
towards the synthesis of



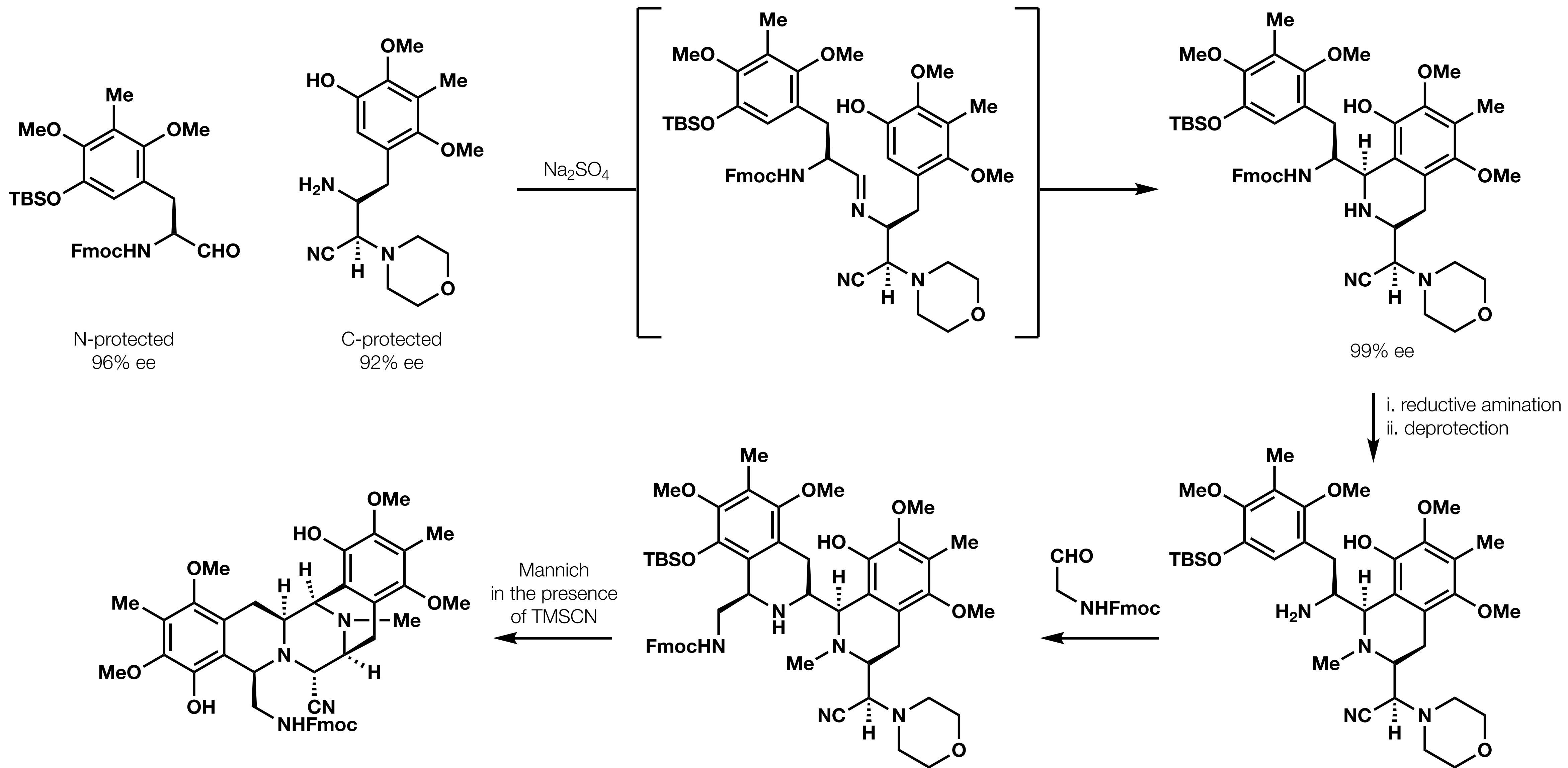
(-)quinocarcin

*JACS* 2005; 127(48): 16796–16797.

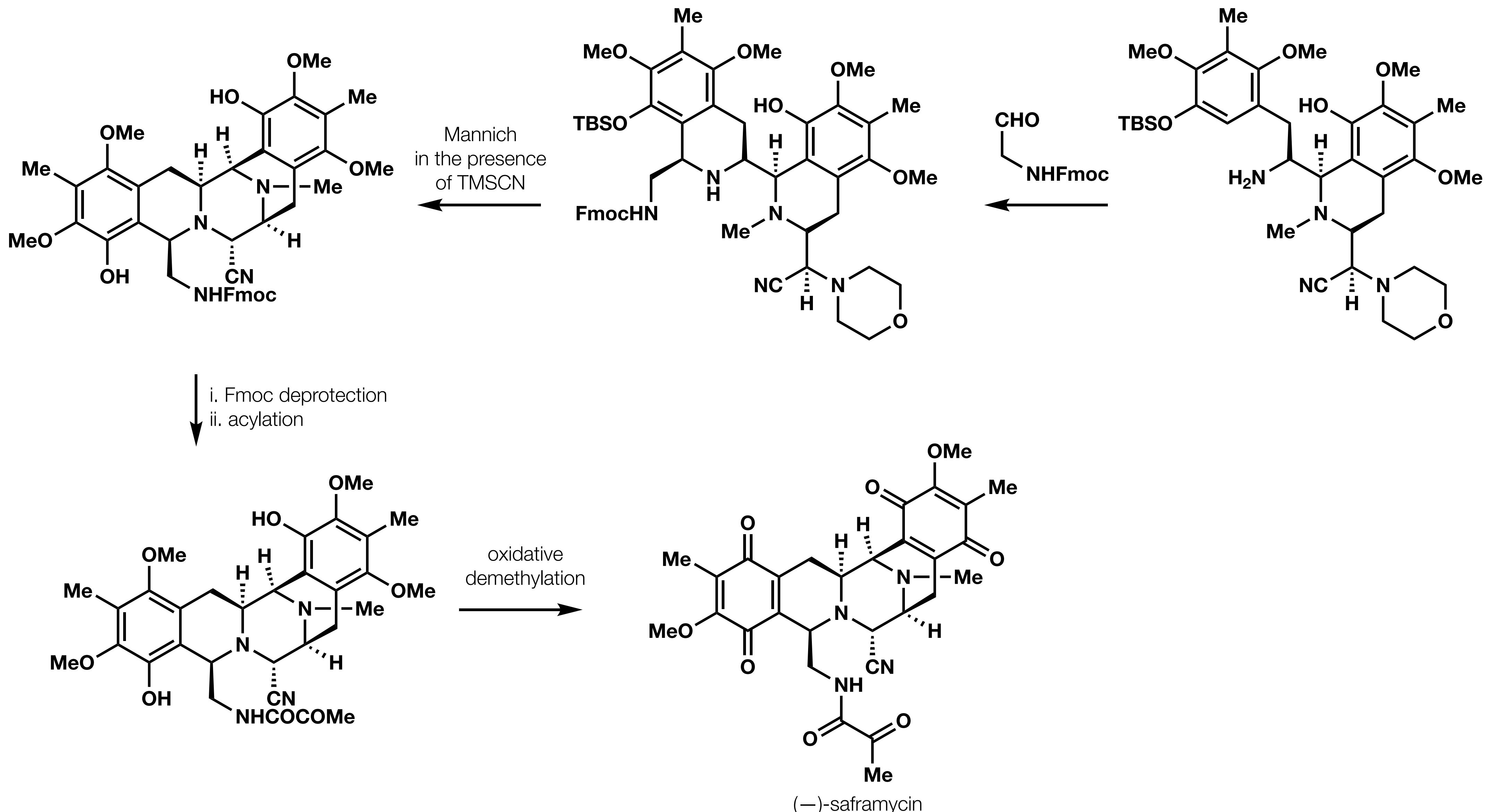
# Synthesis of Saframycin



# Synthesis of Saframycin

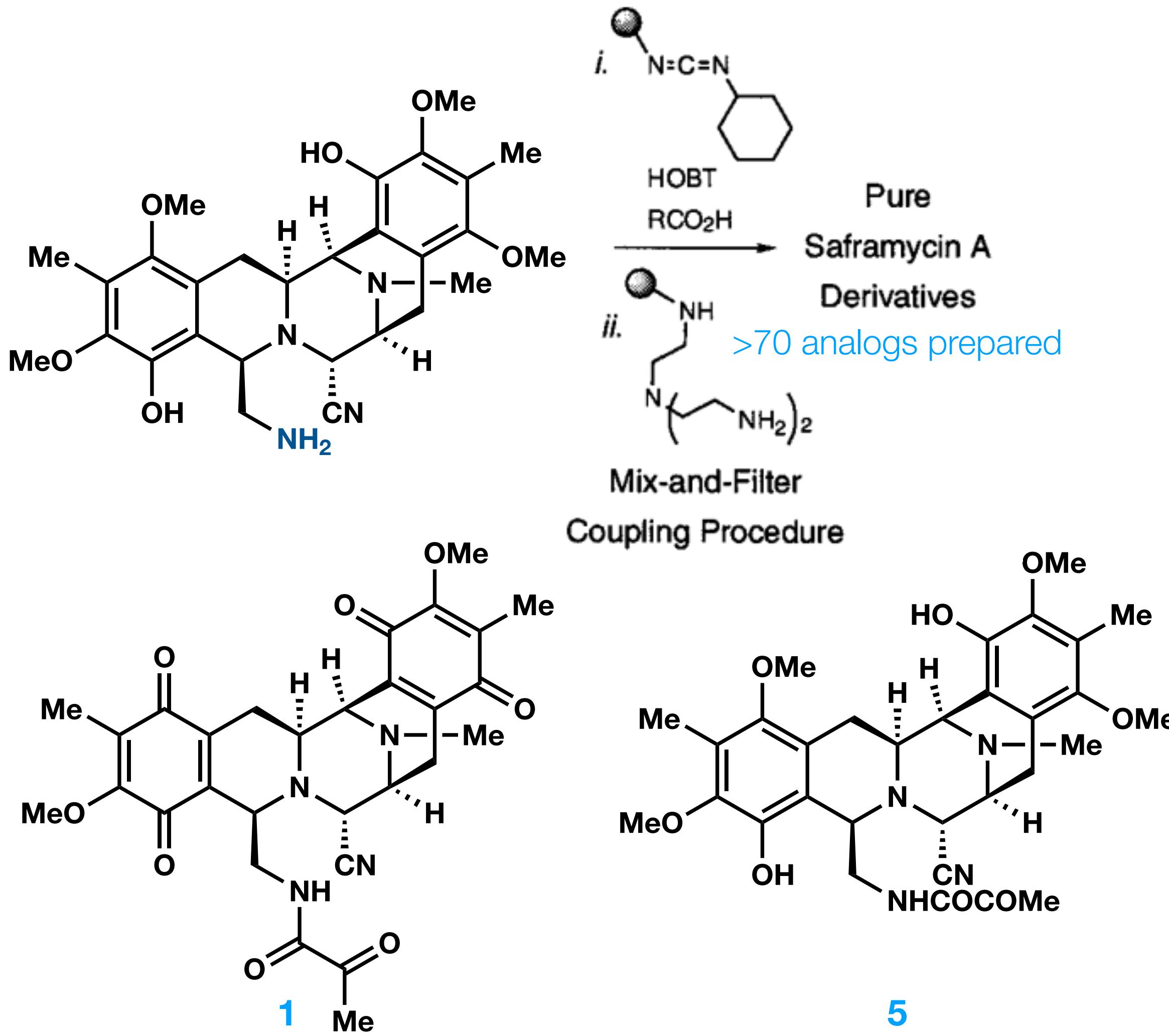


# Synthesis of Saframycin



# Saframycin Analogs and Antiproliferative Activities

Prior to approval of Yondelis, Saframycin derivatives were studied as a potential alternative that mitigates side effects such as body weight loss and hepatic toxicity.



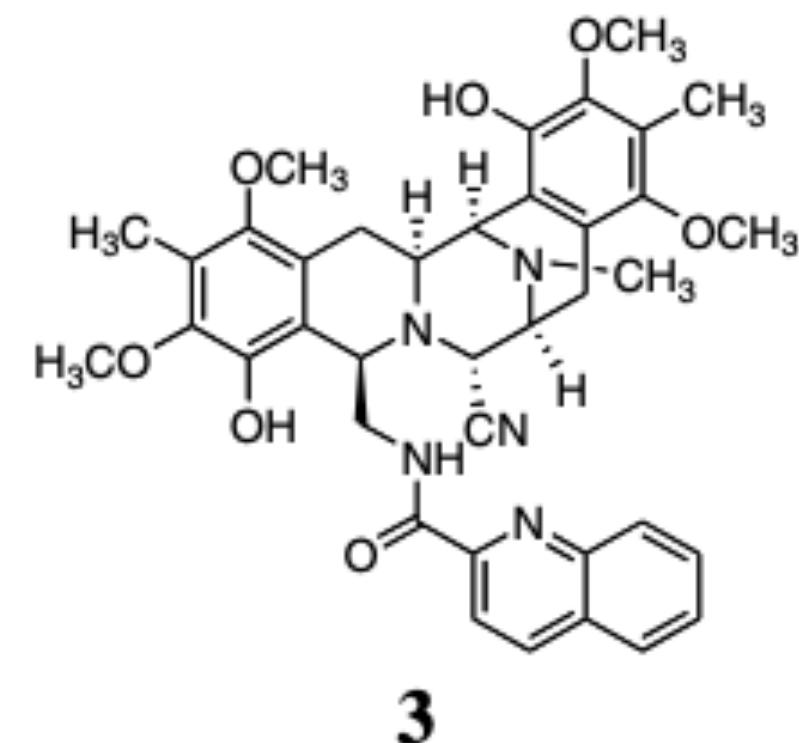
**Table 1.** Some of the Most Potent Bishydroquinone Derivatives of Saframycin A and Their Antiproliferative Activities

<b>N =</b>	<b>IC<sub>50</sub>, nM</b>		<b>N =</b>	<b>IC<sub>50</sub>, nM</b>	
	A375	A549		A375	A549
Saframycin A ( <b>1</b> )	5.3	133	( <b>10</b> )	2.7	31
( <b>5</b> )	4.5	160	( <b>11</b> )	1.7	9.2
			( <b>12</b> )	3.3	40
			( <b>13</b> )	2.5	32
			( <b>14</b> )	1.3	4.4
			( <b>15</b> )	1.4	4.6
			( <b>16</b> ) <sup>10</sup>	2.0	3.5
				1.5	4.1
				1.2	4.7
				3.6	78

A375 melanoma and A549 lung carcinoma

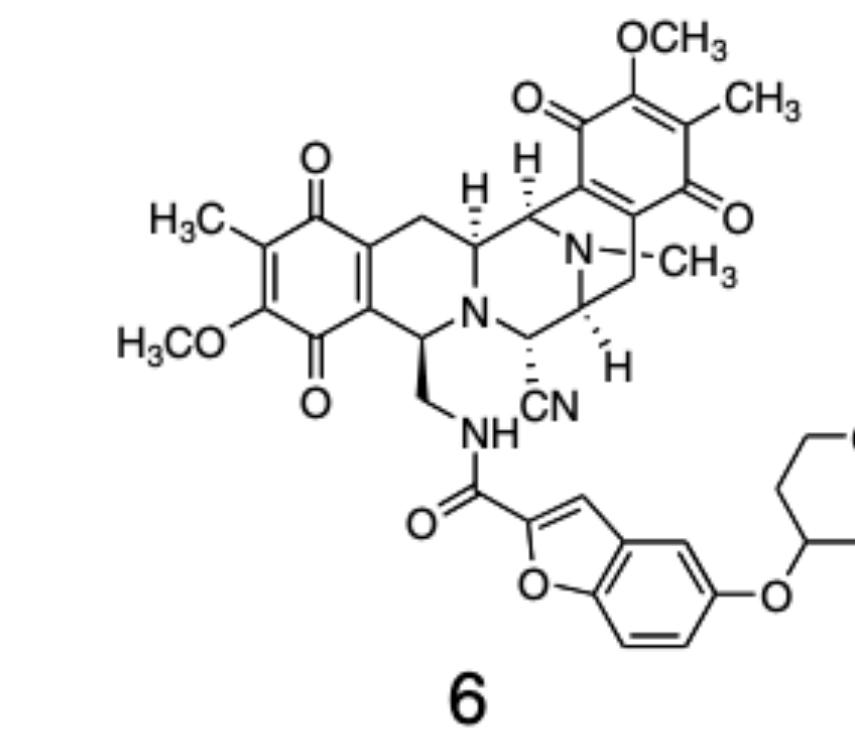
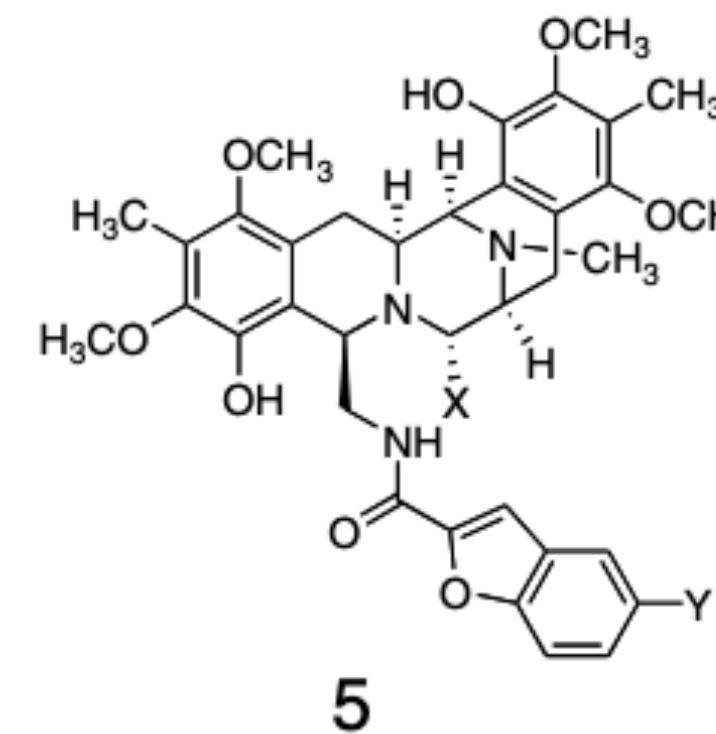
# Saframycin Analogs and Antiproliferative Activities

**Table 1.** GI50 values for the in vitro growth inhibitory effects of reference compound **3** in tumor cells from different tissues



Cell type	Histology	GI50 <sup>a</sup> (nM)
DLD-1	Colon	8.1
HCT-116		5.8
HT-29		7.5
CWR 22Rv1	Prostate	1.1
DU145		6.9
PC-3		6.2
A549	Lung	10.7
H1299		6.2
MDA-MB-231	Breast	4.5
HeLa-S3	Cervical	6.6
HT-1080	Fibrosarcoma	5.0
BxPC-3	Pancreatic	5.8
Nalm-6	B-cell leukemia <sup>b</sup>	0.6
P12	T-cell leukemia <sup>b</sup>	1.9

**Table 2.** GI50 values for the growth inhibitory effects of saframycin analogs in tumor cells



X	Y	Yield (%)	GI50 (nM)			
			HCT-116	DLD-1	PC-3	A549
<b>5a</b>	CN	H	72	5.5	ND <sup>b</sup>	ND <sup>b</sup>
<b>5b</b>	CN	Tetrahydropyran-4-yloxy	60	3.2	8.3	2.7
<b>5c</b>	CN	2-( <i>N</i> -Morpholino)-ethoxy	ND <sup>a</sup>	8.1	22.4	57.5
<b>5d</b>	OH	Tetrahydropyran-4-yloxy	58	5.5	10.4	5.0
<b>5e</b>	H	Tetrahydropyran-4-yloxy	45	>300	ND <sup>b</sup>	ND <sup>b</sup>
<b>6</b>			44	17.3	26.5	16.5

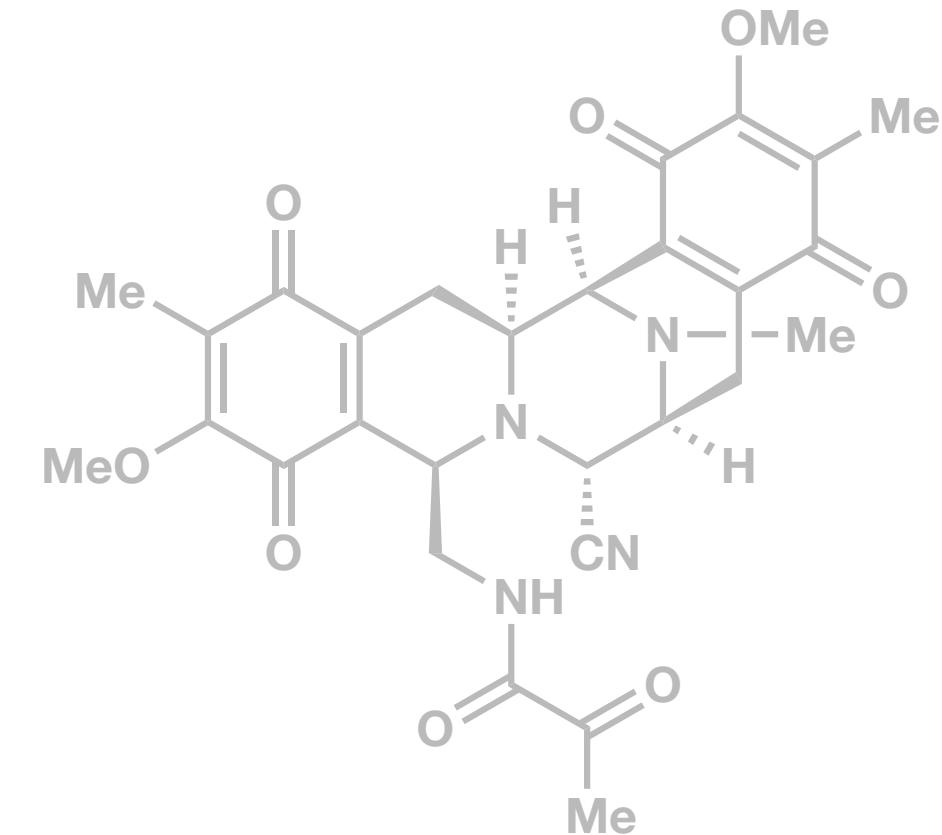
<sup>a</sup>Yield of last step not determined; <sup>b</sup>GI50 value not determined.

<sup>a</sup> Cells were grown in culture to a specific cell density and treated with compound for a period equal to two cell doubling times.

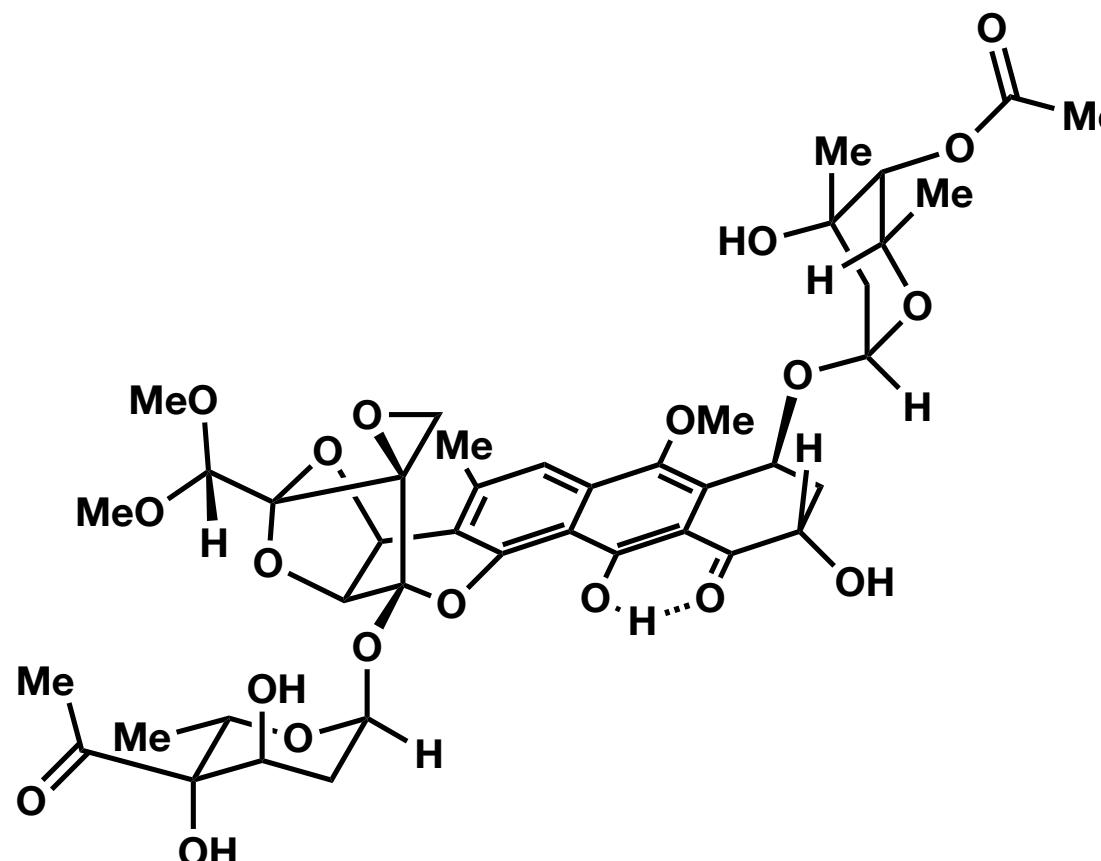
<sup>b</sup> Ref. 13.

# DNA Alkylators

(-)-saframycin

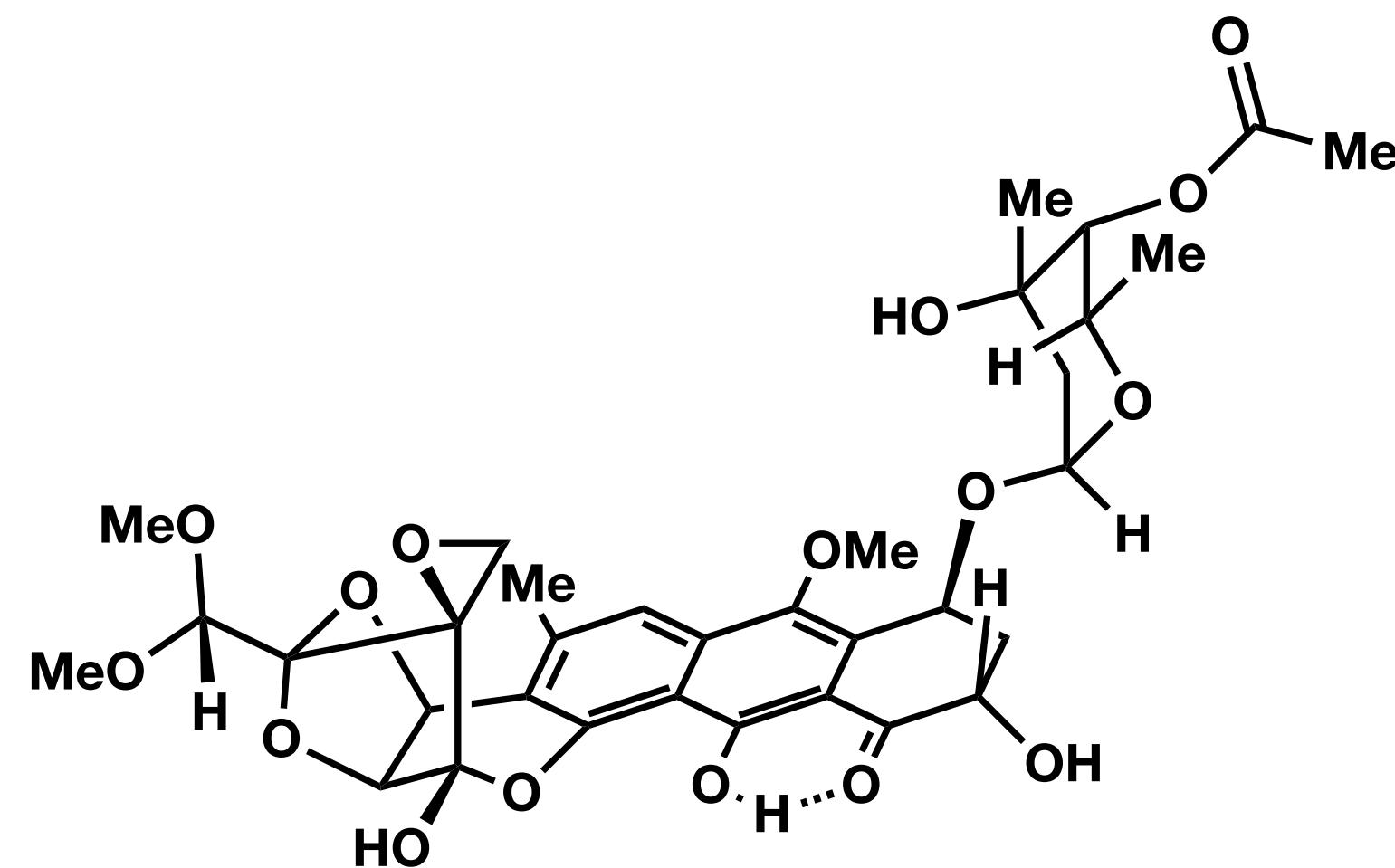


trioxacarcins

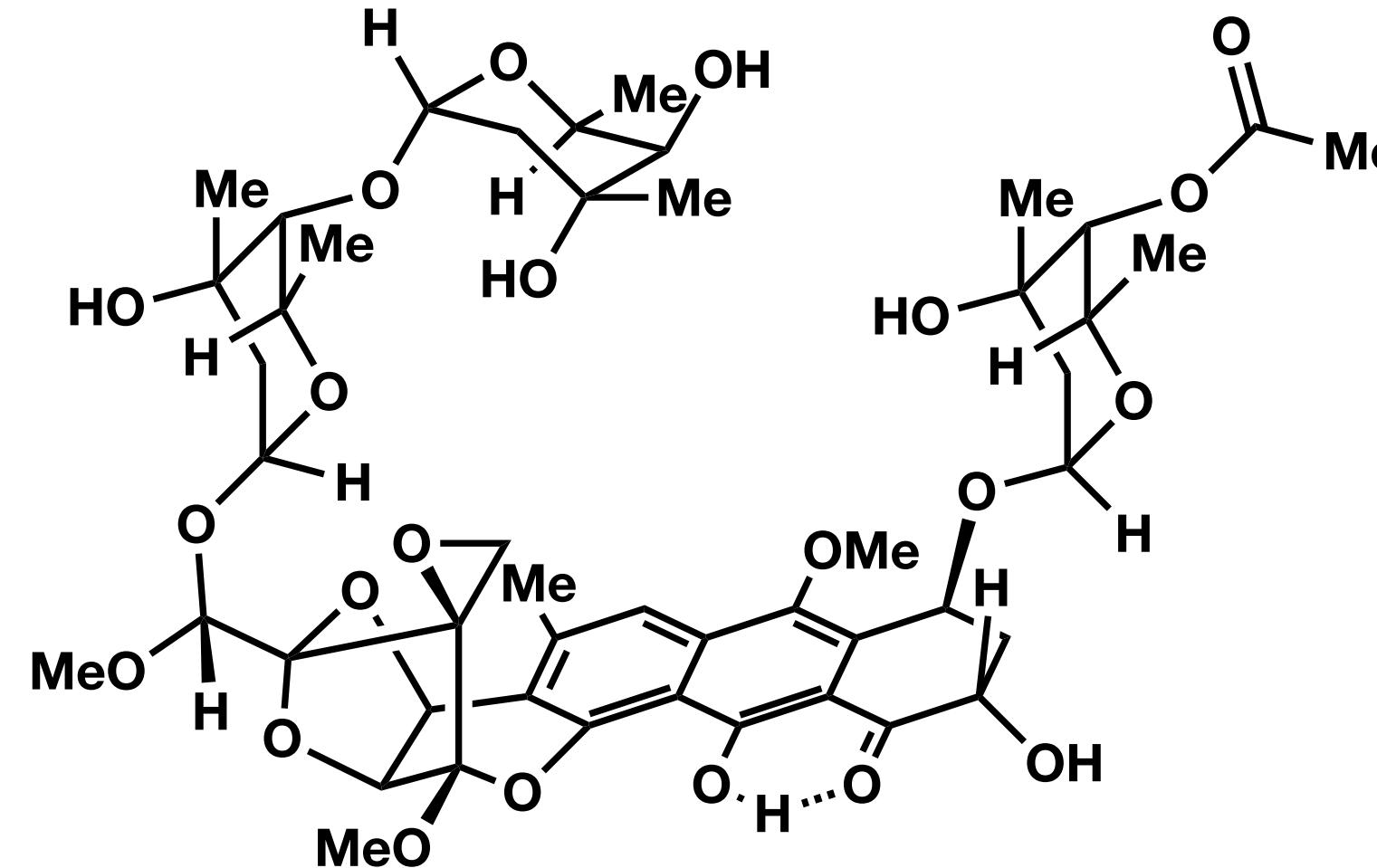


# Trioxacarcins

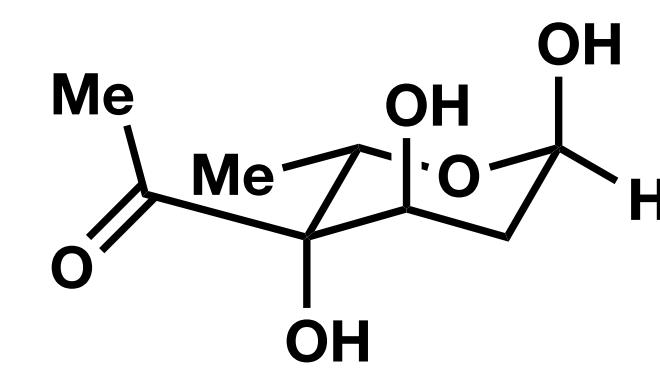
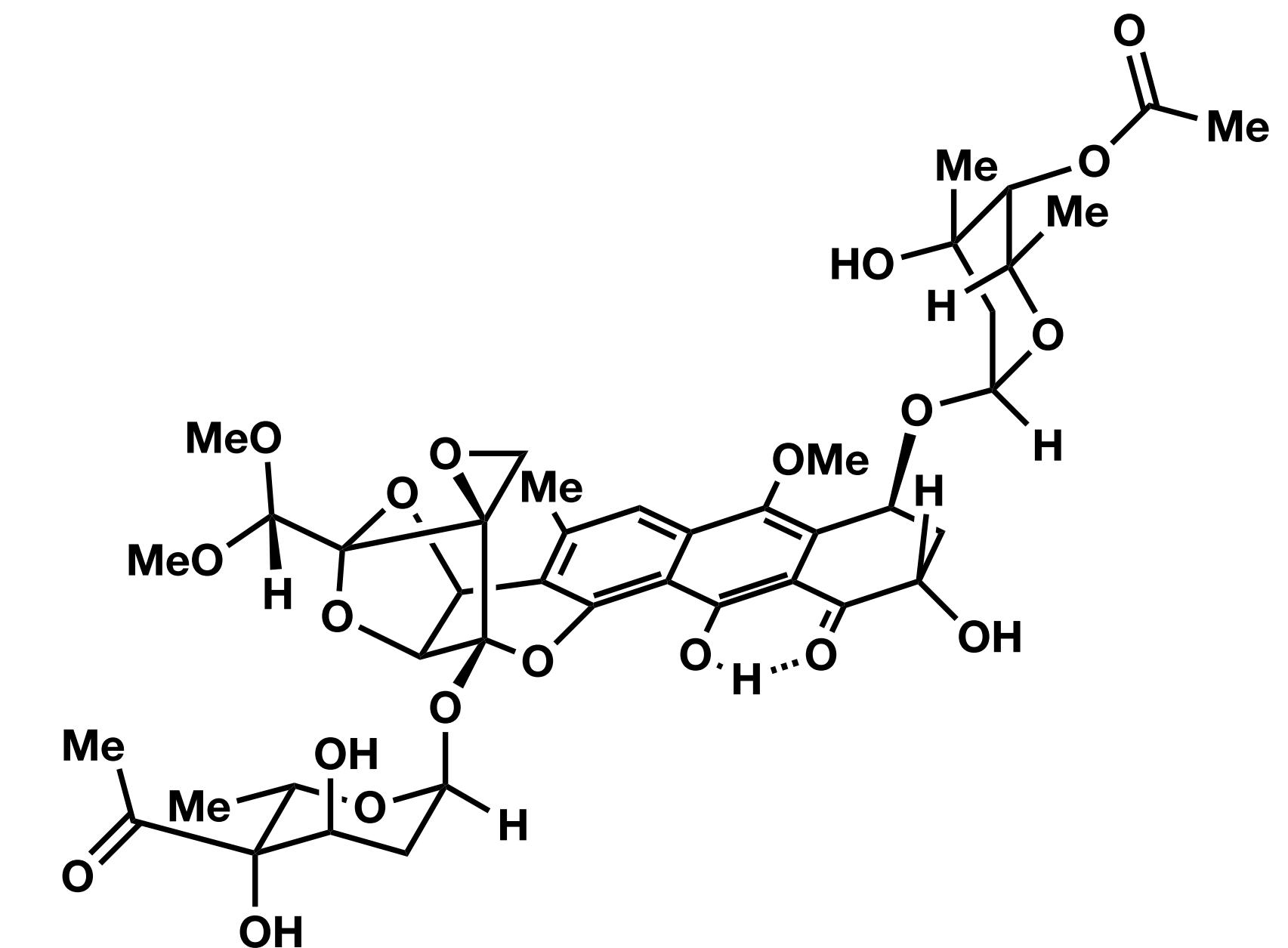
DC-45-A1



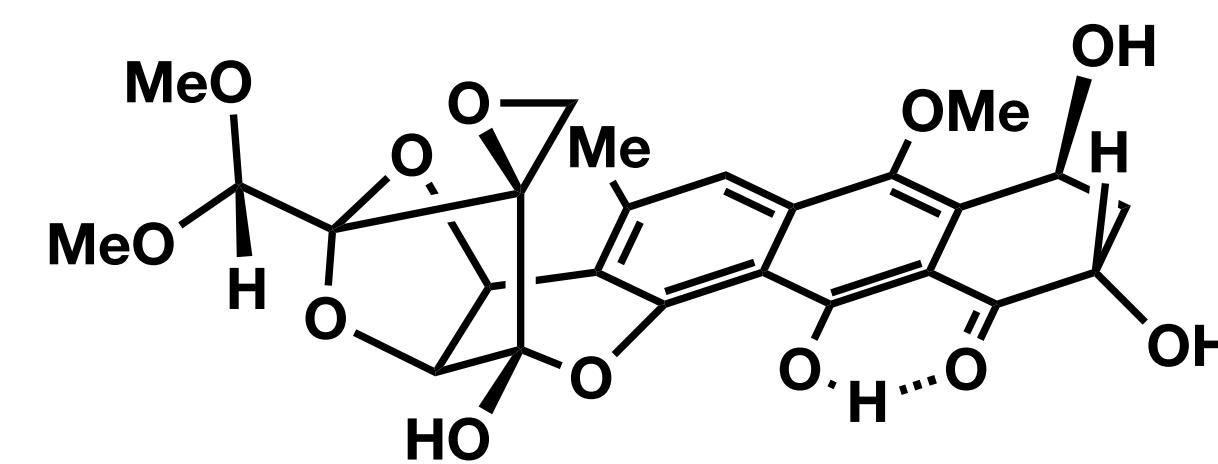
LL-D49194a1



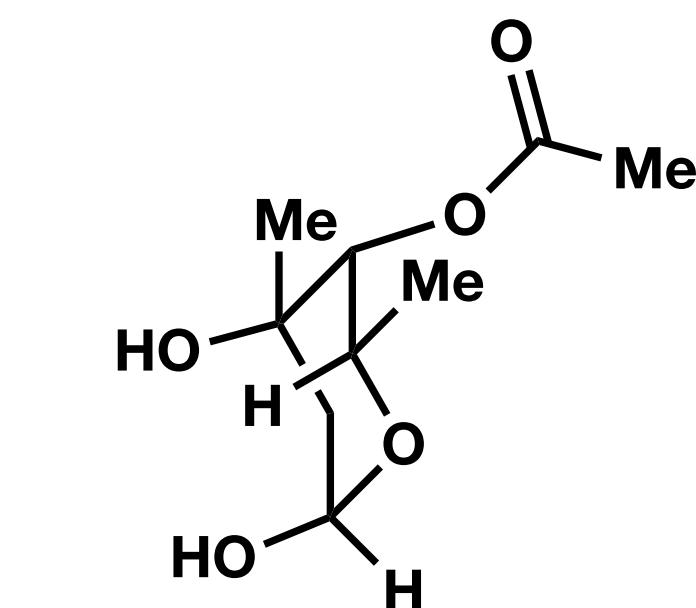
Trioxacarcin A



$\alpha$ -Trioxacarcinose B



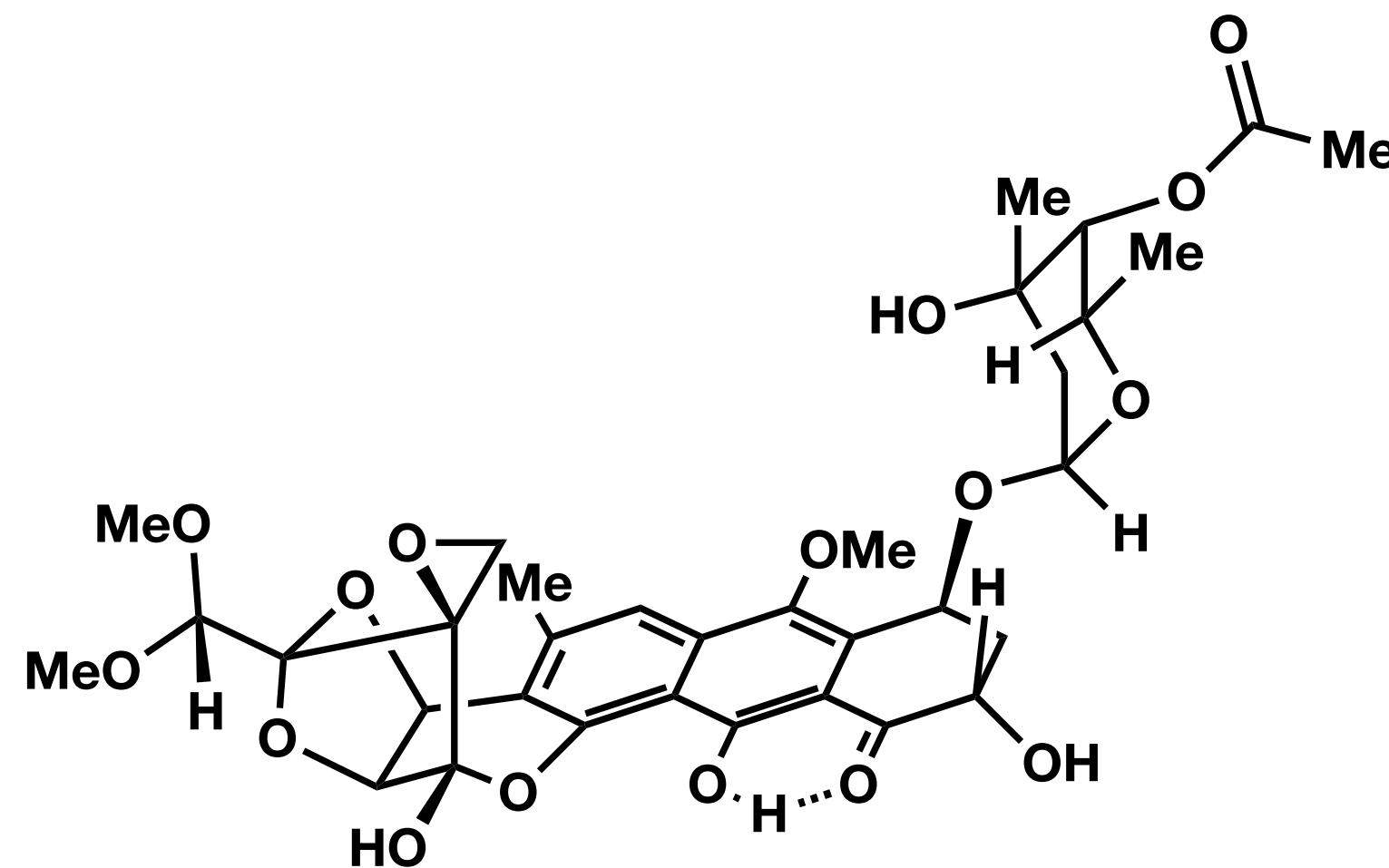
DC-45-A2



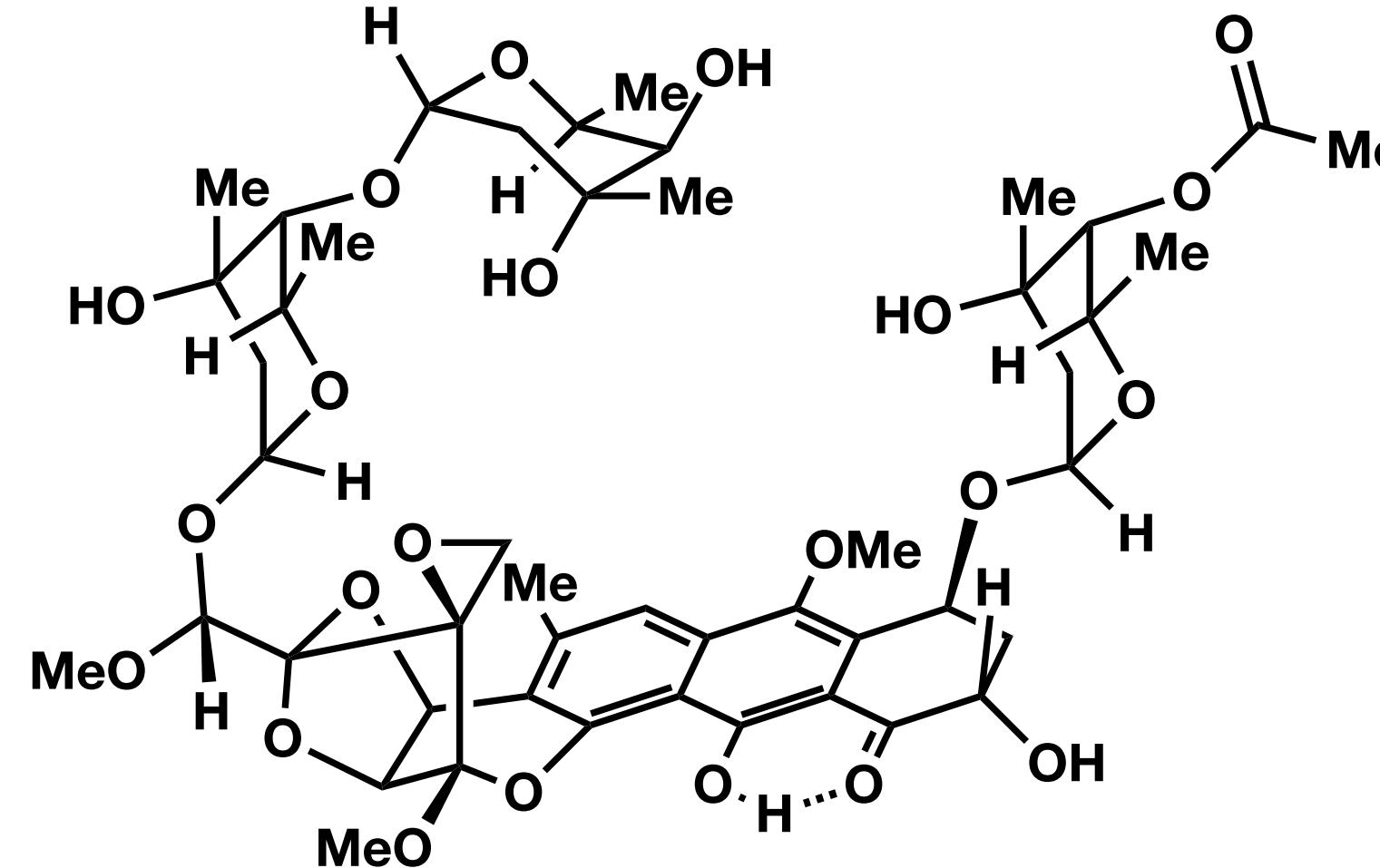
$\alpha$ -Trioxacarcinose A

# Trioxacarcins

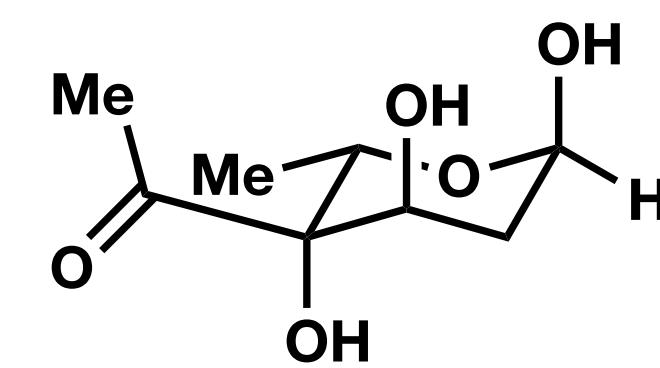
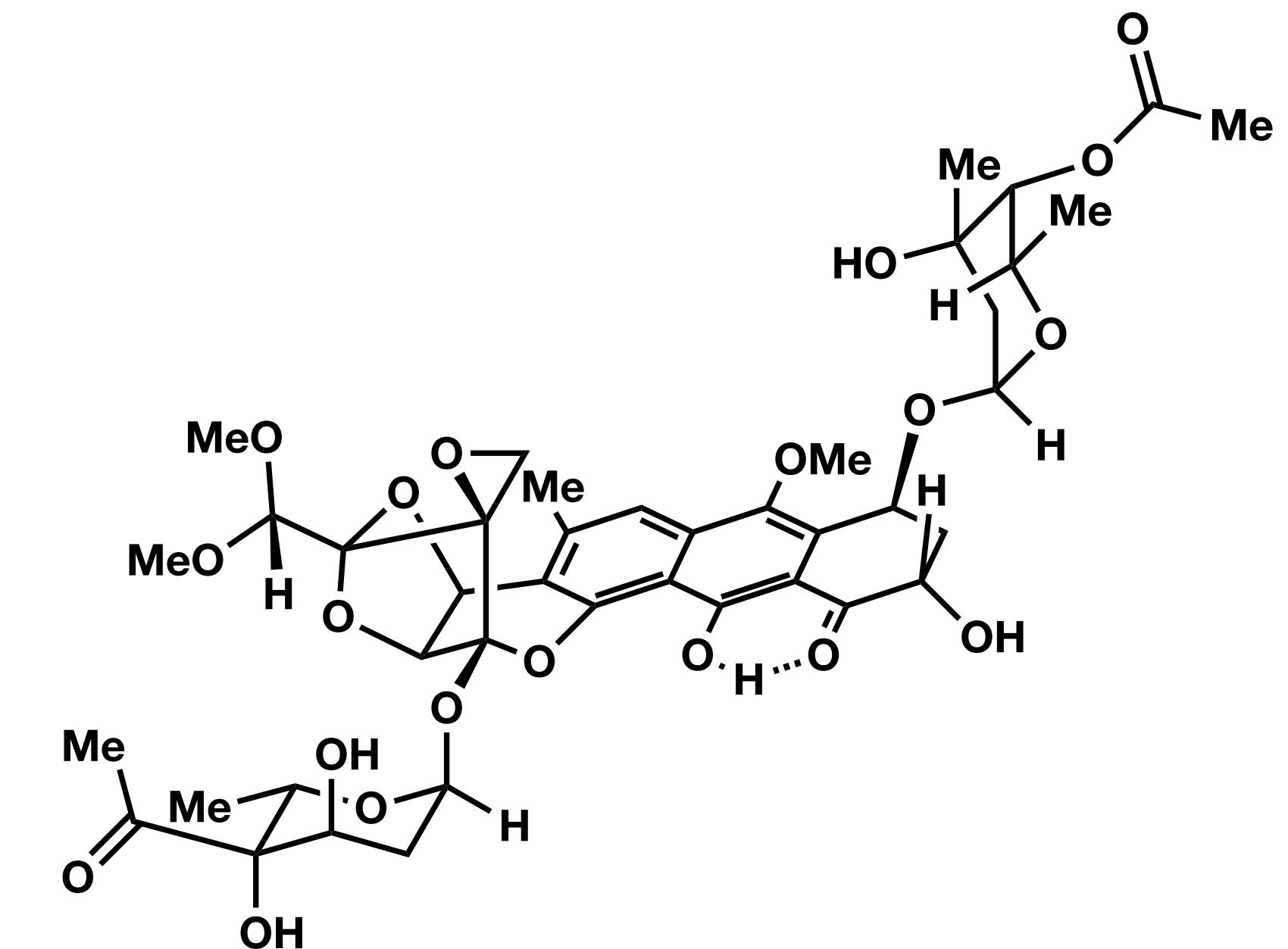
DC-45-A1



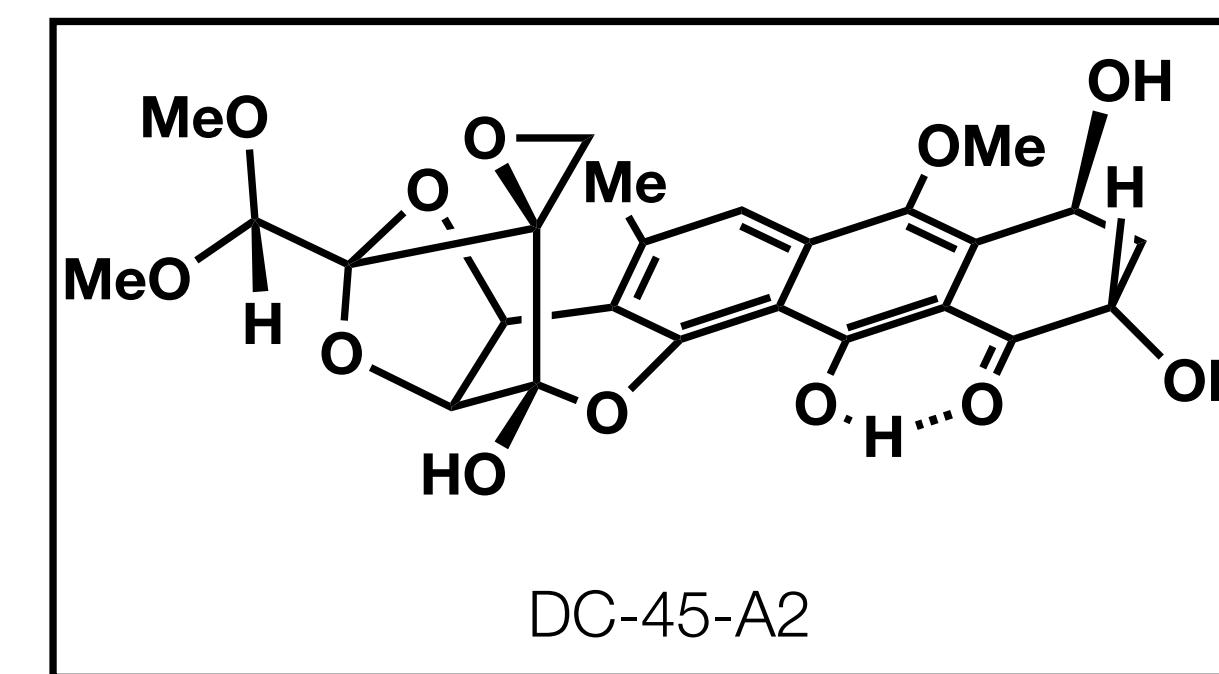
LL-D49194a1



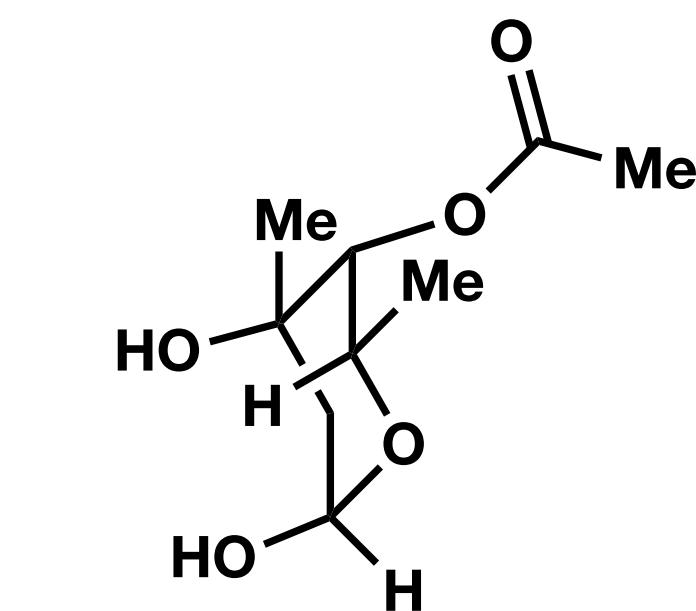
Trioxacarcin A



$\alpha$ -Trioxacarcinose B

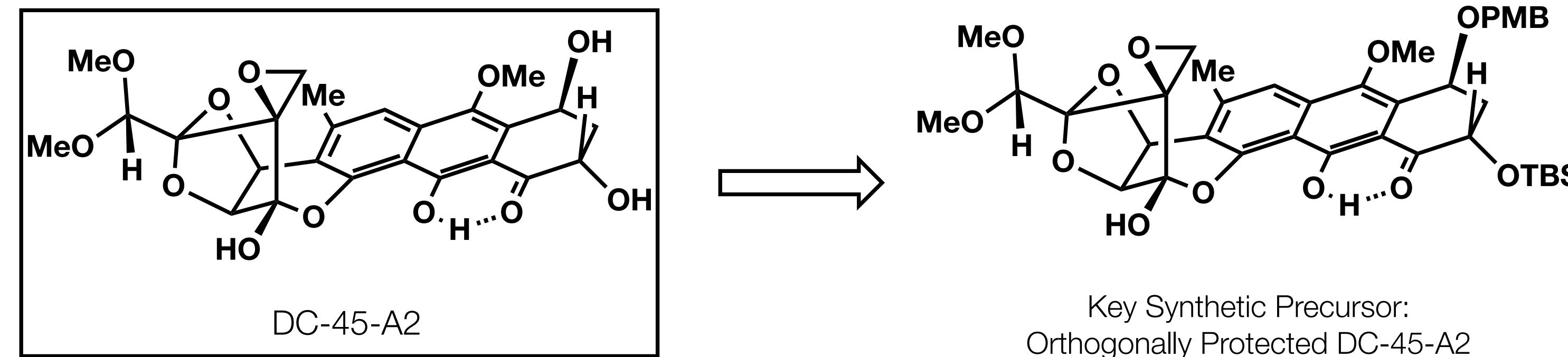


DC-45-A2

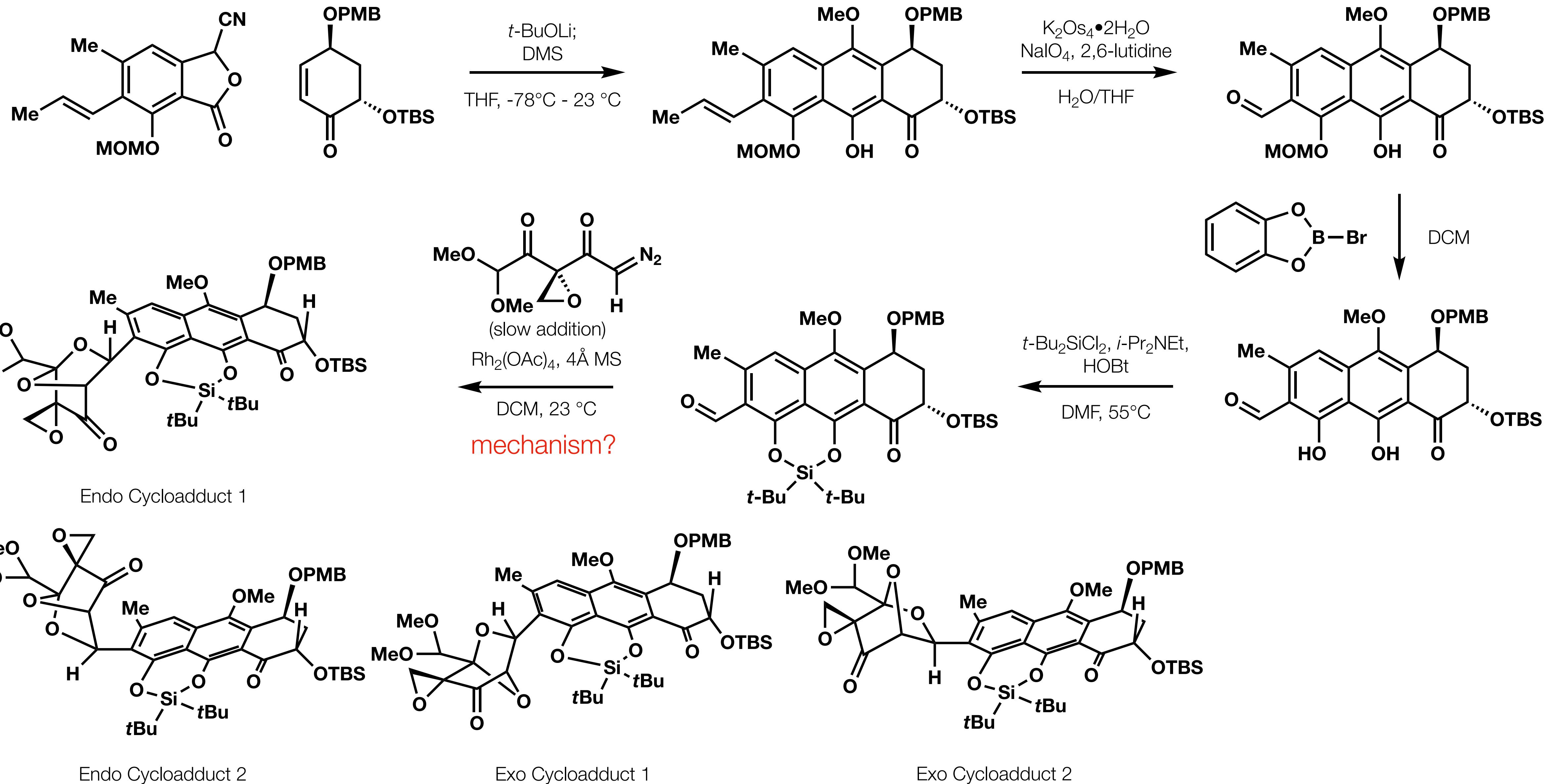


$\alpha$ -Trioxacarcinose A

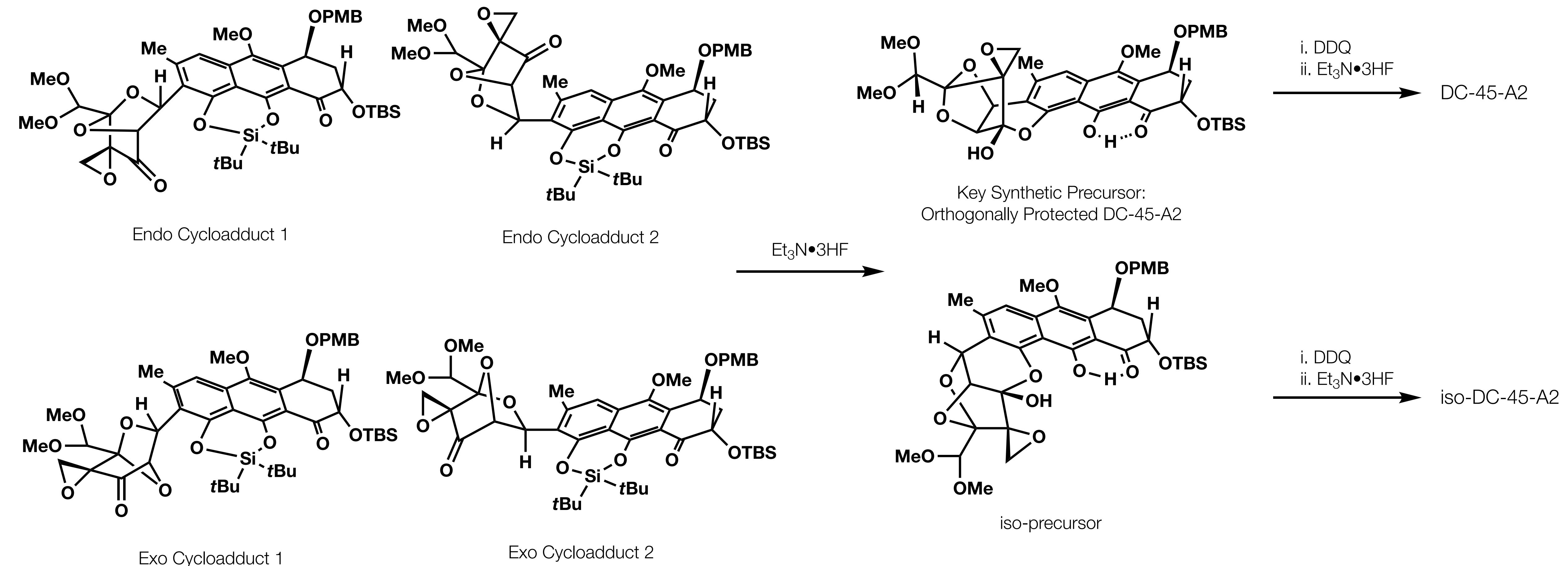
# Trioxacarcins



# Trioxacarcins



# Trioxacarcins



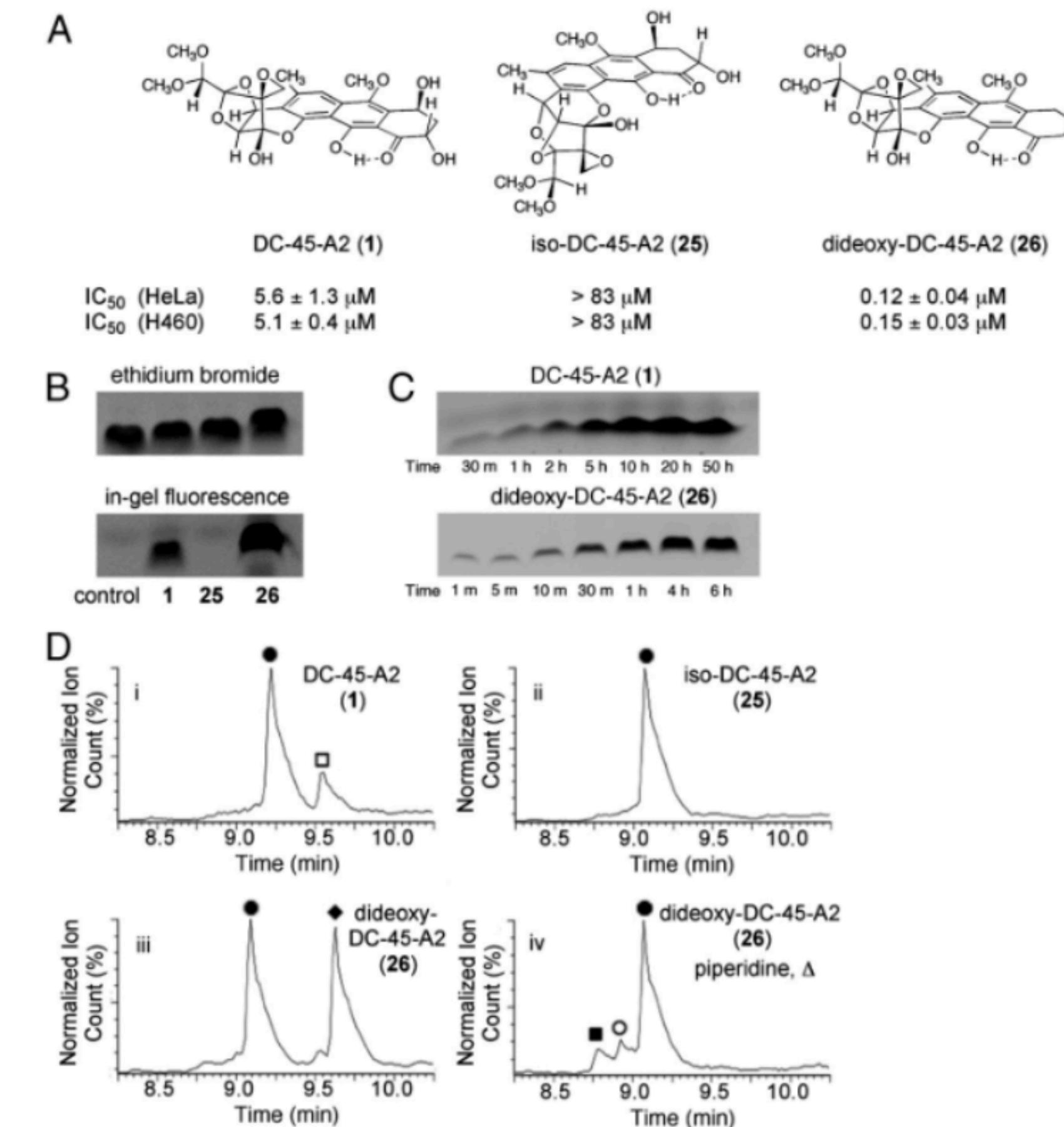
# Trioxacarcins

## Analogs of DC-A5-42

dideoxy-DC-A5-42 (**1**) is more potent than DC-45-A2 (**26**)

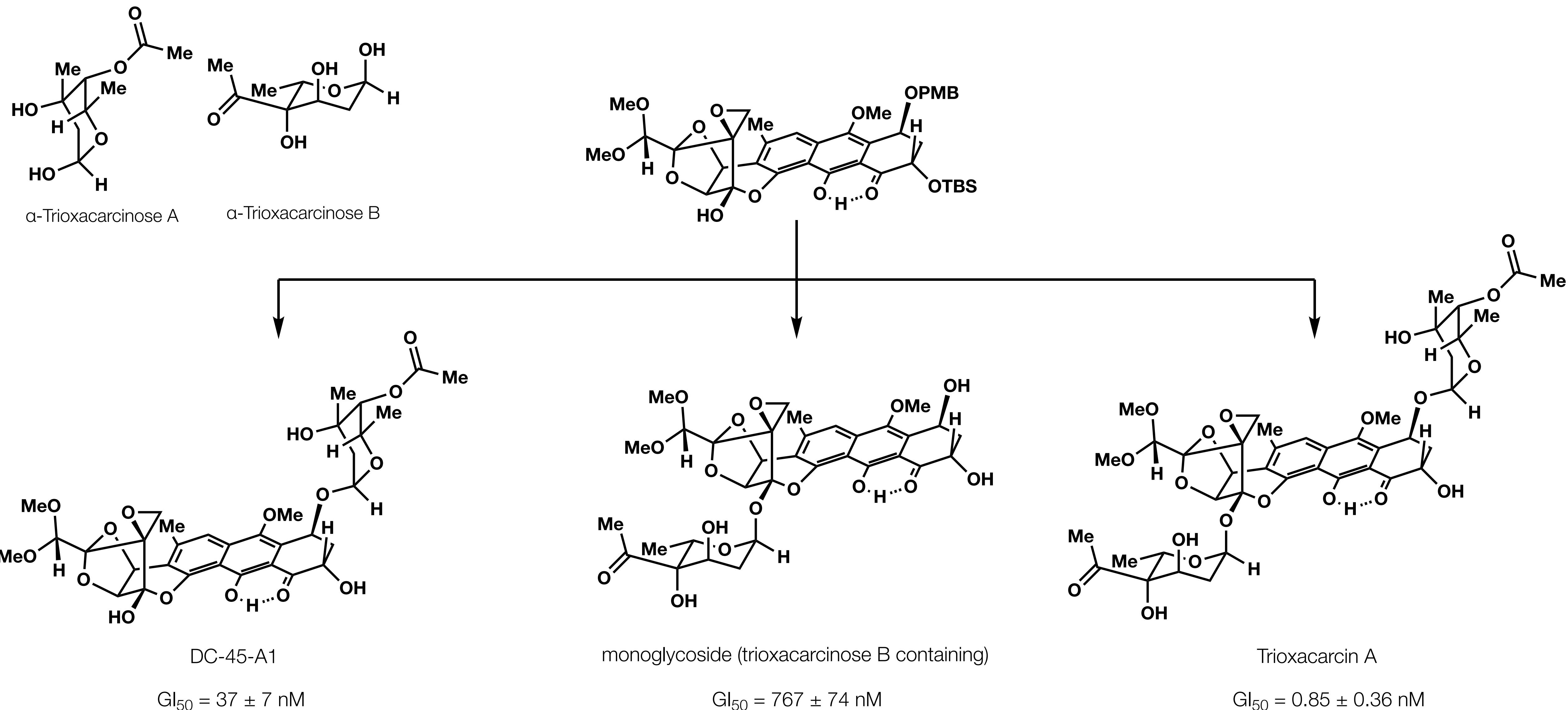
iso-DC-45-A2 (**25**) is inactive

**1** and **26** are shown to modify a short DNA duplex (12 base pairs) with a single guanine (G) in the center.

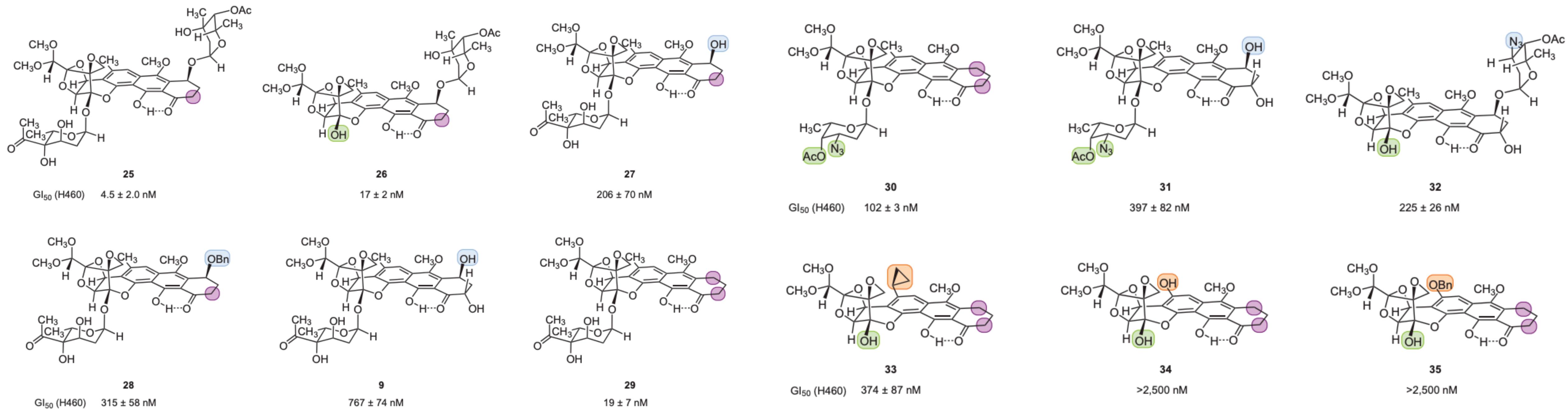


**Fig. 4.** Synthetic nonglycosylated trioxacarcins, their antiproliferative activities in cultured human cancer cells, and DNA-modifying effects. (A) IC<sub>50</sub> values for DC-45-A2, iso-DC-45-A2, and dideoxy-DC-45-A2 measured in HeLa (cervical cancer) and H460 (lung cancer) cell lines. (B) Images of TBE 20% polyacrylamide gels of the products of the reaction of the self-complementary duplex 12-mer d(AATTACGTAATT) (100 μM) with DC-45-A2 (lane 2, 100 μM), iso-DC-45-A2 (lane 3, 100 μM), or dideoxy-DC-45-A2 (lane 4, 100 μM) for 2 h at 23 °C; visualized with ethidium bromide and by in-gel fluorescence. (C) Images of TBE gels of the products of the reaction of the self-complementary duplex 12-mer d(AATTACGTAATT) (100 μM) with DC-45-A2 or dideoxy-DC-45-A2 (25 μM) at 23 °C for the indicated times; visualized by in-gel fluorescence. (D) LC-MS chromatograms of reaction mixtures of the self-complementary duplex 12-mer d(AATTACGTAATT) (100 μM) and (i) DC-45-A2 (100 μM), (ii) iso-DC-45-A2 (100 μM), or (iii) dideoxy-DC-45-A2 (100 μM) after 24 h at 23 °C; iv depicts the LC-MS chromatogram of the reaction mixture of the self-complementary duplex 12-mer d(AATTACGTAATT) (100 μM) and dideoxy-DC-45-A2 (100 μM) after 24 h at 23 °C followed by the addition of piperidine (1 M) and heating for 30 min at 95 °C; ●, (AATTACGTAATT); □, (AATTACGTAATT • 1); ♦, (AATTACGTAATT • 26); ■, (pTAATT); ○, (AATTACp).

# Trioxacarcins - Glycosydic Analogs

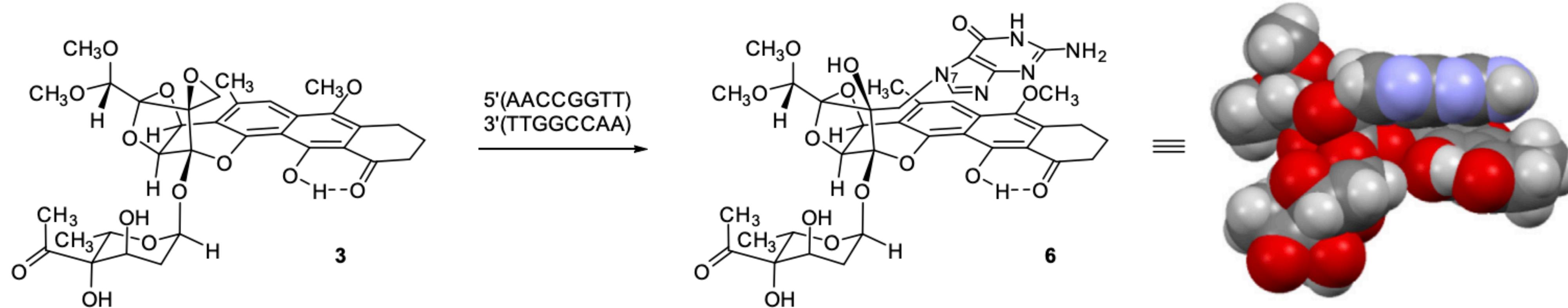


# Trioxacarcins - Glycosyidic Analogs

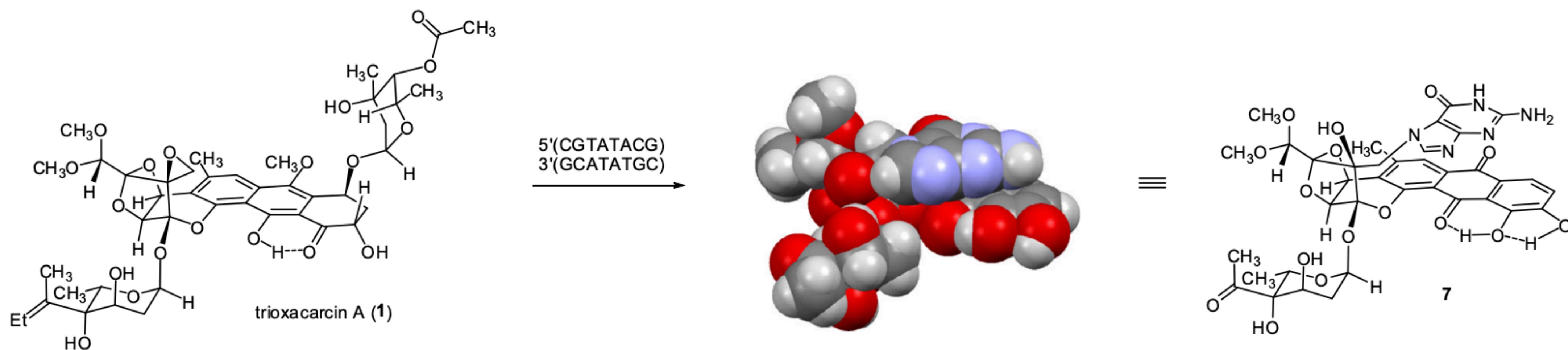


No SAR discussed, but synthetic strategy is clearly effective at generating a variety of analogs

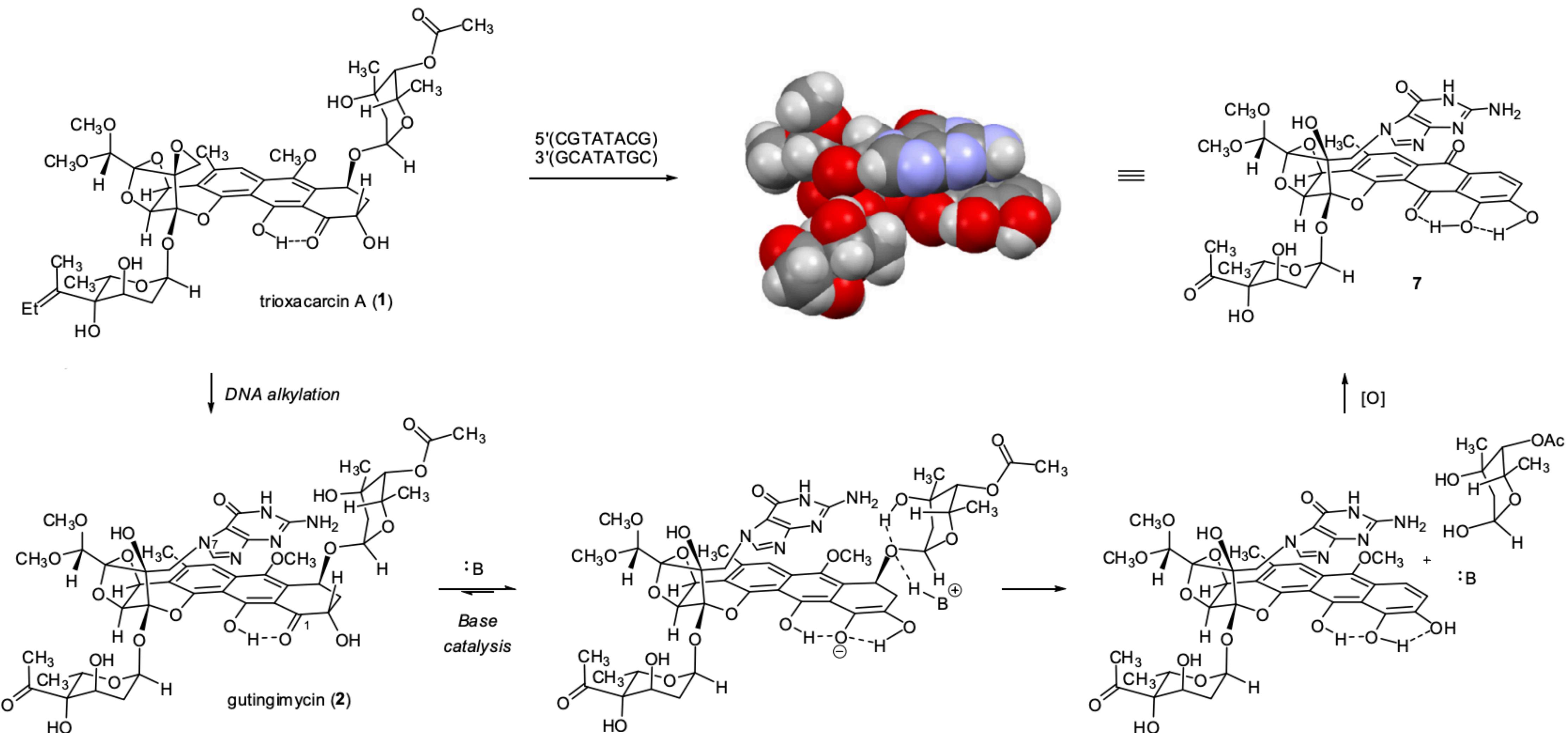
# Trioxacarcins - Evidence for Mechanism of Action



**Scheme 1.** Formation of guanine adduct **6** from the fully synthetic trioxacarcin analog **3**.

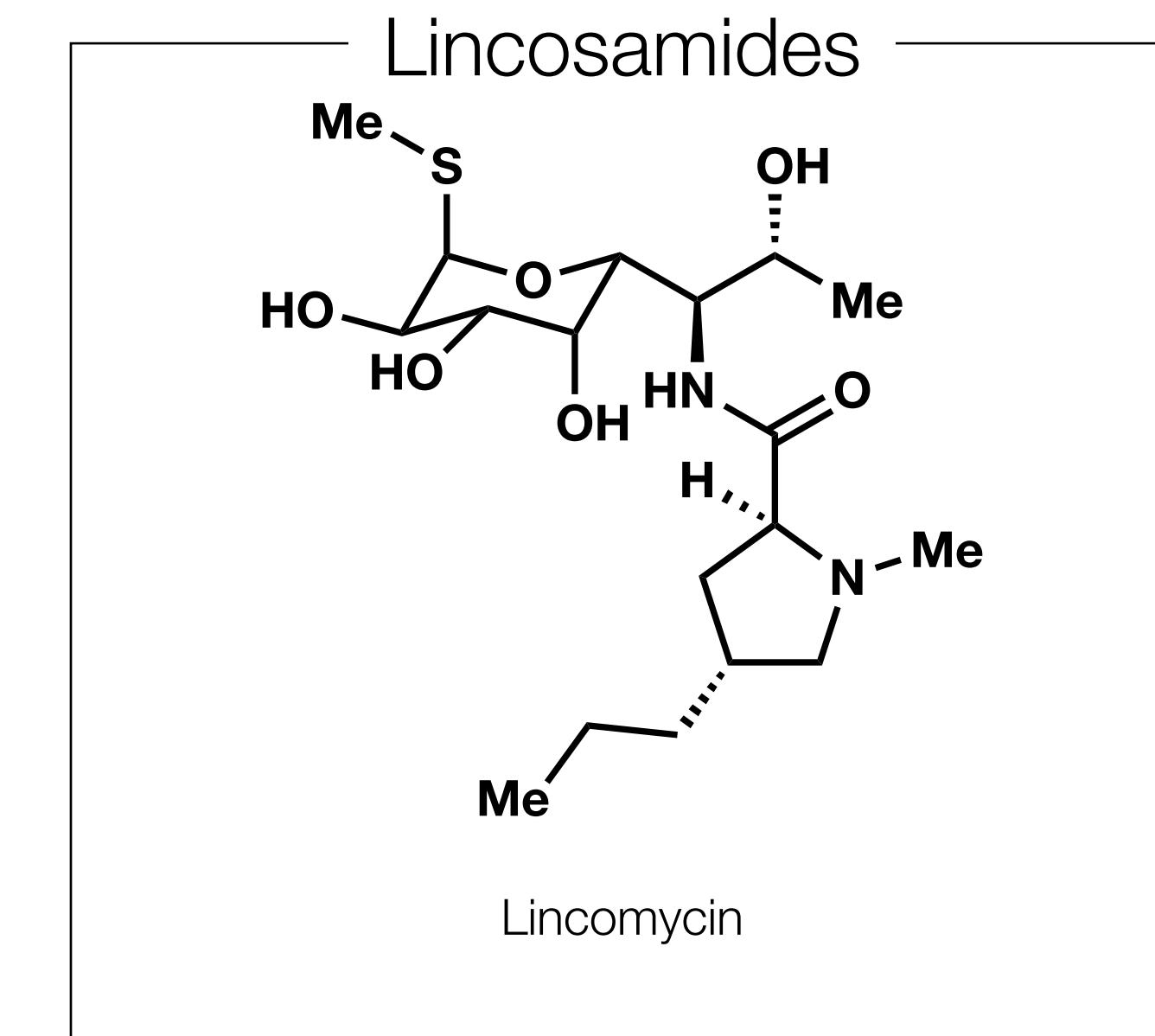
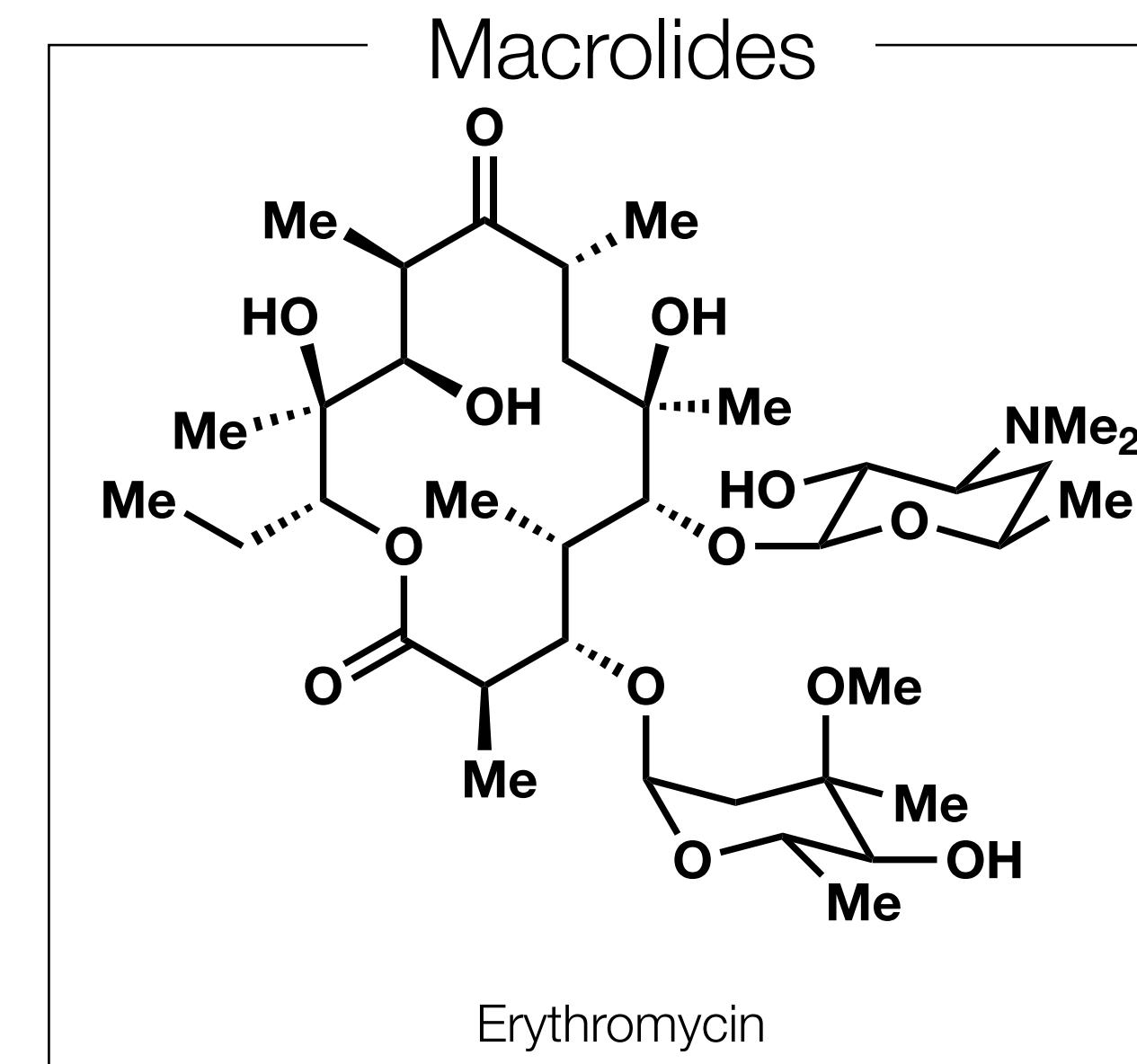
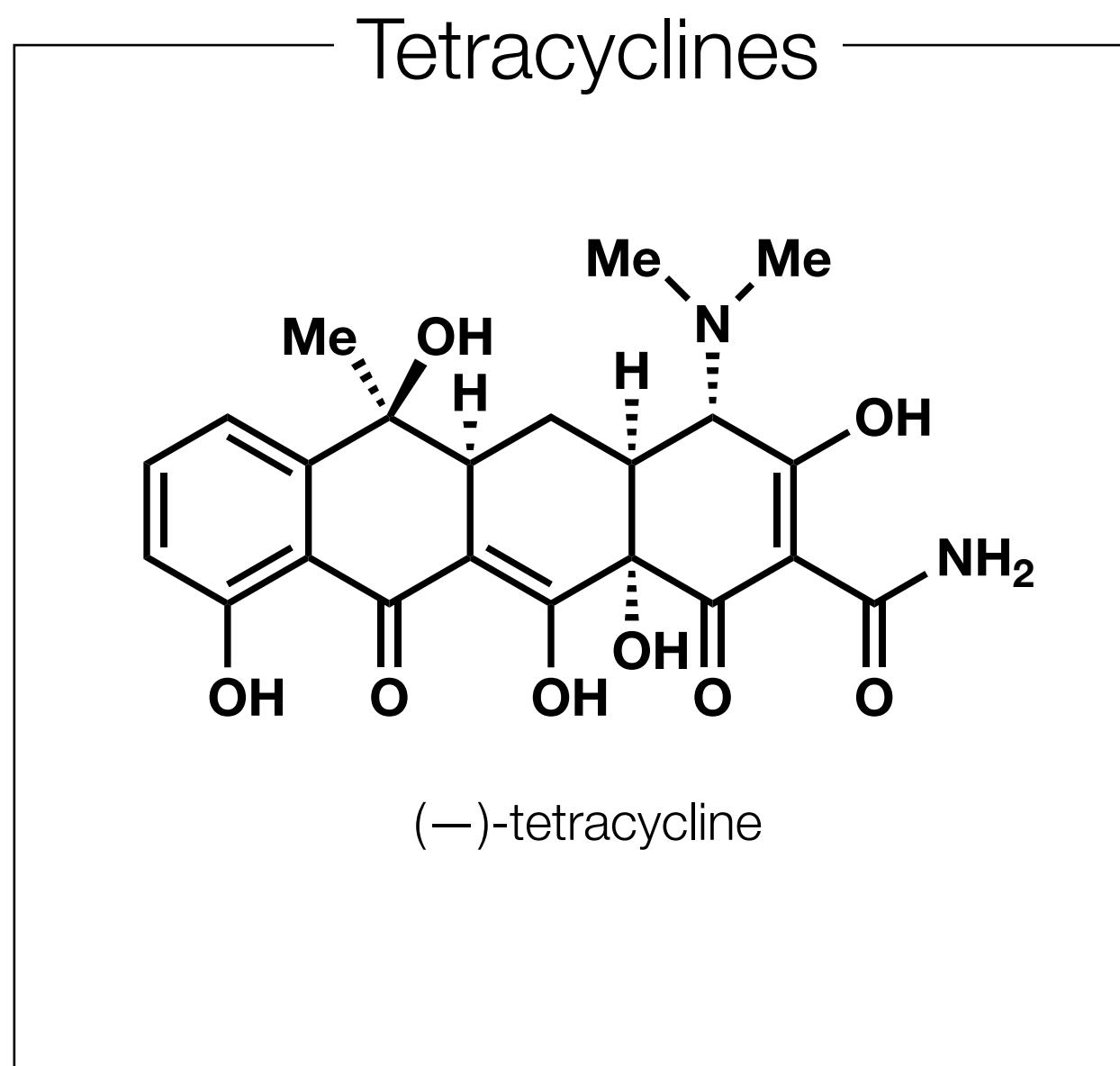


# Trioxacarcins - Evidence for Mechanism of Action



# Combatting Antibiotic Resistance

*Perhaps Myers' Most Impactful Work*



Overarching Research Theme

Convergent Routes with High Modularity



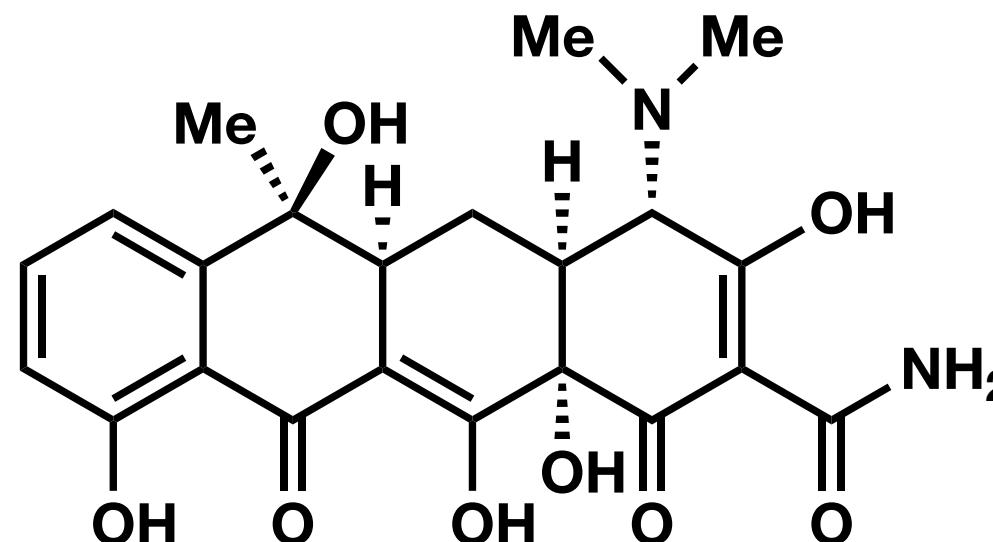
SAR Exploration



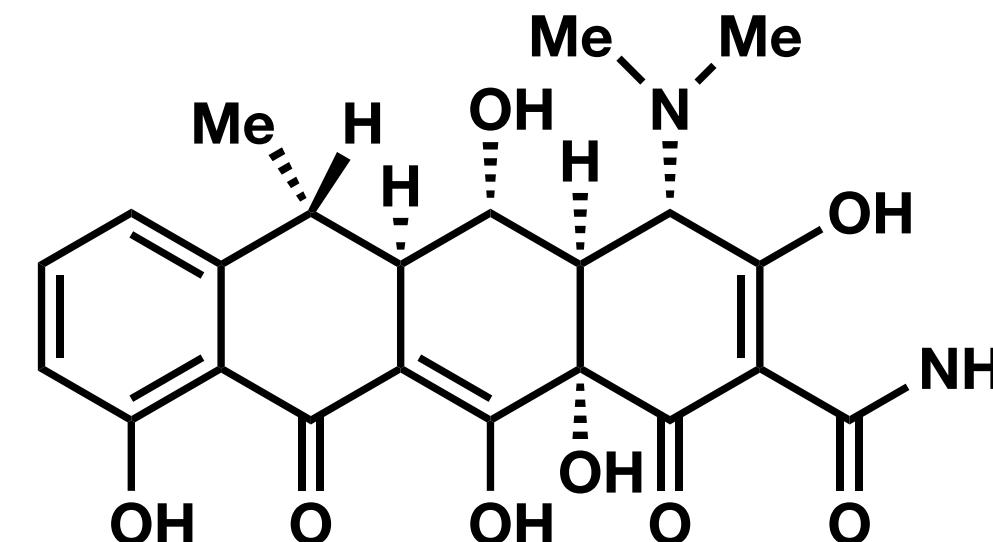
Identification of Clinically Relevant Targets

# Tetracyclines

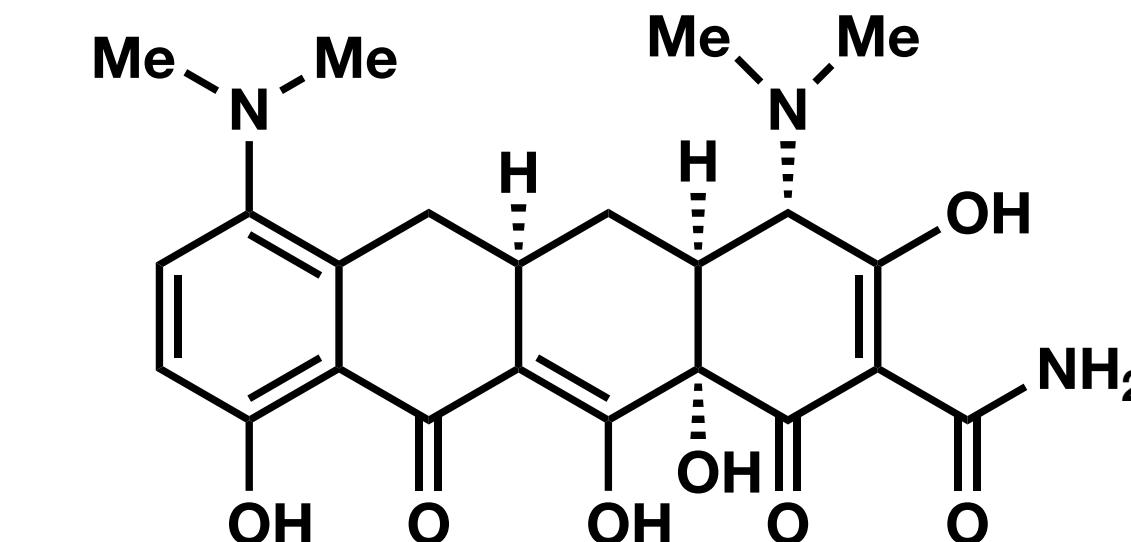
Widely prescribed class of antibiotics since the 1950s



(-)-tetracycline



(-)-doxycycline



(-)-minocycline

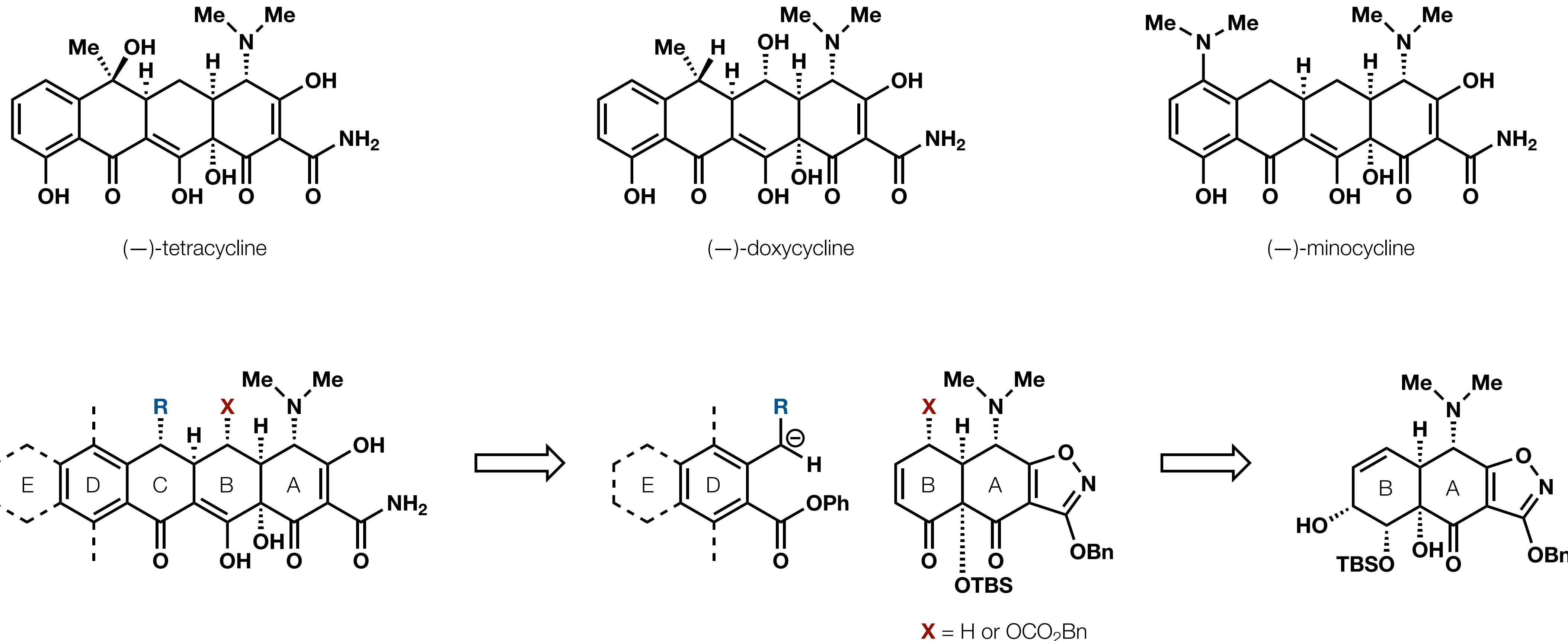
natural product produced on large scale by fermentation

both are non-natural antibiotics and are manufactured by semisynthesis

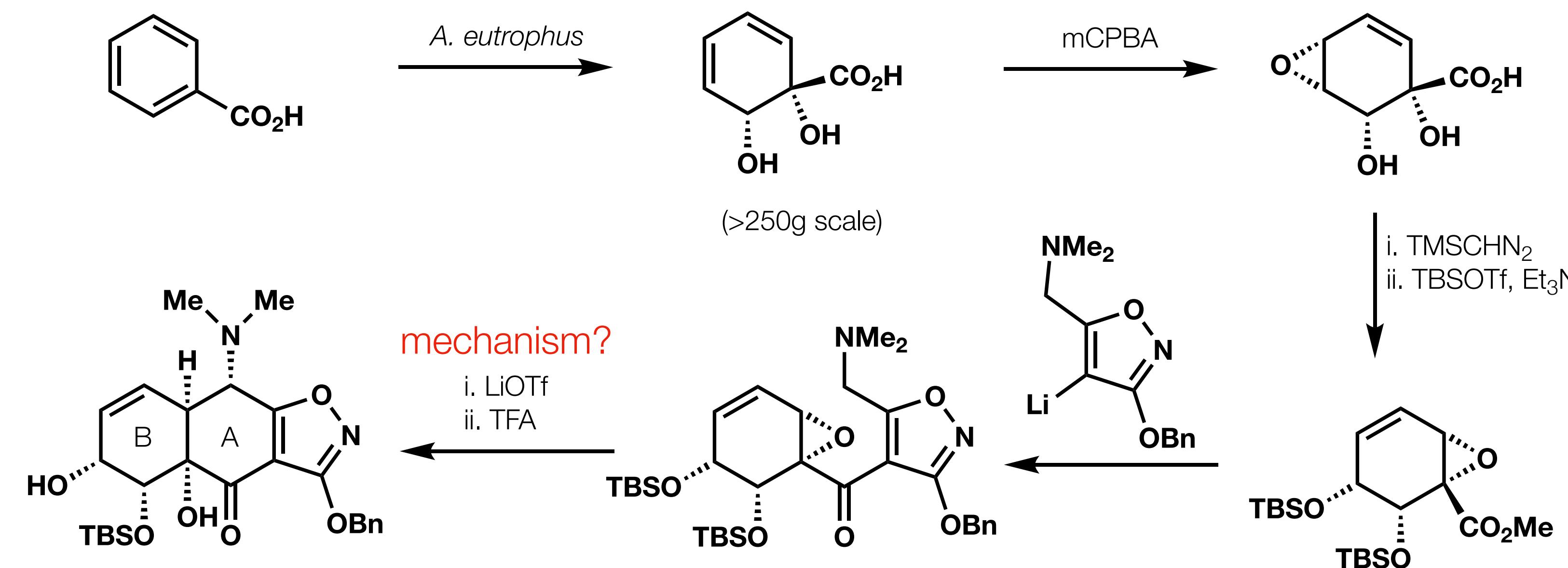
Commonly prescribed for

- respiratory infections
- STIs
- UTIs
- Lyme disease
- Other systematic bacterial infections
- acne
- rosacea

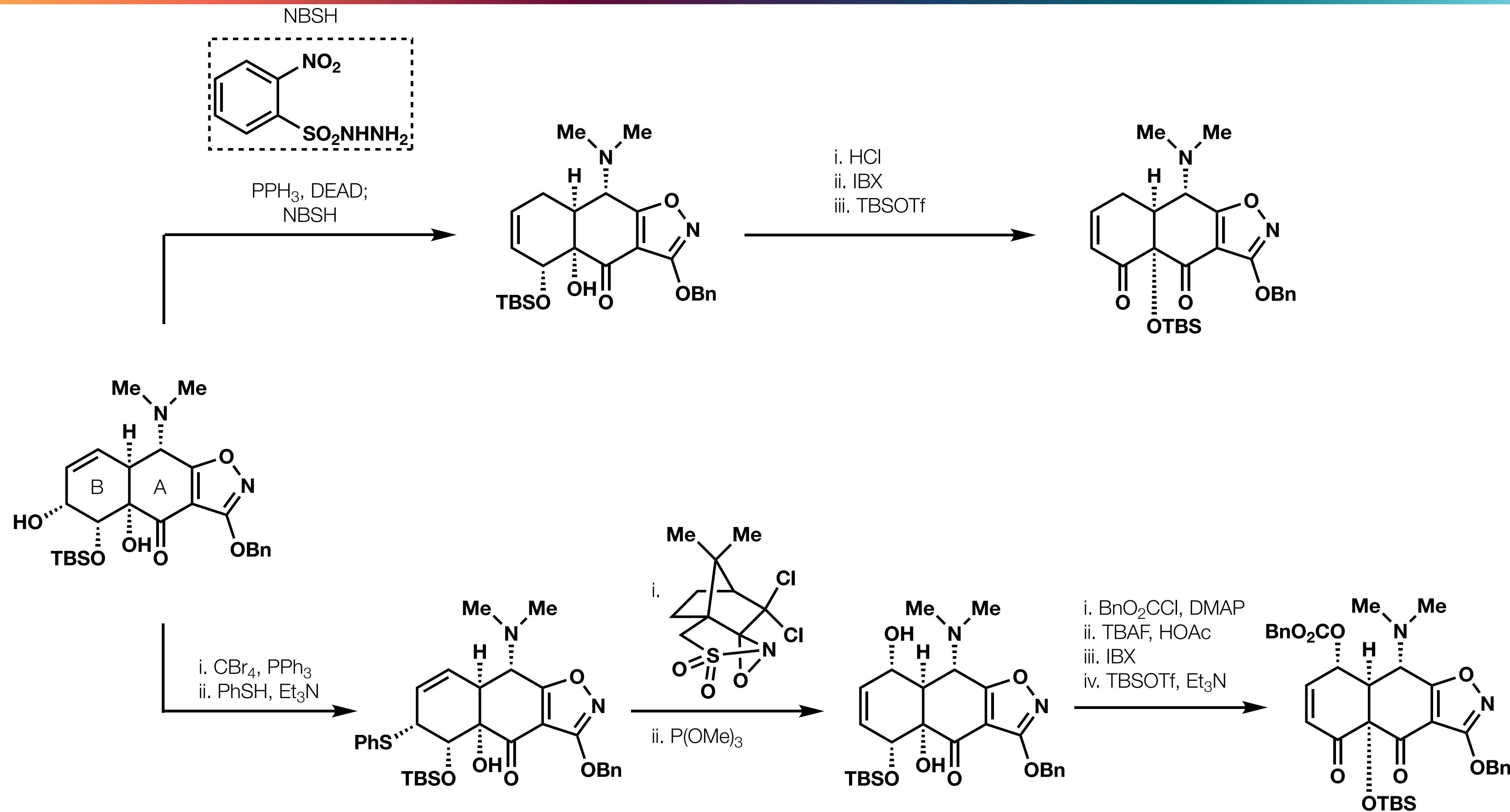
# Tetracyclines - Convergent Synthesis



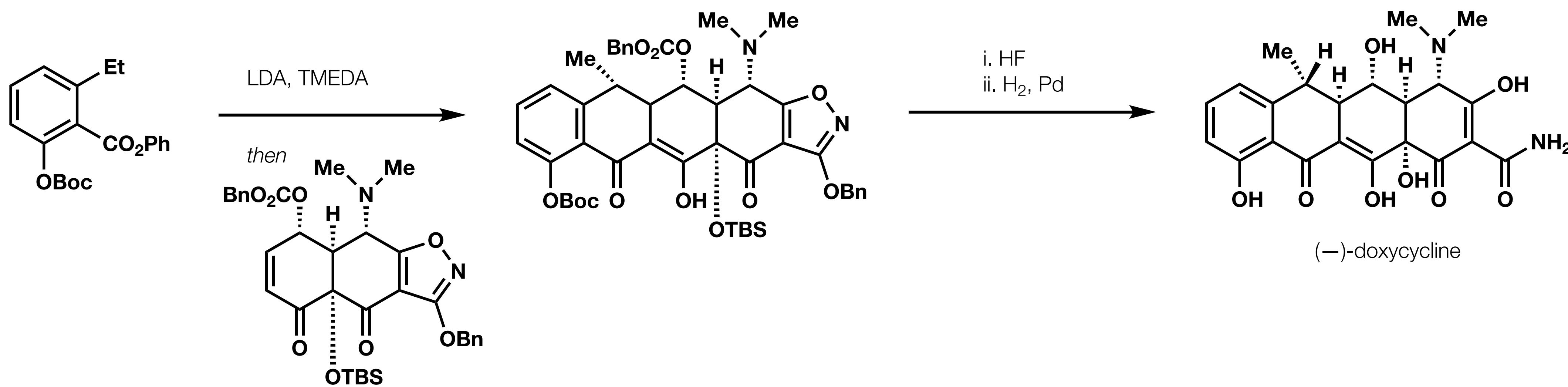
# Tetracyclines - Convergent Synthesis



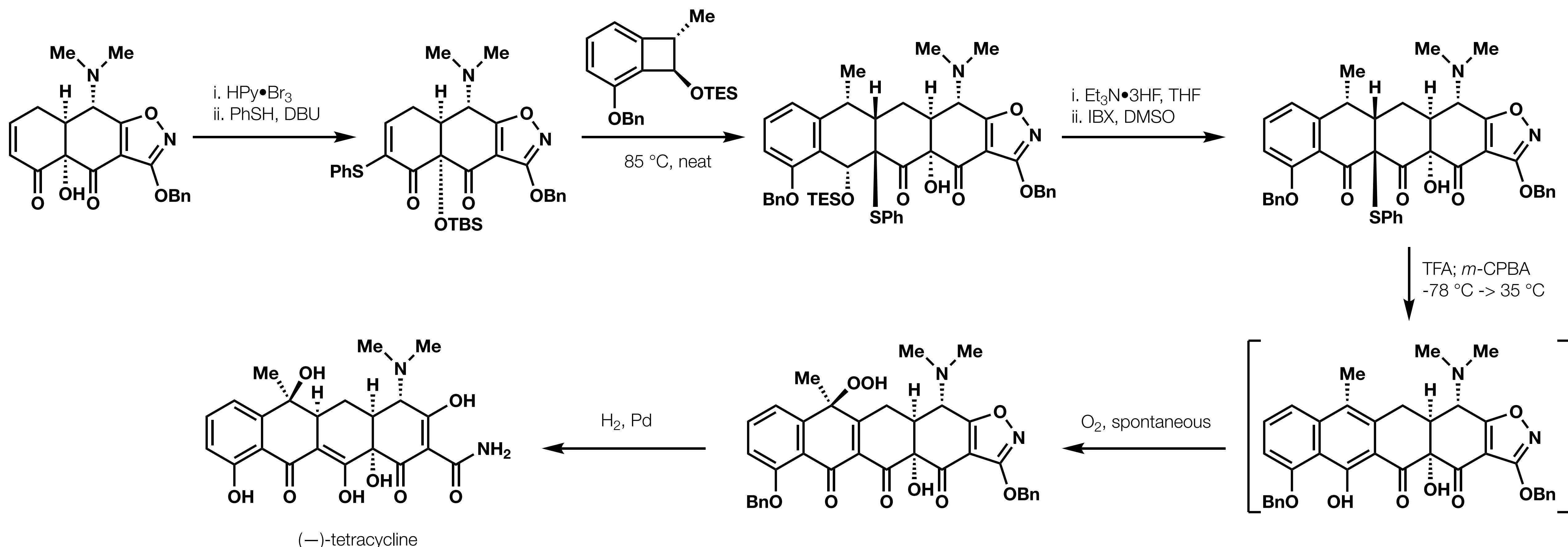
# Tetracyclines - Convergent Synthesis



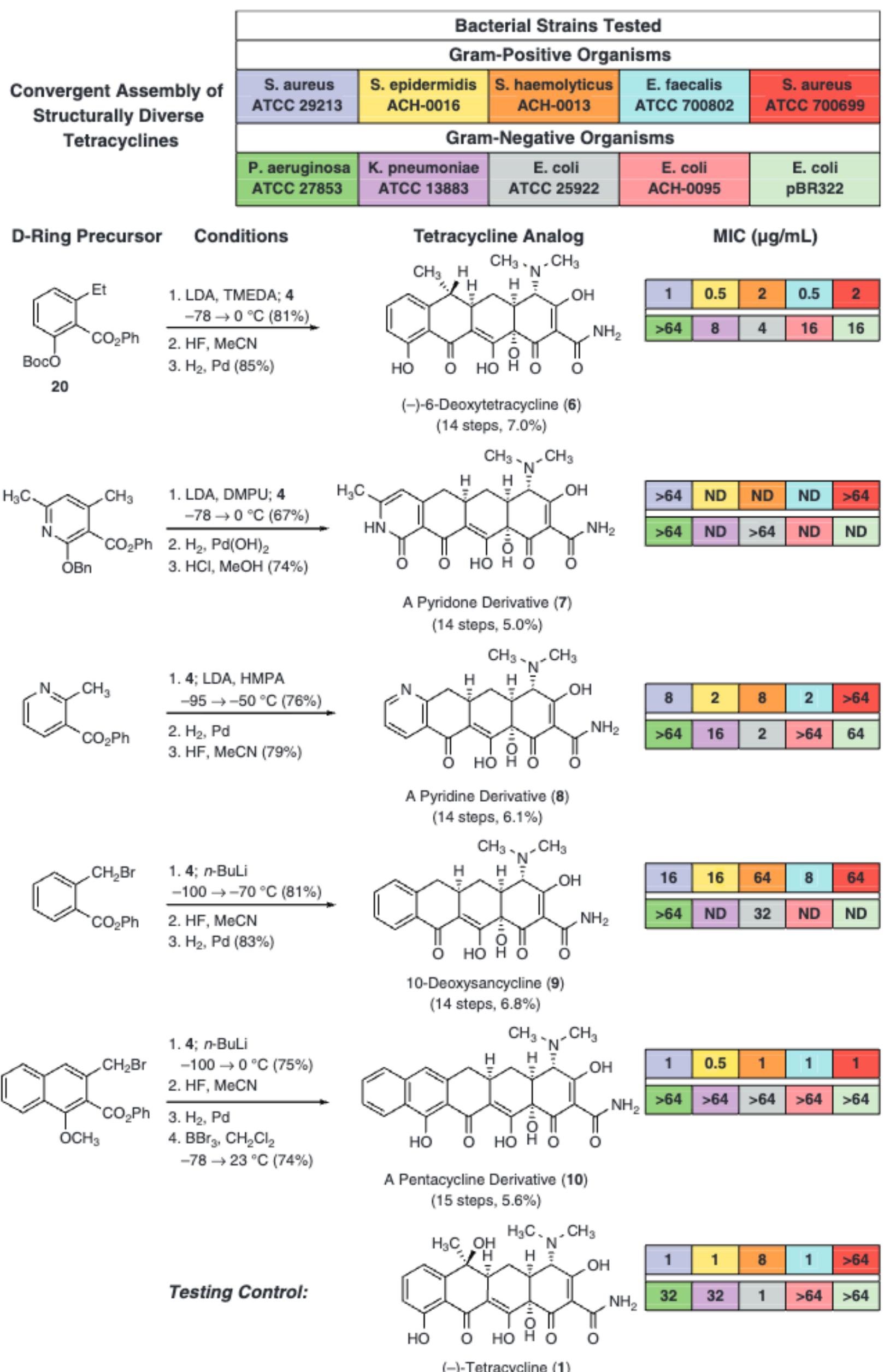
# Tetracyclines - Convergent Synthesis



# Tetracyclines - Convergent Synthesis

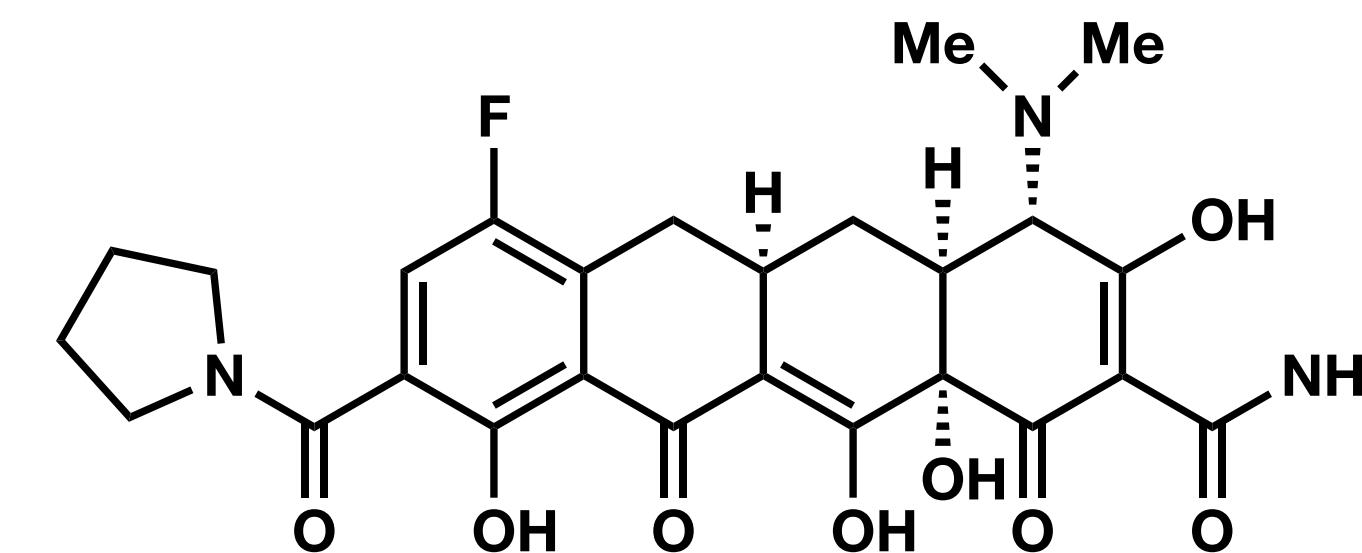


# Tetracyclines - Convergent Synthesis



Initiated a platform that allowed for the synthesis of more than 3000 tetracycline analogs

# Identification of Most Potent Analog



TP-434 (eravacycline)

Systematic exploration of analogs led to identification of TP-434 as a lead target

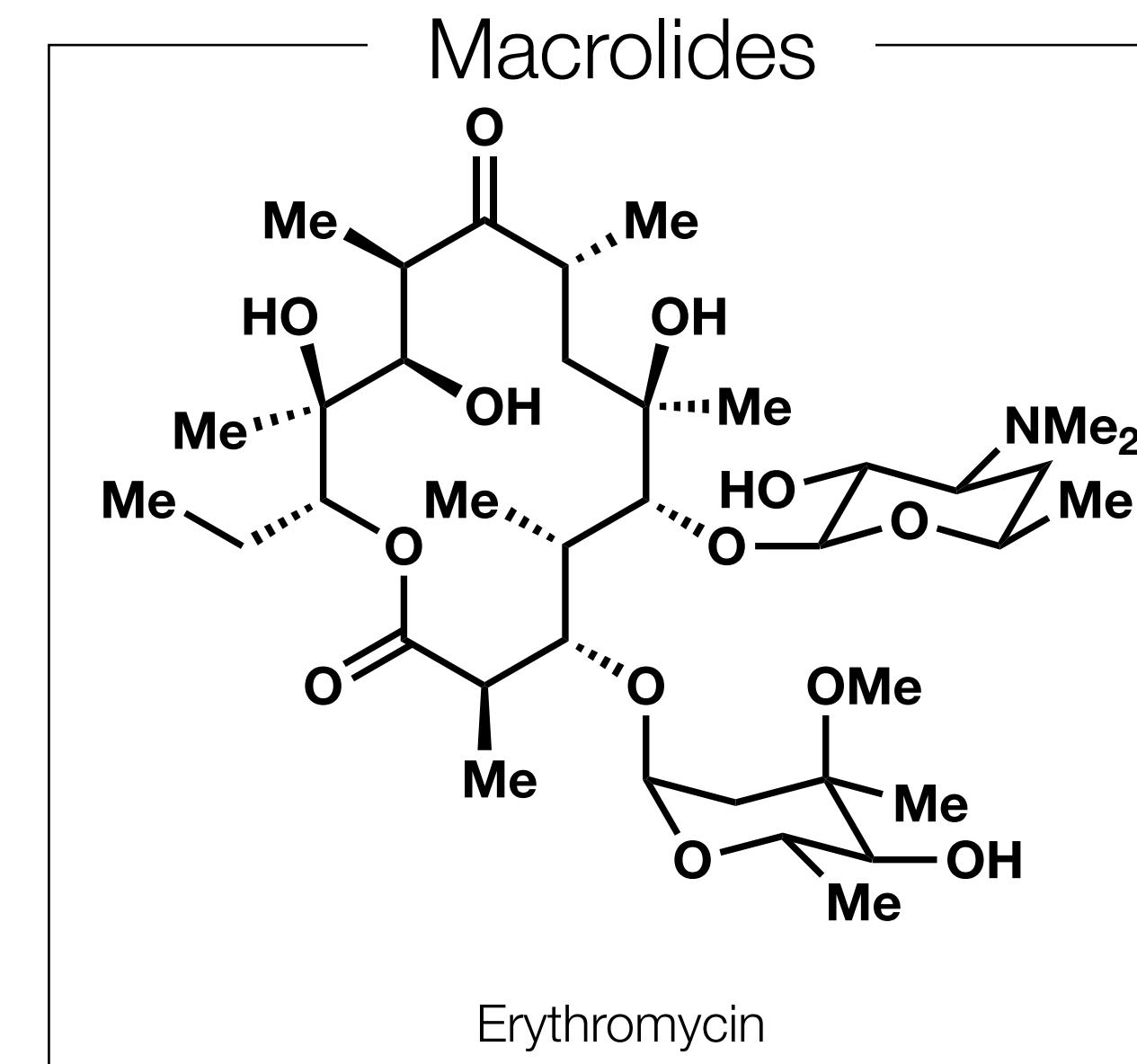
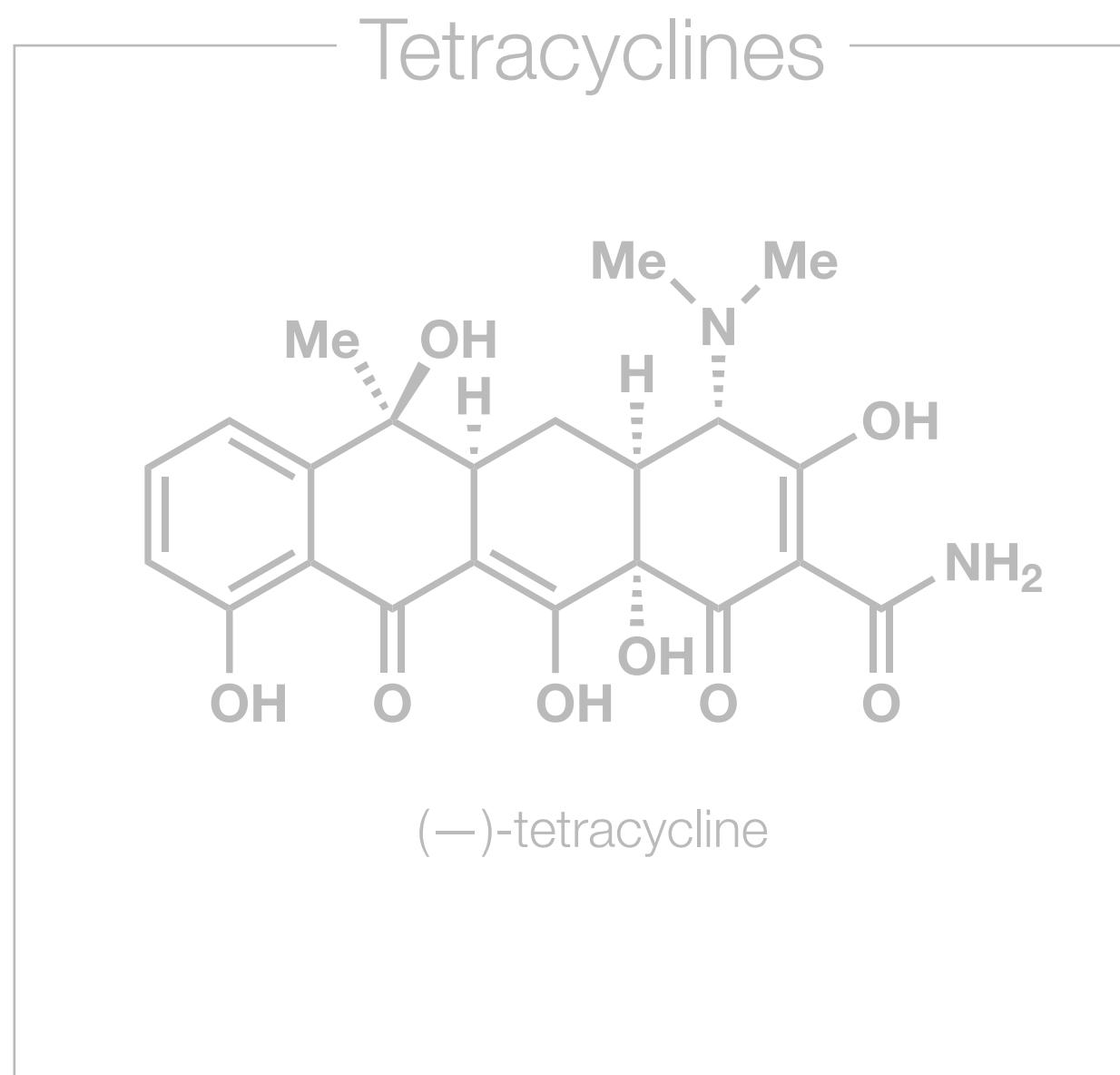
MIC<sub>90</sub> values of  $\leq 2 \mu\text{g/ml}$  against panels of all major bacterial species (Both Gram-positive and Gram-negative) except for *Pseudomonas aeruginosa* and *Burkholderia cenocepacia*

Tetraphase Pharmaceuticals (founded by Myers) began advancing TP-434 in preclinical development in 2008

FDA approved eravacycline (Xerava) for complicated intra-abdominal infections in 2018

# Combatting Antibiotic Resistance

*Perhaps Myers' Most Impactful Work*



Overarching Research Theme

Convergent Routes with High Modularity



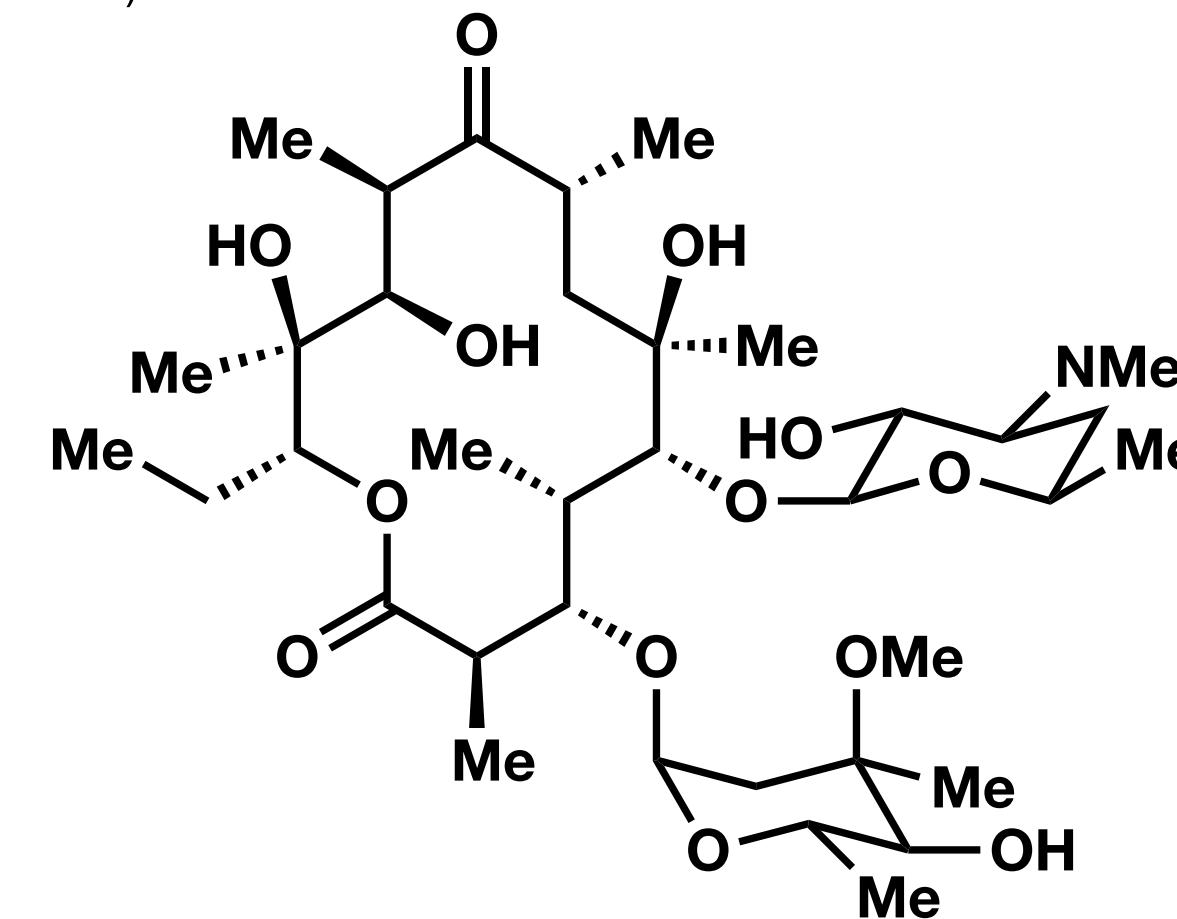
SAR Exploration



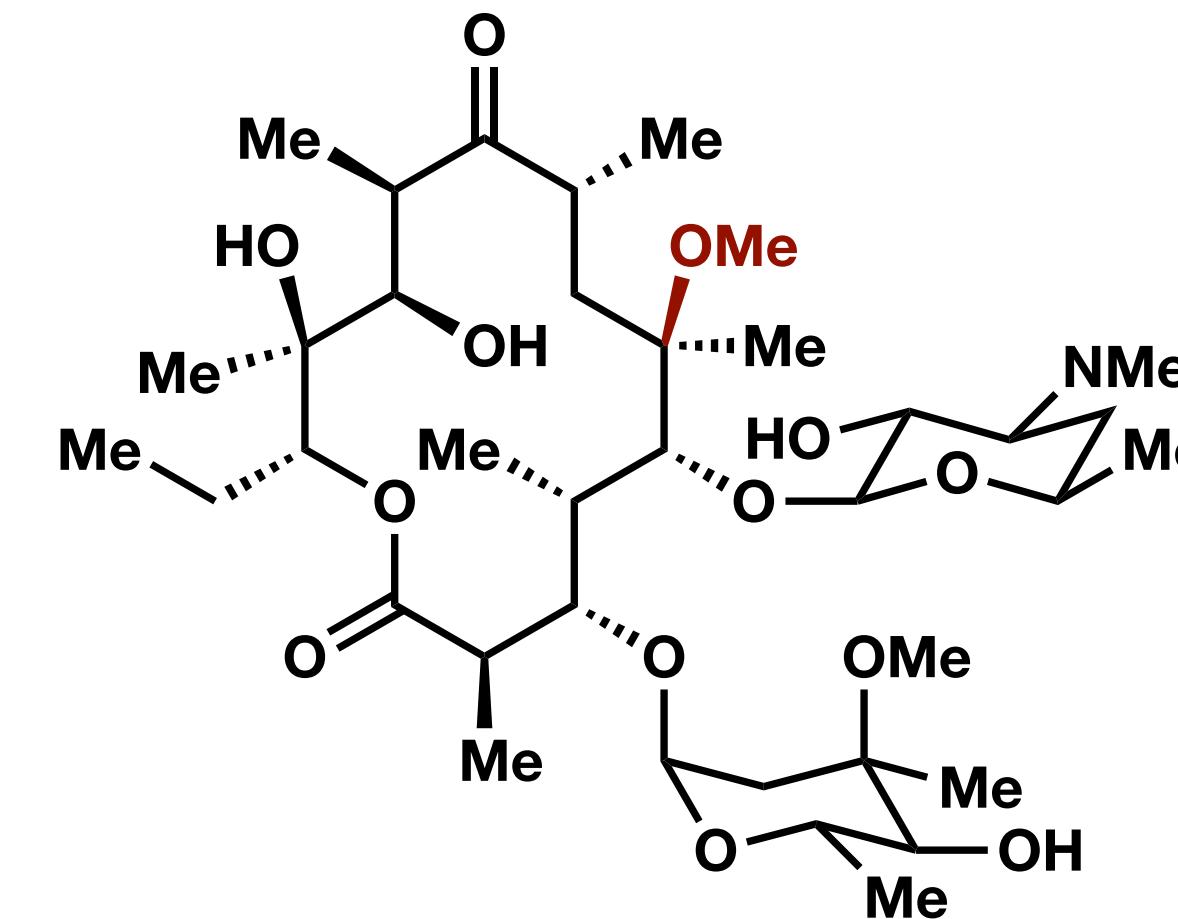
Identification of Clinically Relevant Targets

# Macrolides - Dependency on Semisynthesis

*Nature*. **2016**: 533(7603): 338–345.

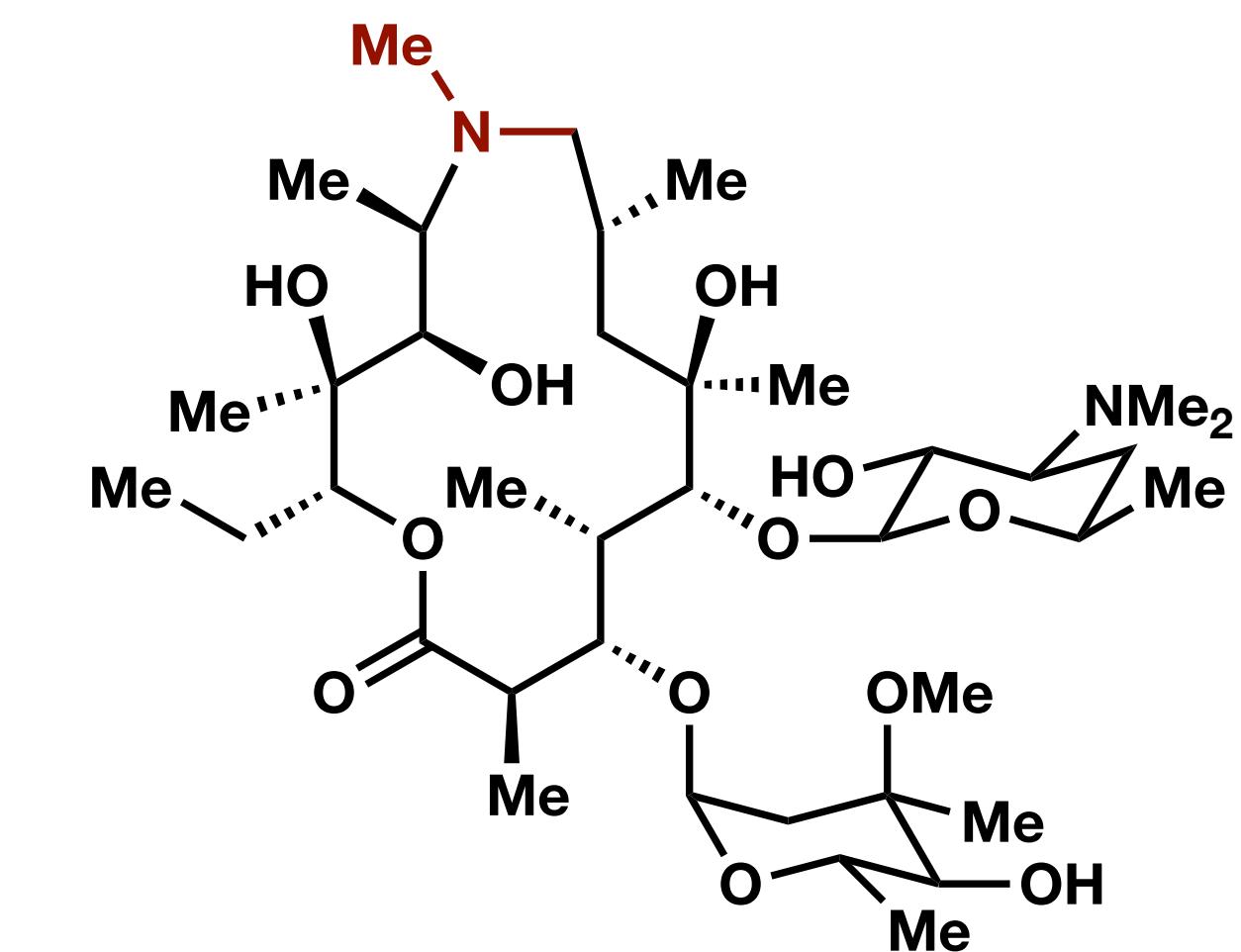


# Erythromycin (Fermentation product 1952

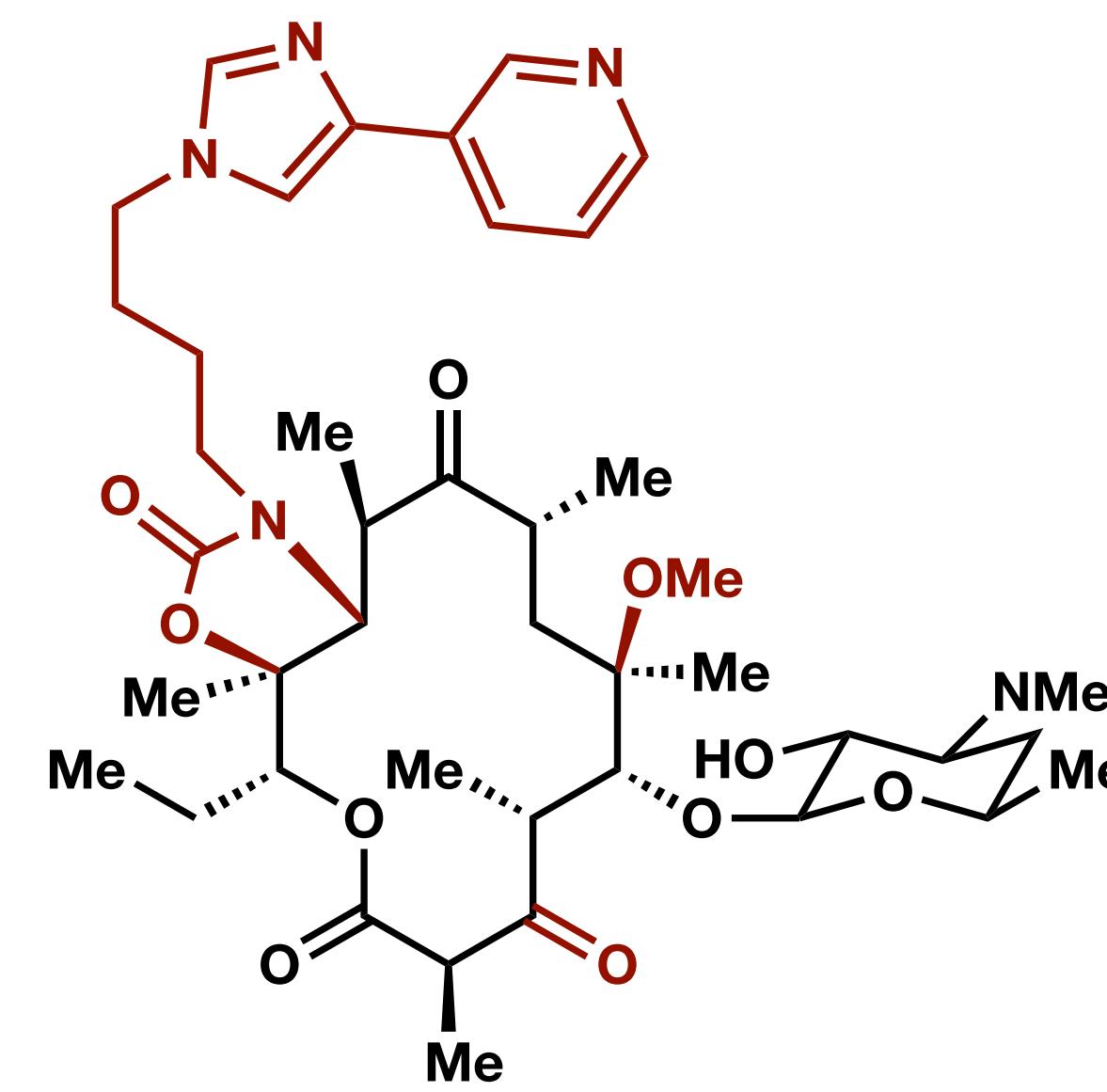


# Clarithromycin (**6** steps from erythromycin)

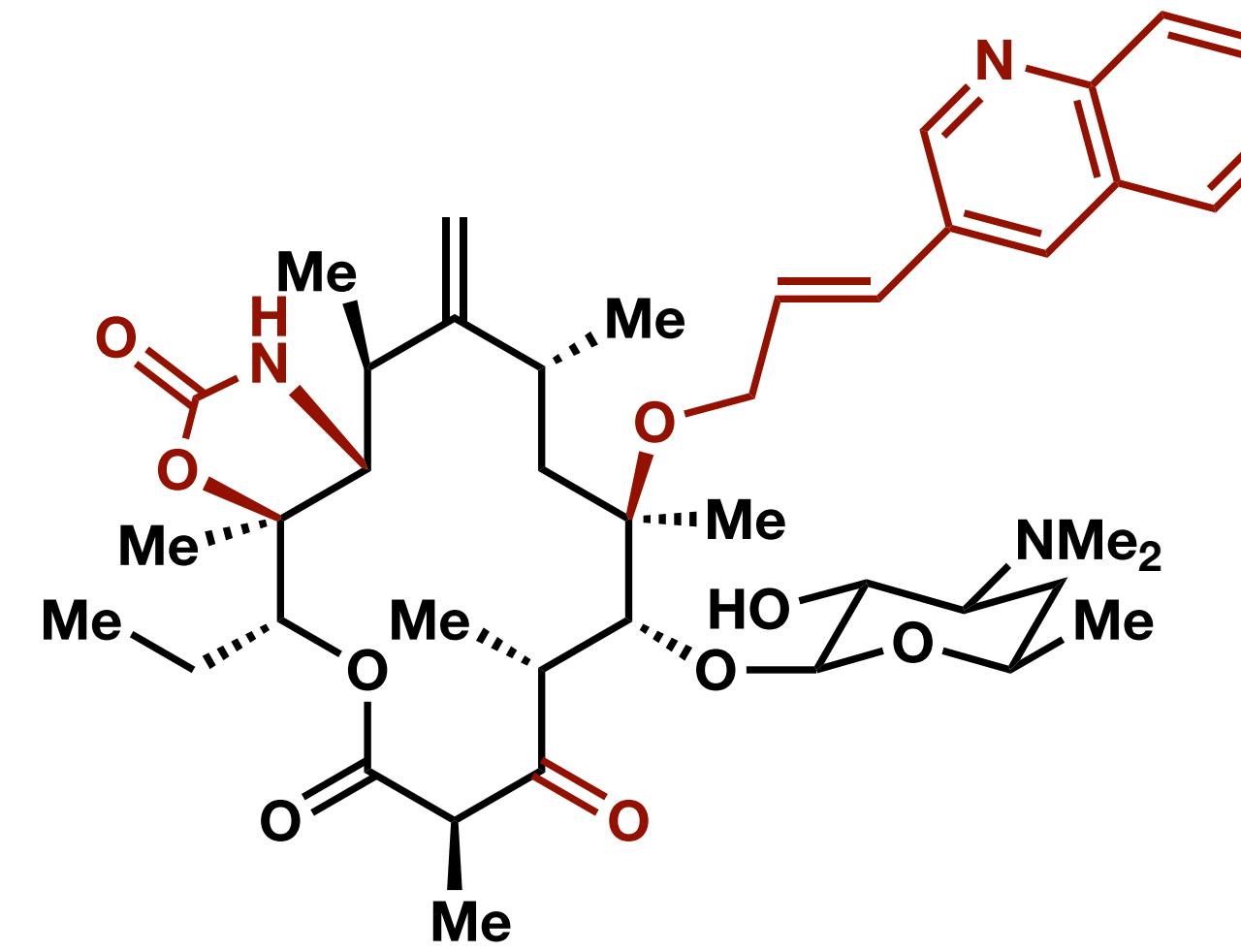
## 1991



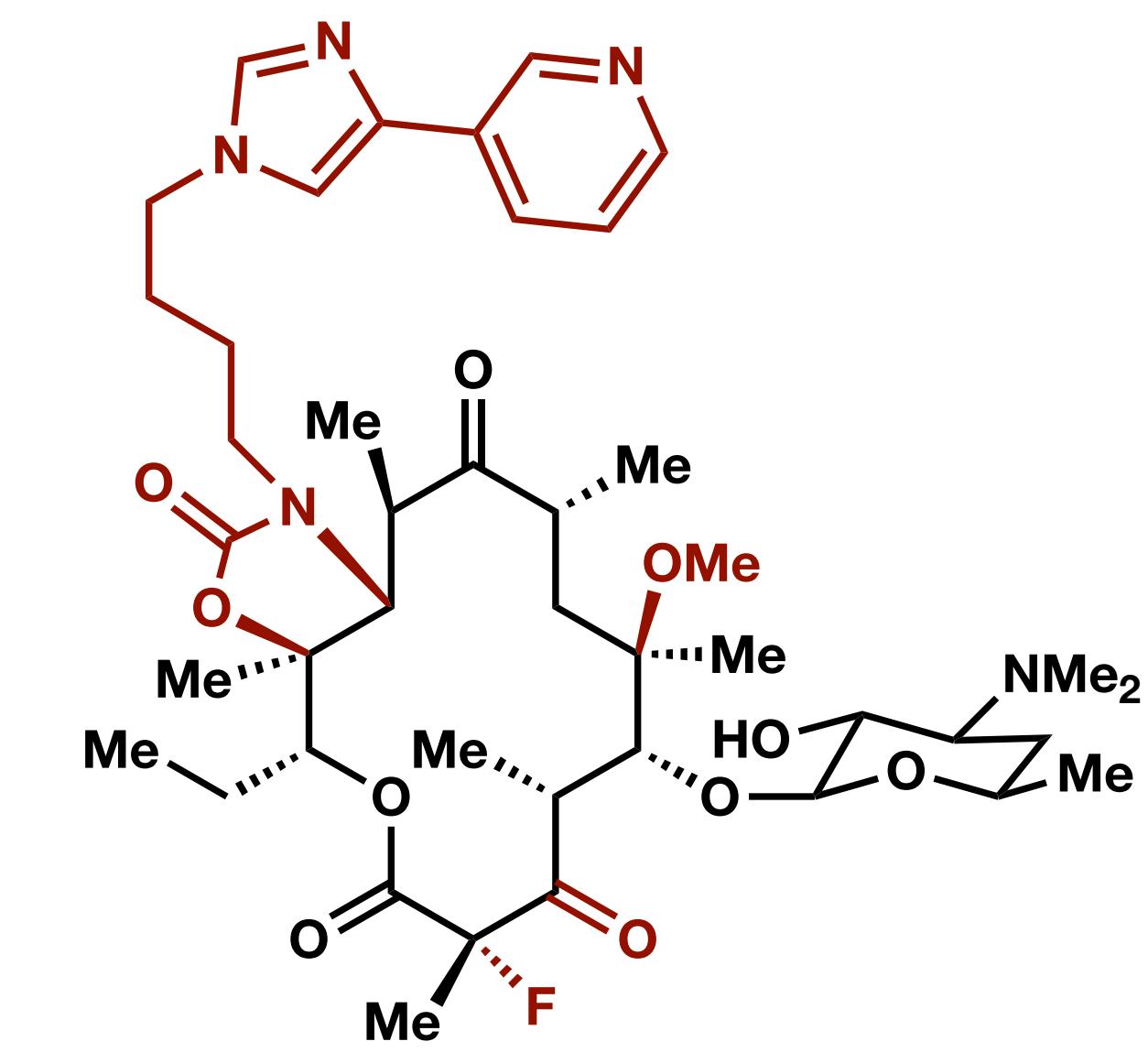
# Azithromycin (**4** steps from erythromycin) 1991



# Telithromycin (**12** steps from erythromycin) 2004



# Cethromycin (**9** steps from erythromycin) Former clinical candidate

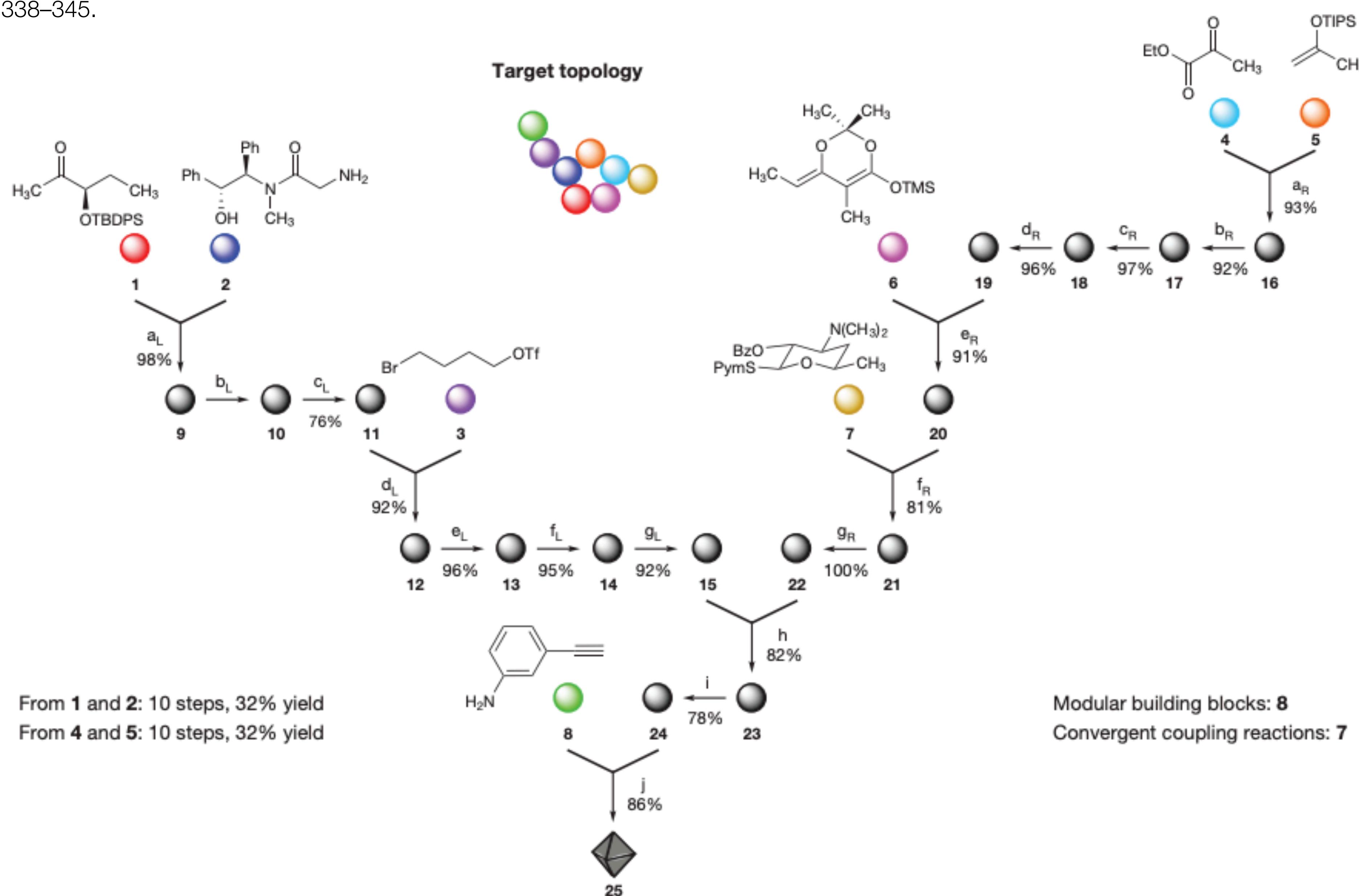


# Solithromycin (**16** steps from erythromycin) Former clinical candidate

# Macrolides - Convergent Strategy

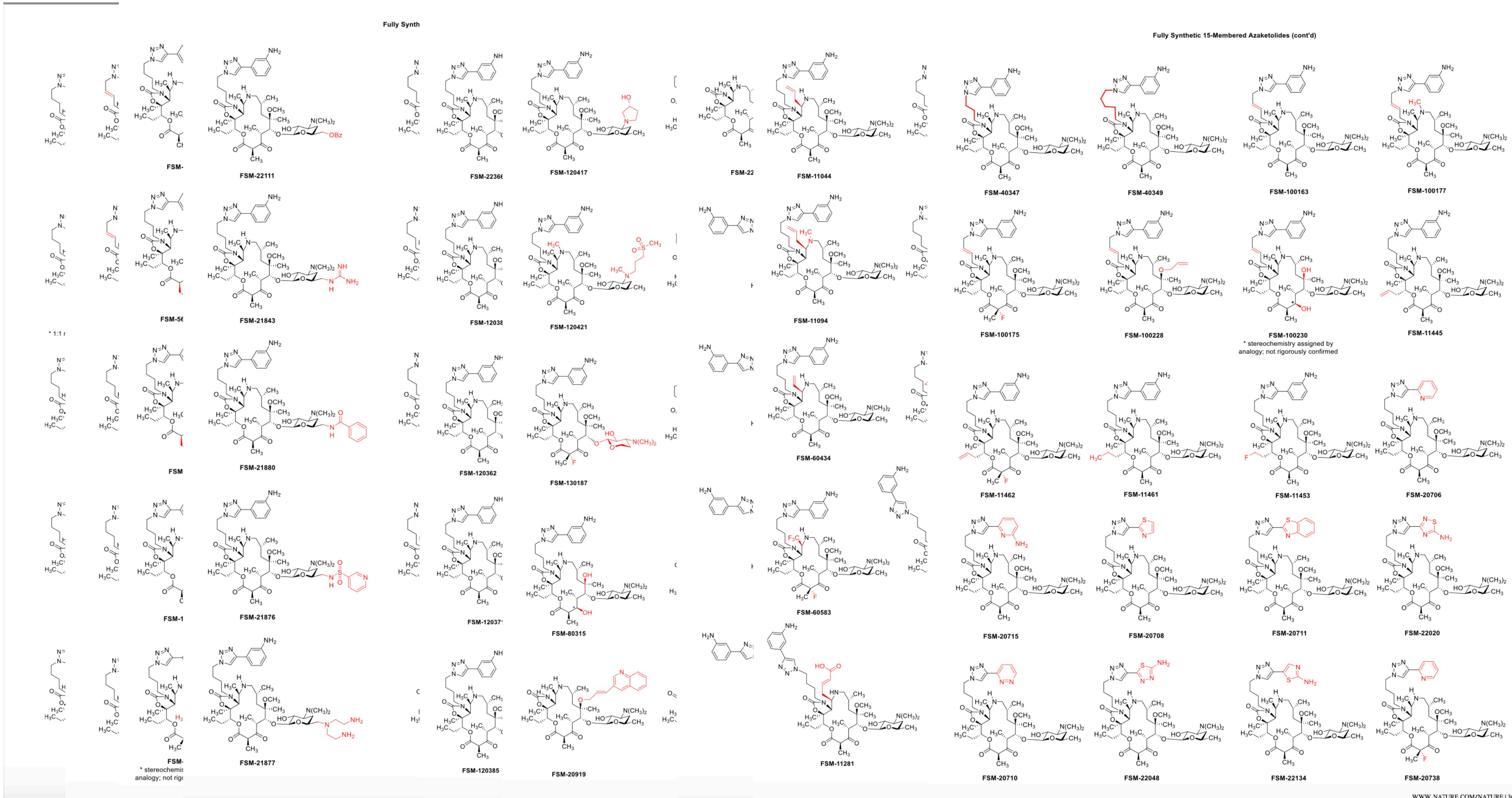
Nature. 2016; 533(7603): 338–345.

a



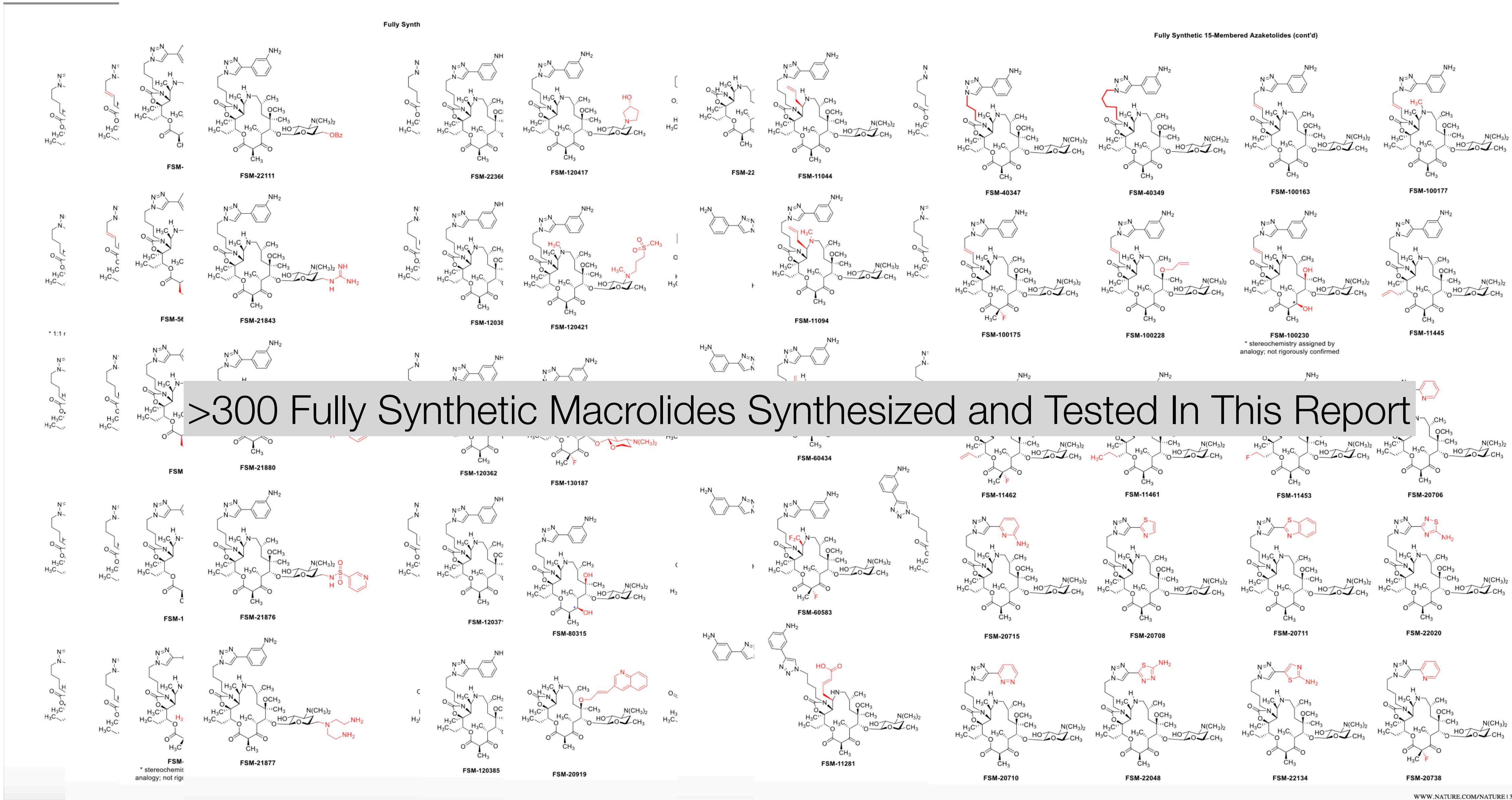
# Macrolides - Convergent Strategy

Nature. 2016; 533(7603): 338–345.



# Macrolides - Convergent Strategy

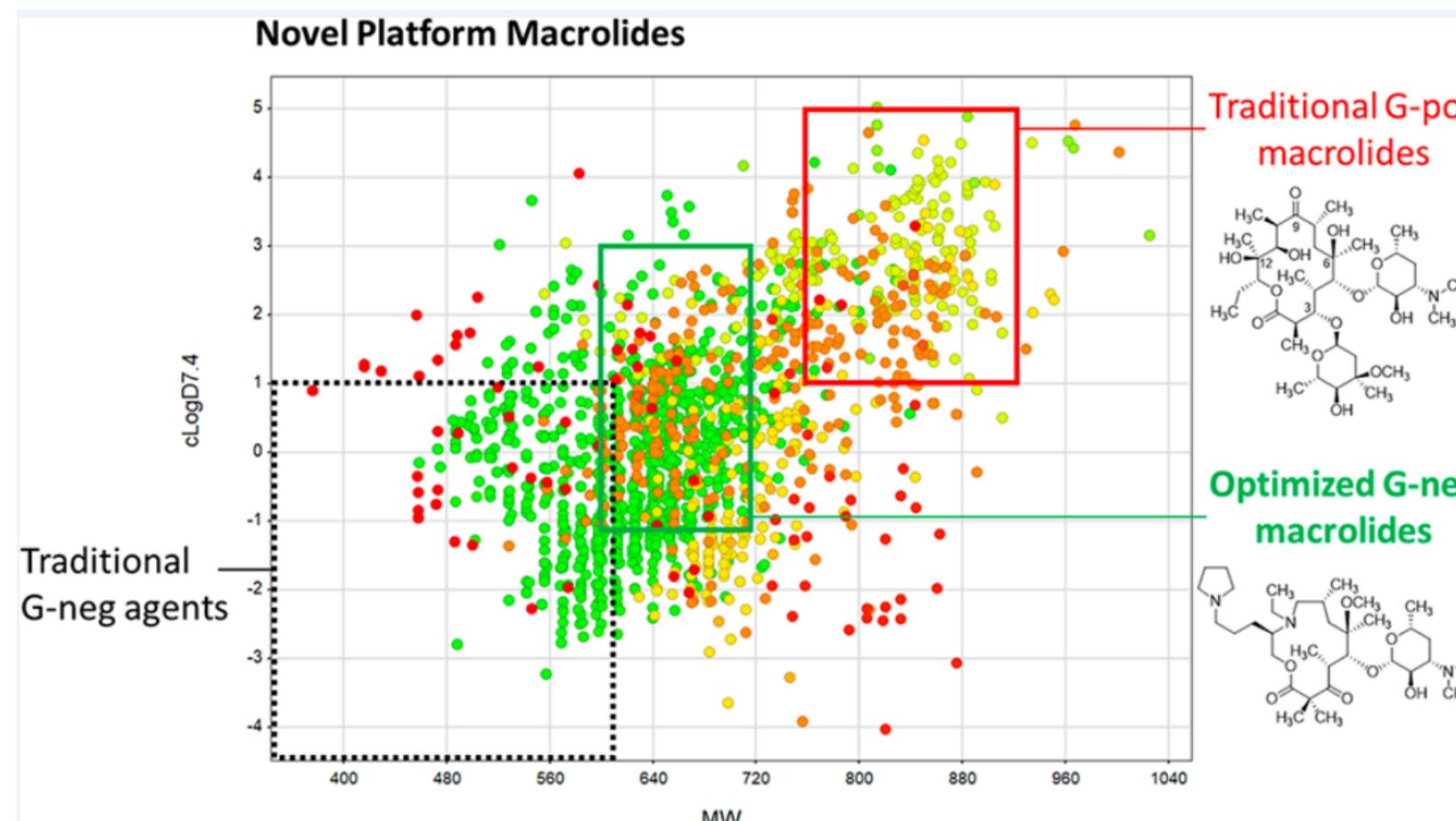
Nature. 2016; 533(7603): 338–345.



# Macrolides - Convergent Strategy

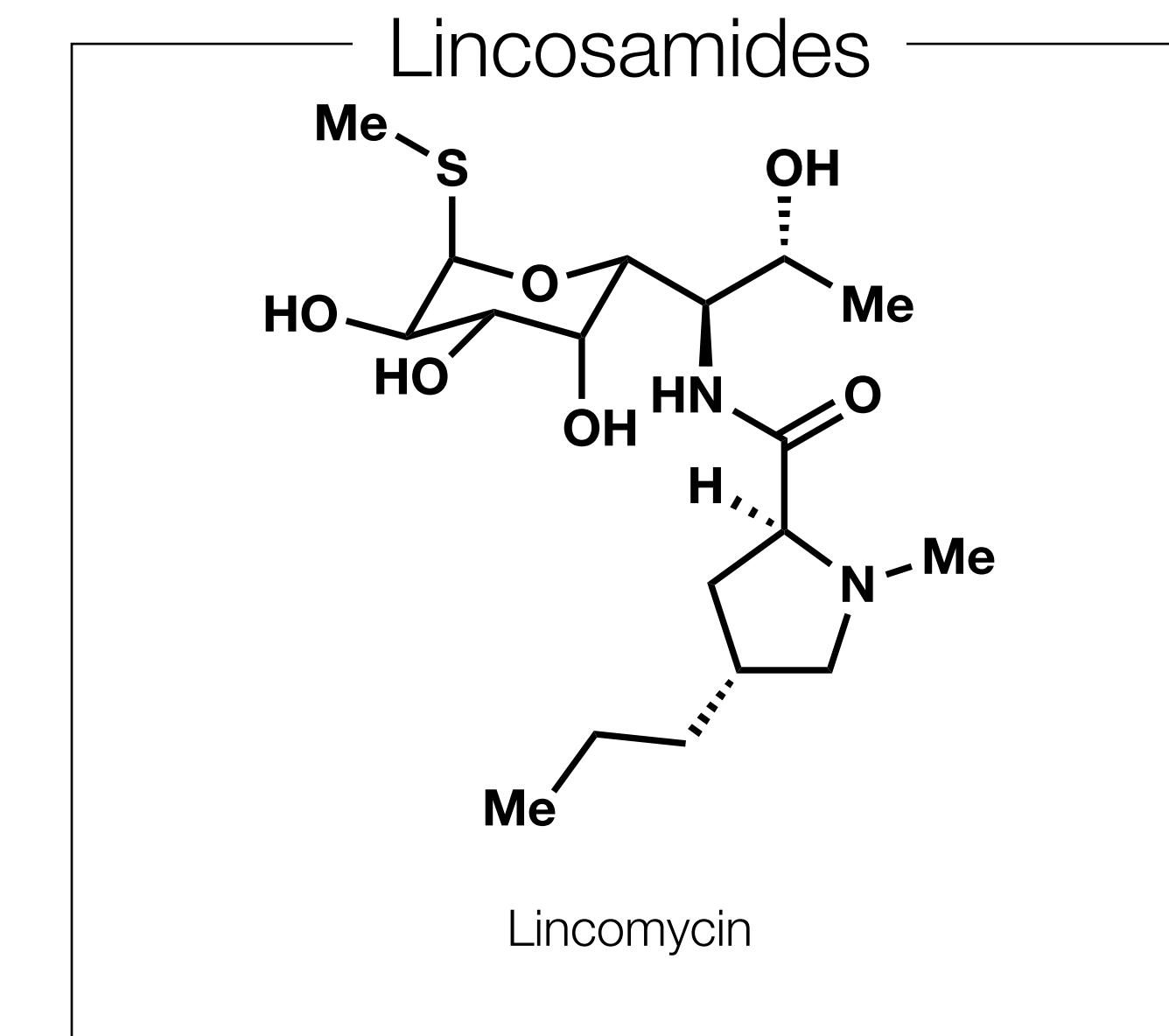
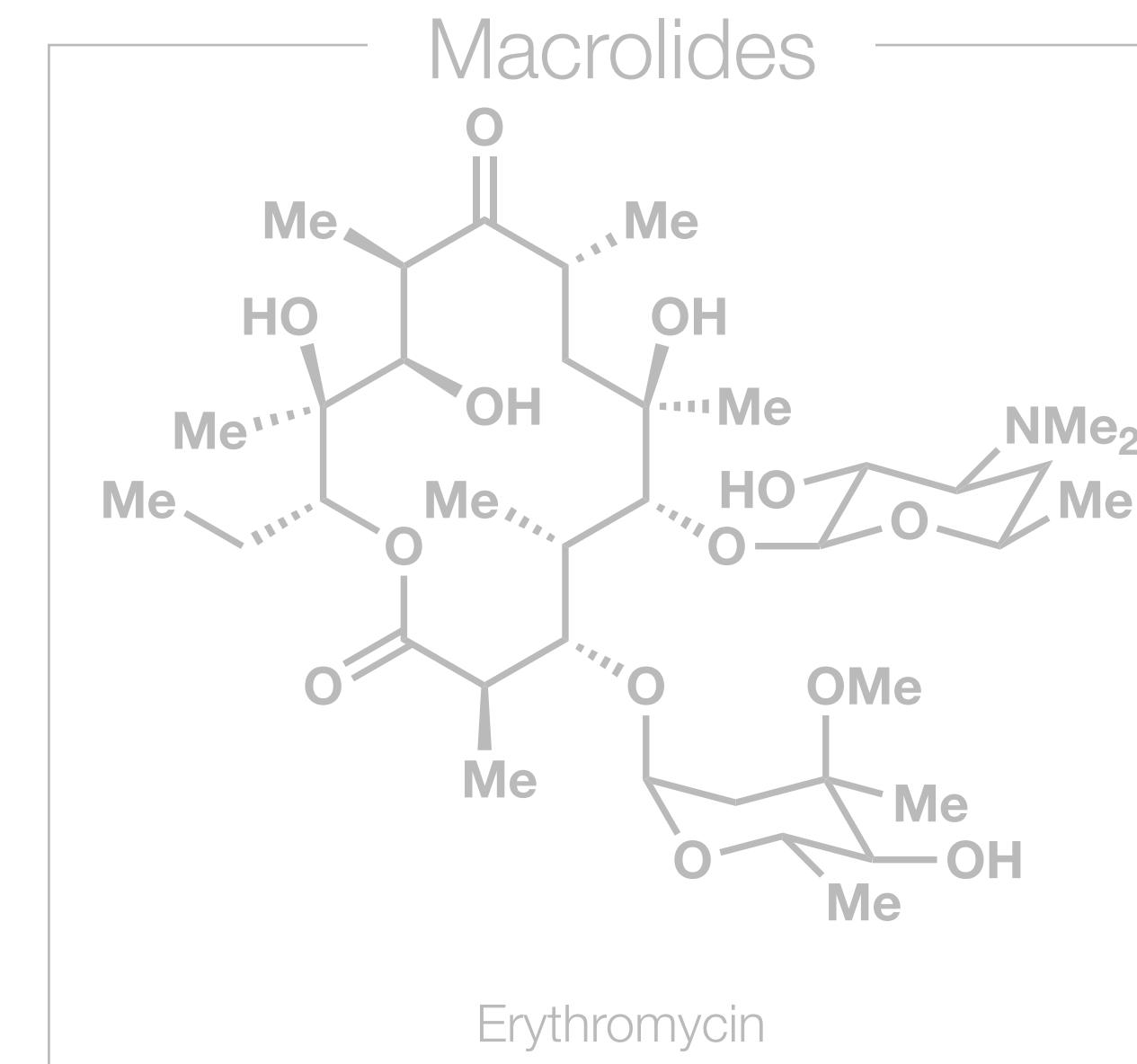
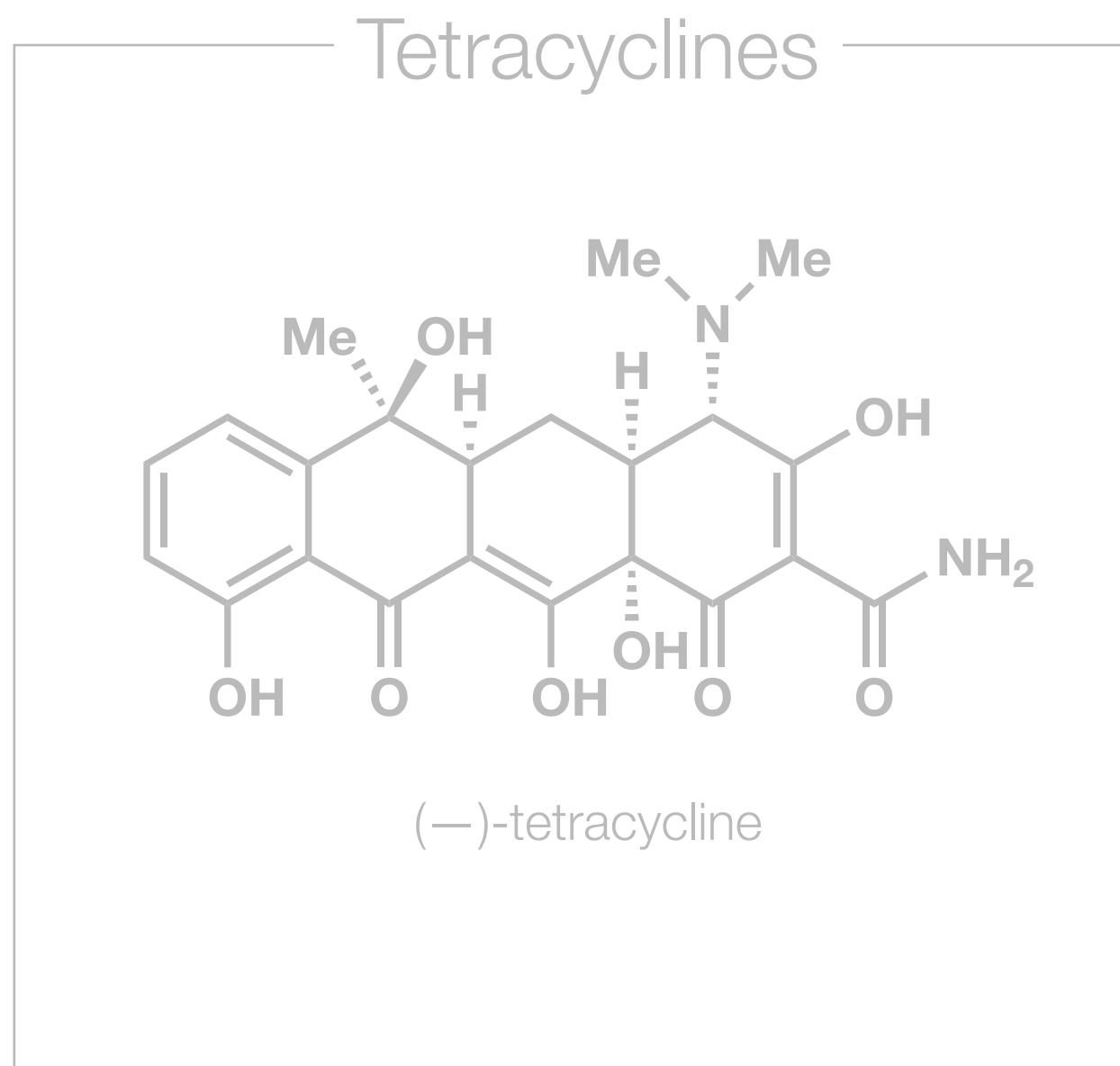
As of 2021, more than 2500 macrolides have been prepared using Myers' strategy

This huge library has enabled correlation of physicochemical properties to enhanced activity, especially towards the development of antibiotics for Gram-negative bacteria



# Combatting Antibiotic Resistance

*Perhaps Myers' Most Impactful Work*



Overarching Research Theme

Convergent Routes with High Modularity

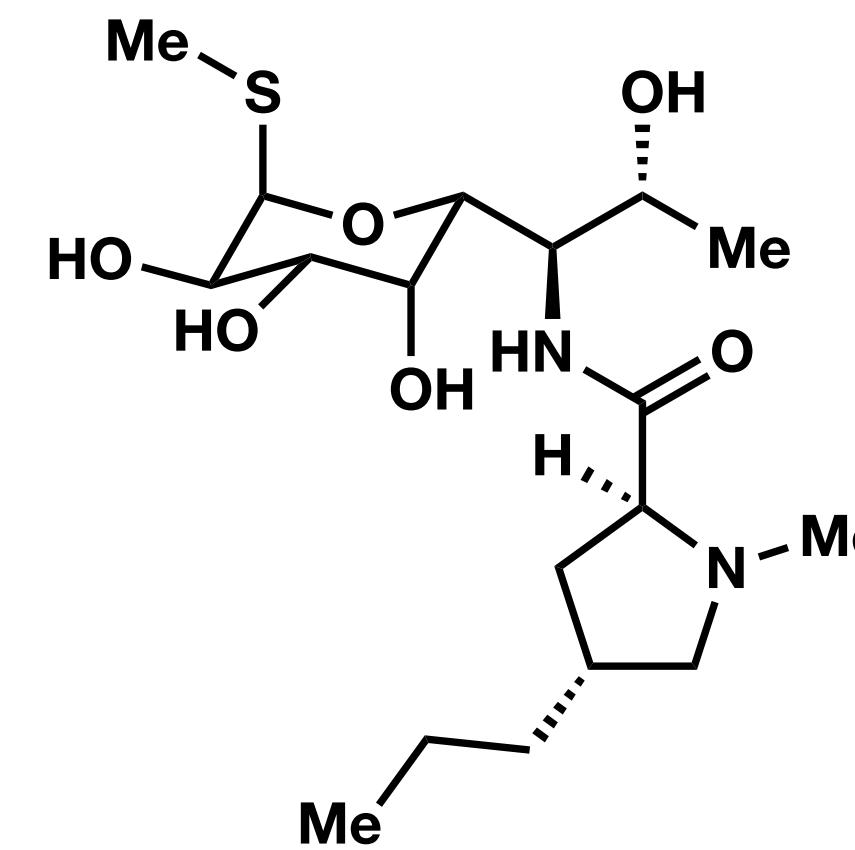


SAR Exploration

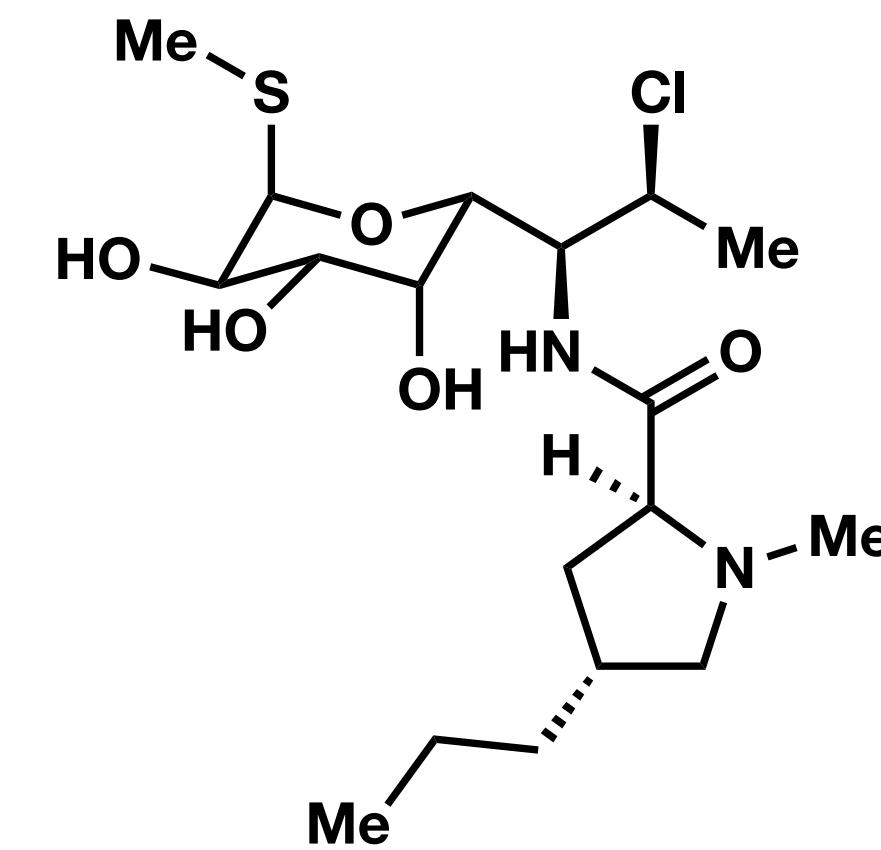


Identification of Clinically Relevant Targets

# Lincosamides

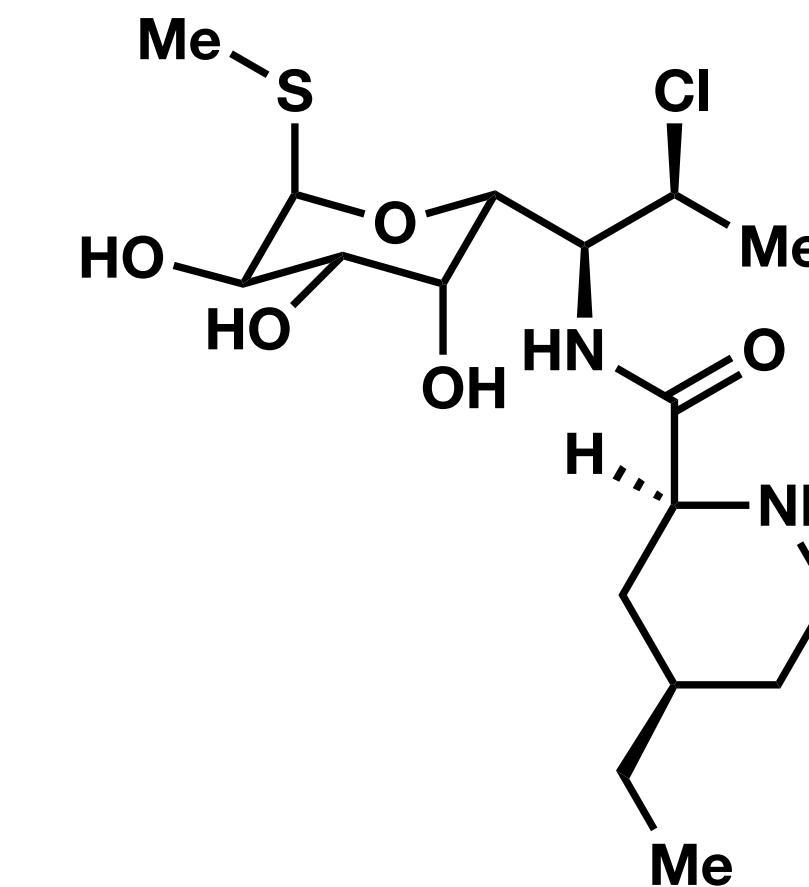


Lincomycin



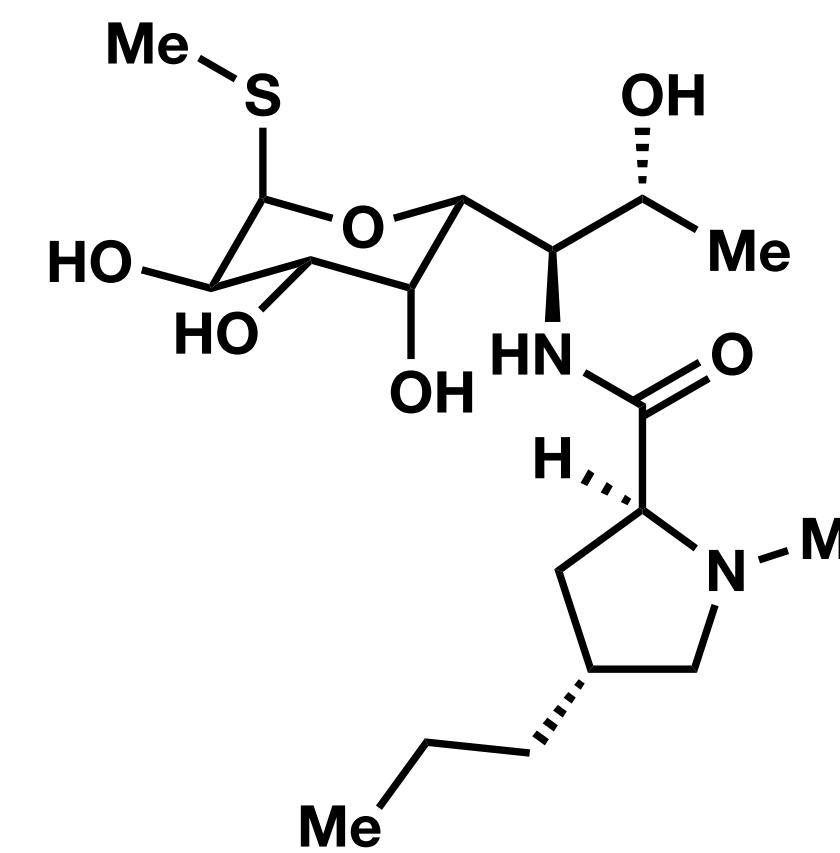
Clindamycin

FDA Approved in 1970

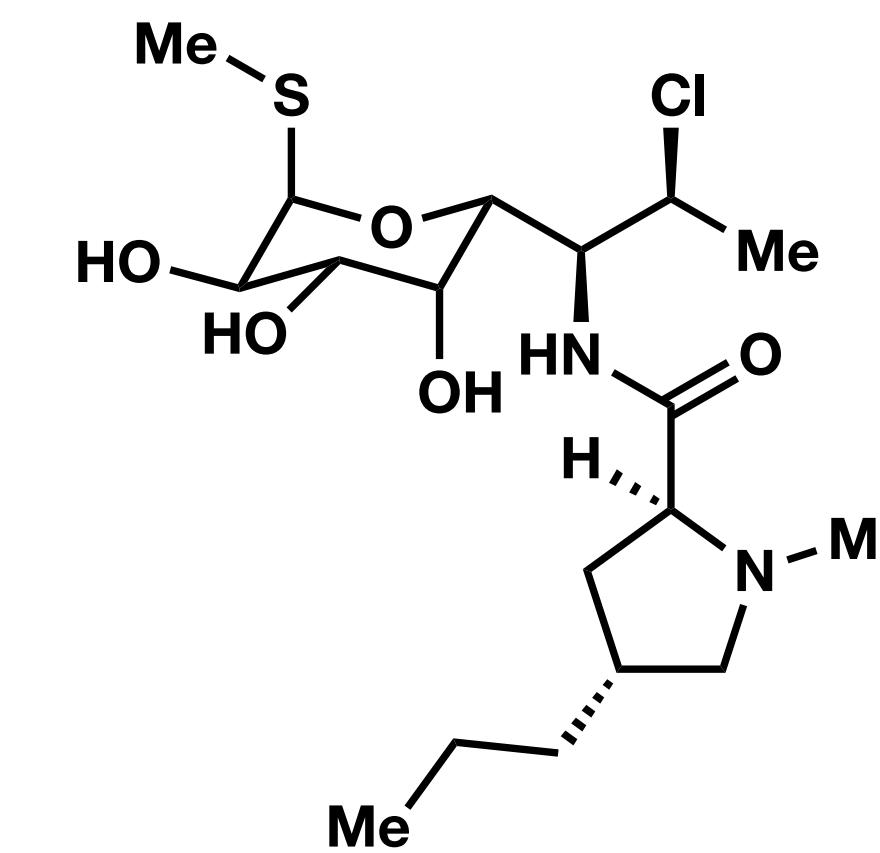


Pirlimycin (Pirsue)

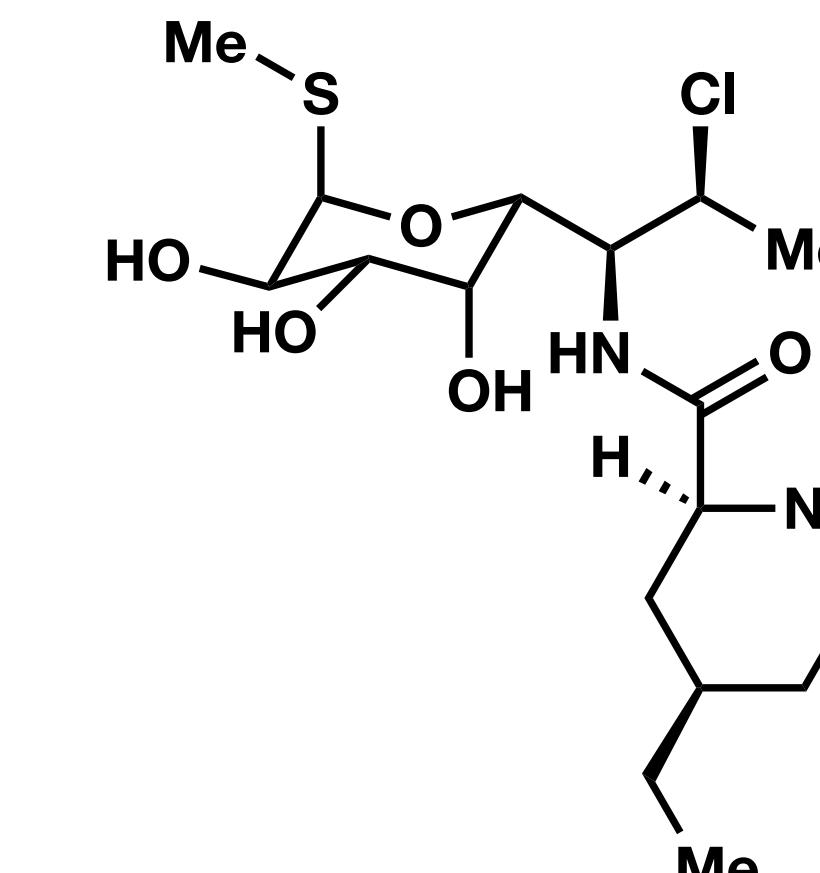
# Lincosamides



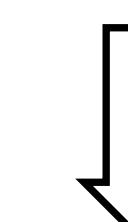
Lincomycin



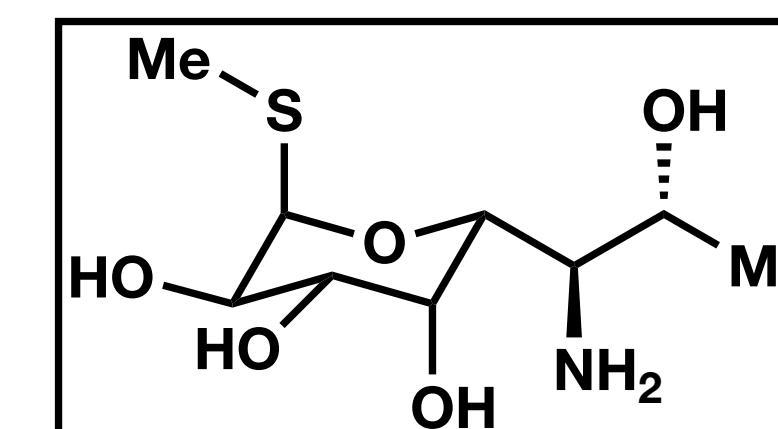
Clindamycin



Pirlimycin (Pirsue)

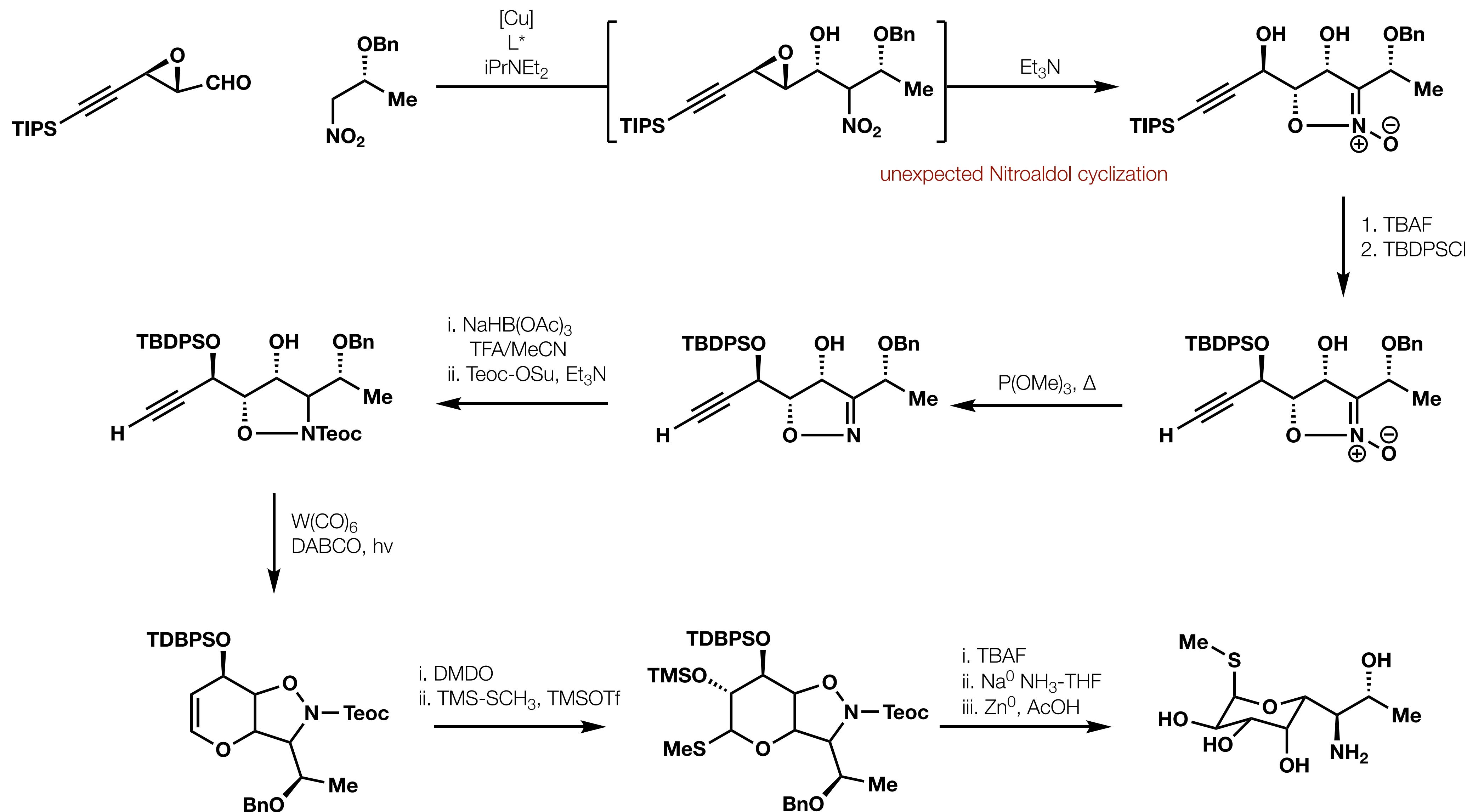


de novo synthesis needed!



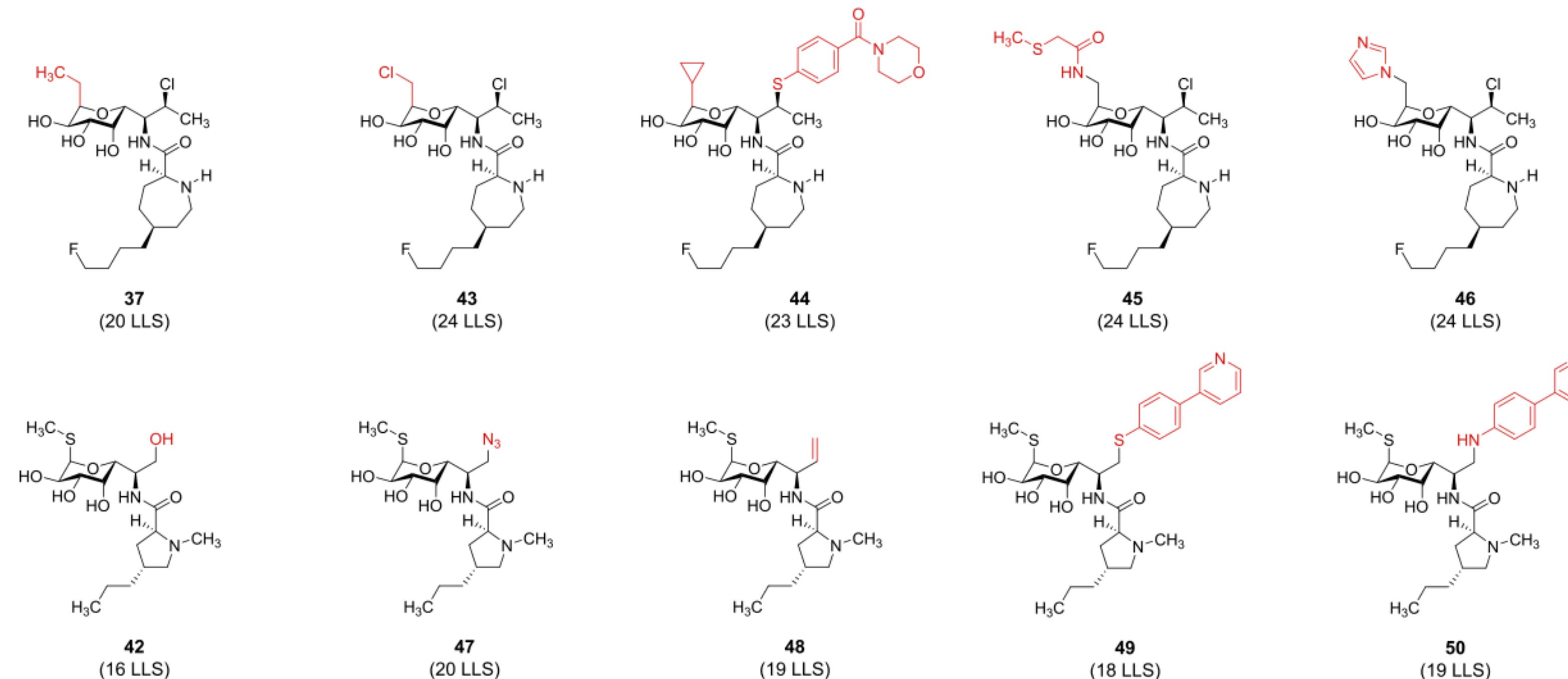
Methylthiolincosamine (MTL)

# Lincosamides



# Lincosamides

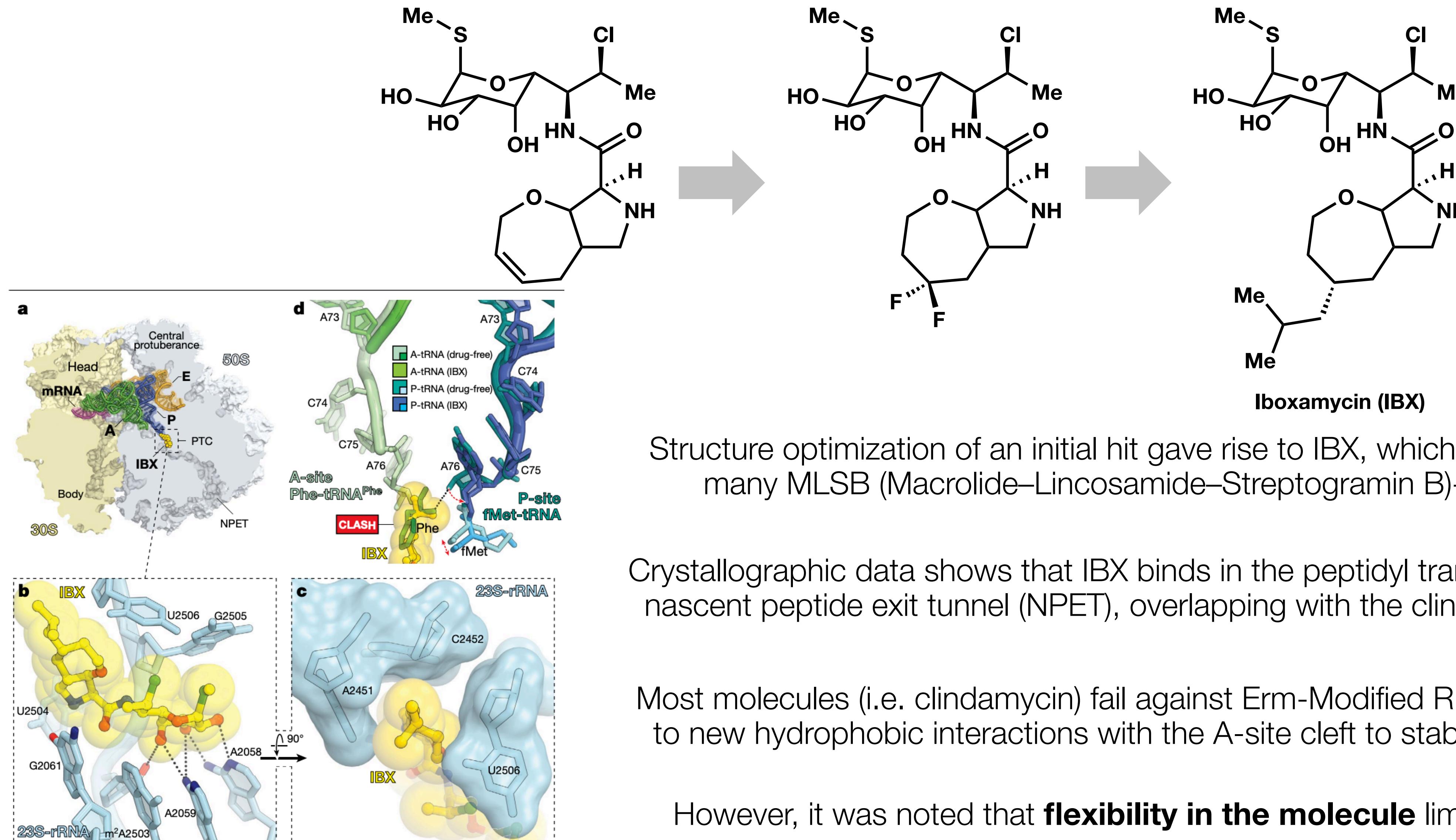
**Table 1. Structures and Minimum Inhibitory Concentrations ( $\mu\text{g}/\text{mL}$ ) of Selected Lincosamides Prepared by the Route Described**



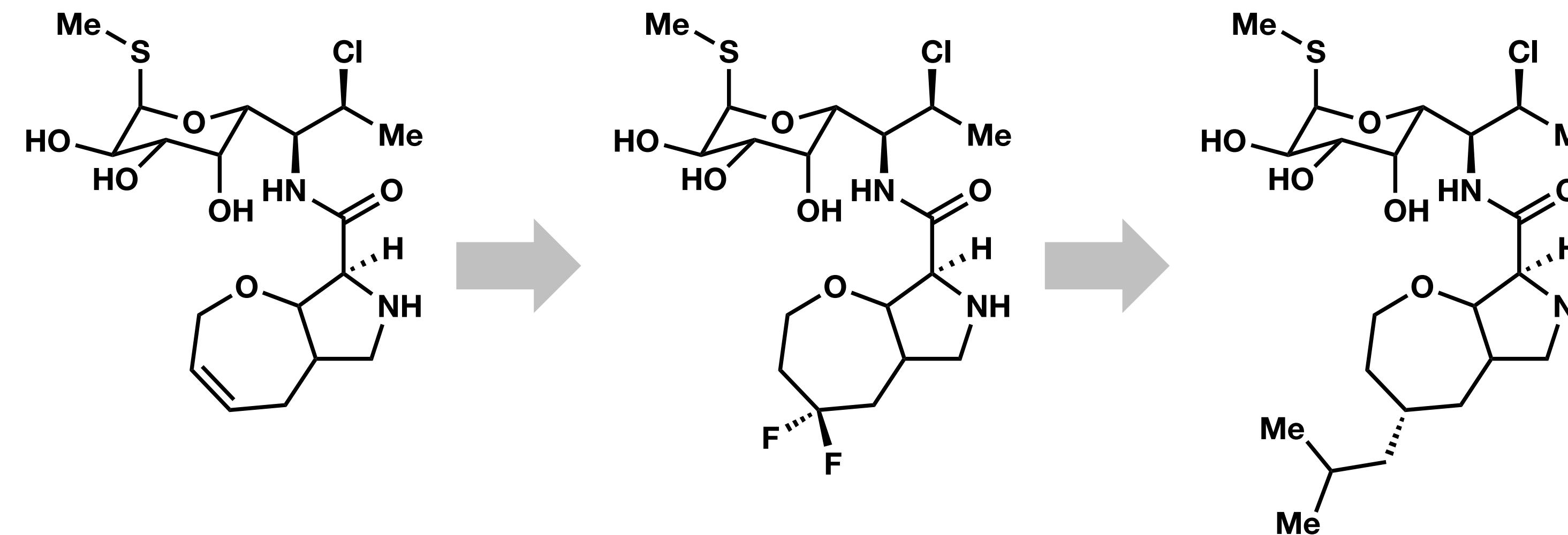
Species	Description	Linco	Clinda	5	37	43	44	45	46	42	47	48	49	50
Gram +	<i>S. aureus</i> ATCC 29213	1	0.25	$\leq 0.06$	$\leq 0.06$	16	8	>64	>64	32	8	8	0.5	32
	<i>S. aureus</i> BAA 977; <i>i-ermA</i>	1	0.25	$\leq 0.06$	$\leq 0.06$	4	8	>64	>64	NT	NT	8	NT	NT
	<i>S. pneumoniae</i> ATCC 49619	0.5	0.12	$\leq 0.06$	$\leq 0.06$	1	0.12	2	8	4	4	2	0.25	8
	<i>S. pneumoniae</i> MMX 3028; <i>c-ermB</i>	>64	>64	8	64	>64	16	>64	>64	NT	>64	NT	>64	>64
	<i>S. pneumoniae</i> MMX 3031; <i>c-mefA</i>	0.25	0.06	$\leq 0.06$	$\leq 0.06$	1	0.12	4	8	NT	0.12	NT	0.25	32
	<i>S. pyogenes</i> ATCC 19615	$\leq 0.06$	0.06	$\leq 0.06$	$\leq 0.06$	4	$\leq 0.06$	2	4	8	2	2	$\leq 0.06$	8
	<i>S. pyogenes</i> MMX 946; <i>MLS<sub>B</sub></i>	>64	>64	4	64	>64	4	>64	>64	NT	>64	NT	>64	>64
	<i>E. faecalis</i> ATCC 29212	32	16	$\leq 0.06$	>64	>64	>64	>64	>64	64	>64	NT	>64	>64
Gram -	<i>K. pneumoniae</i> ATCC 10031	NT	8	0.5	1	NT	NT	NT	NT	NT	NT	>64	NT	NT
	<i>E. coli</i> ATCC 25922	>64	>64	4	32	>64	>64	>64	>64	>64	>64	>64	>64	>64
	<i>P. aeruginosa</i> ATCC 27853	>64	>64	>64	>64	>64	>64	>64	>64	NT	NT	NT	NT	NT
	<i>H. influenzae</i> ATCC 49247	32	16	0.25	1	>64	>64	>64	>64	NT	NT	NT	NT	NT

MIC Color Scale ( $\mu\text{g}/\text{mL}$ )     $\leq 0.06$     0.12    0.25    0.5    1    2    4    8    16    32    64    >64

# Lincosamides - Iboxamycin



# Lincosamides - Iboxamycin



# Iboxamycin (IB)

<b>a</b>	<b>Species</b>	<b>Strain description</b>	<b>IBX</b>	<b>CLI</b>							
Gram-positive	<i>S. aureus</i>	ATCC 29213	0.06	0.125							
	<i>S. aureus</i>	Clinical; MDR, <i>c-ermA</i>	1	>256							
	<i>S. aureus</i>	Clinical; <i>msrA</i>	0.06	0.125							
	<i>S. aureus</i>	Clinical; <i>cfr</i>	2	>128							
	<i>S. epidermidis</i>	Clinical; <i>cfr</i>	8	>128							
	<i>S. haemolyticus</i>	Clinical; LNZ-R, MEC-R	0.06	2							
	<i>S. pneumoniae</i>	Clinical; <i>c-ermB</i>	0.25	256							
	<i>S. pneumoniae</i>	ATCC 700673; MDR	0.5	>64							
	<i>S. pyogenes</i>	ATCC 19615	0.03	0.06							
	<i>E. faecalis</i>	ATCC 29212; <i>IsaA</i>	0.06	16							
Gram-negative	<i>E. faecalis</i>	Clinical; <i>c-ermB</i>	1	>256							
	<i>E. faecium</i>	Clinical; VRE, <i>vanA</i>	1	>64							
	<i>C. difficile</i>	ATCC 700057	0.25	8							
	<i>B. fragilis</i>	ATCC 25285	0.5	1							
	<i>E. coli</i>	ATCC 25922	8	>128							
	<i>K. pneumoniae</i>	ATCC 10031	0.25	8							
	<i>K. pneumoniae</i>	Clinical; FQ-R	8	>128							
	<i>K. oxytoca</i>	Clinical	8	>128							
	<i>A. baumannii</i>	ATCC 19606	4	>128							
	<i>P. aeruginosa</i>	ATCC 27853	128	>128							
	<i>H. influenzae</i>	ATCC 9007	0.5	8							
	<i>N. gonorrhoeae</i>	Clinical	0.125	2							

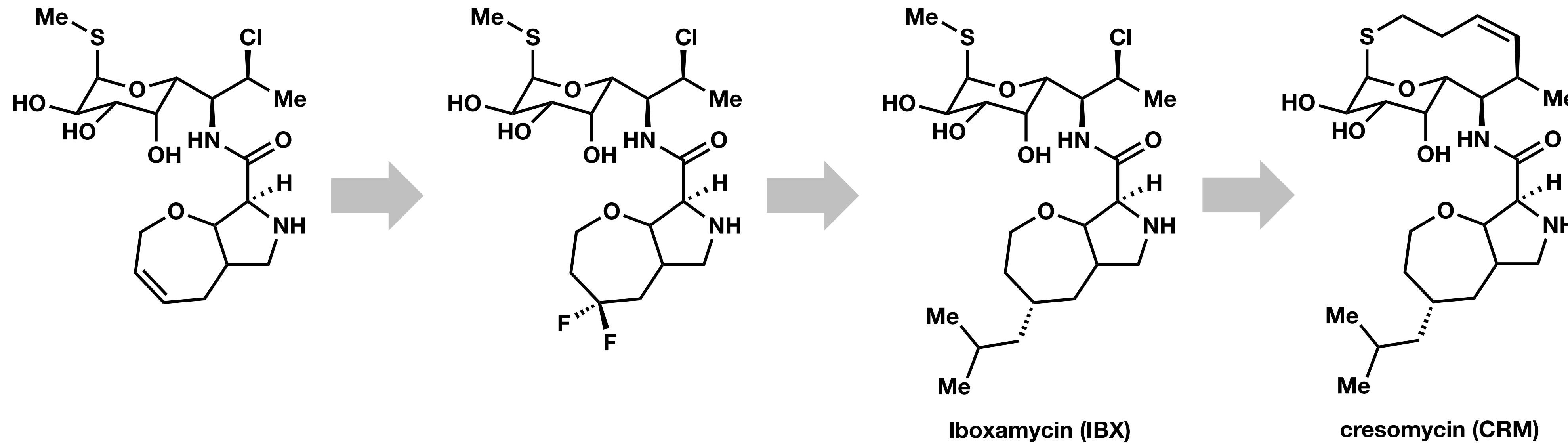
  

<b>b</b>	<b>Species</b>	<b>Strain description</b>	<b>IBX</b>	<b>CLI</b>	<b>CTR</b>	<b>LEVO</b>	<b>AZM</b>	<b>DOXY</b>	<b>LNZ</b>	<b>VAN</b>
<i>S. aureus</i>	ATCC BAA-1707; MRSA	0.06	0.125	64	>128	1	1	0.25	2	1
<i>S. aureus</i>	Clinical; MRSA	0.06	0.25	>128	1	1	64	2	2	2
<i>S. aureus</i>	ATCC 700699; <i>c-ermA</i>	2	>128	>128	32	>128	16	2	2	8
<i>S. aureus</i>	Clinical; <i>cfr</i>	2	>128	128	8	>128	0.25	16	>64	
<i>S. pneumoniae</i>	Clinical; MLS <sub>B</sub>	0.25	>64	1	0.25	>64	2	4	2	2
<i>S. pyogenes</i>	MMX 946; <i>c-ermB</i>	0.25	>256	≤0.03	0.5	>64	0.125	1	0.5	
<i>E. faecalis</i>	Clinical; VRE	1	>256	128	>128	16	2	>128		
<i>E. faecalis</i>	Clinical; VRE	2	>256	>256	64	>128	1	16	>128	
<i>E. faecium</i>	Clinical; VRE, LNZ-R	≤0.06	>64	>256	128	8	16	64	>256	
<i>E. faecium</i>	Clinical; VRE	1	>256	>256	128	>128	0.25	4	>128	

<b>c</b>	<b>Species</b>	<b>Strain description</b>	<b>IBX</b>	<b>CLI</b>	<b>CTR</b>	<b>LEVO</b>	<b>AZM</b>	<b>DOXY</b>	<b>LNZ</b>	<b>GEN</b>
<i>E. coli</i>	Clinical	8	>128	0.125	32	64	0.5	64	2	
<i>E. coli</i>	Clinical; <i>armA</i>	8	>256	≤0.06	≤0.06	4	2	128	>256	
<i>E. coli</i>	Clinical; CRE, NDM-1	8	64	>128	64	8	32	128	4	
<i>E. coli</i>	Clinical; ESBL	8	128	64	16	4	8	64	2	
<i>E. coli</i>	Clinical; MDR, <i>arm</i>	8	>128	>128	32	4	32	128	64	
<i>K. pneumoniae</i>	Clinical; CRE	8	>128	>128	32	32	16	>128	8	
<i>K. pneumoniae</i>	Clinical; 3GC-R	16	>128	64	1	8	2	32	0.5	
<i>K. pneumoniae</i>	Clinical; ESBL	16	>128	128	0.5	4	16	64	0.5	
<i>A. baumannii</i>	Clinical; CRAB	16	128	64	32	4	32	64	4	
<i>A. baumannii</i>	Clinical; CRAB, MDR	16	128	64	64	4	2	64	>128	

# Lincosamides - it gets better!



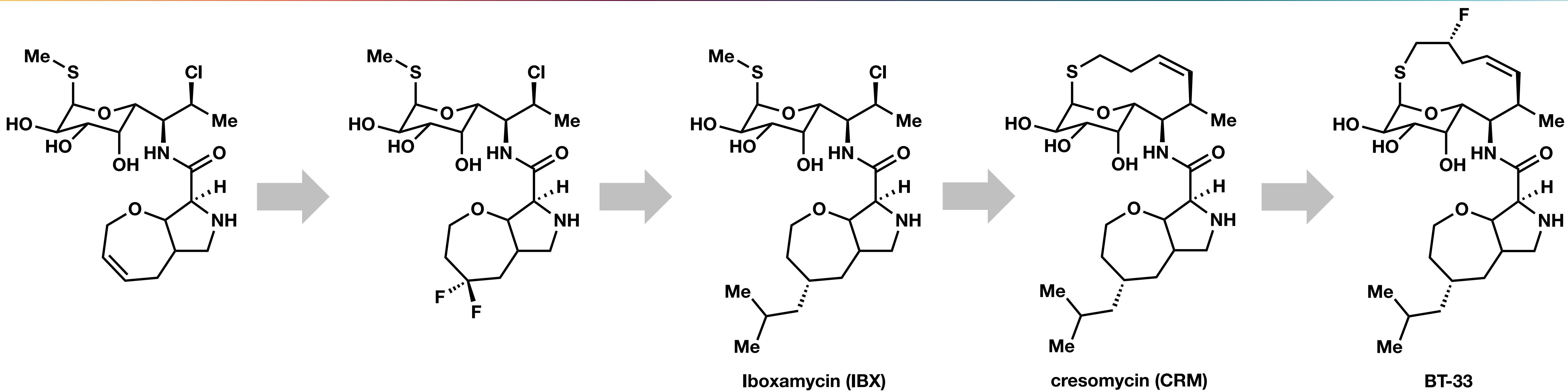
To rigidify IBX, a bridged, macrobicyclic structure was introduced

DFT was used to predict favorable conformations, and synthesis of CRM commenced after seeing that CRM had significantly less low energy conformers than IBX

Successful synthesis allowed for *in vivo* and *in vitro* studies

CRM showed stronger ribosomal engagement, enhanced potency, and retained activity against a wider range of resistant strains

# Lincosamides - it gets EVEN better!



Additional fluoride provides additional van der Waals contact with nucleobase G2505 in the ribosome, enhancing binding affinity

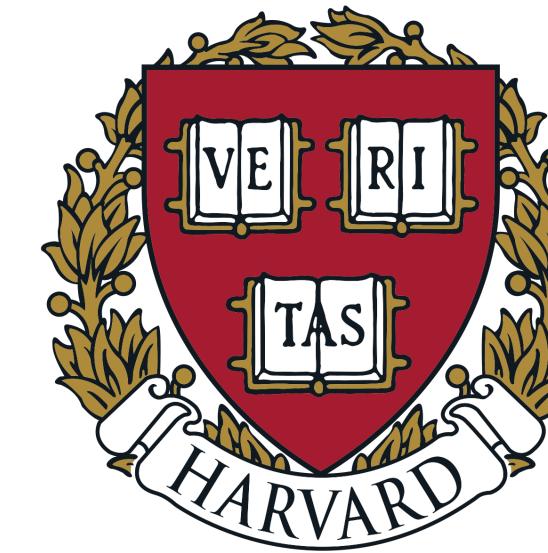
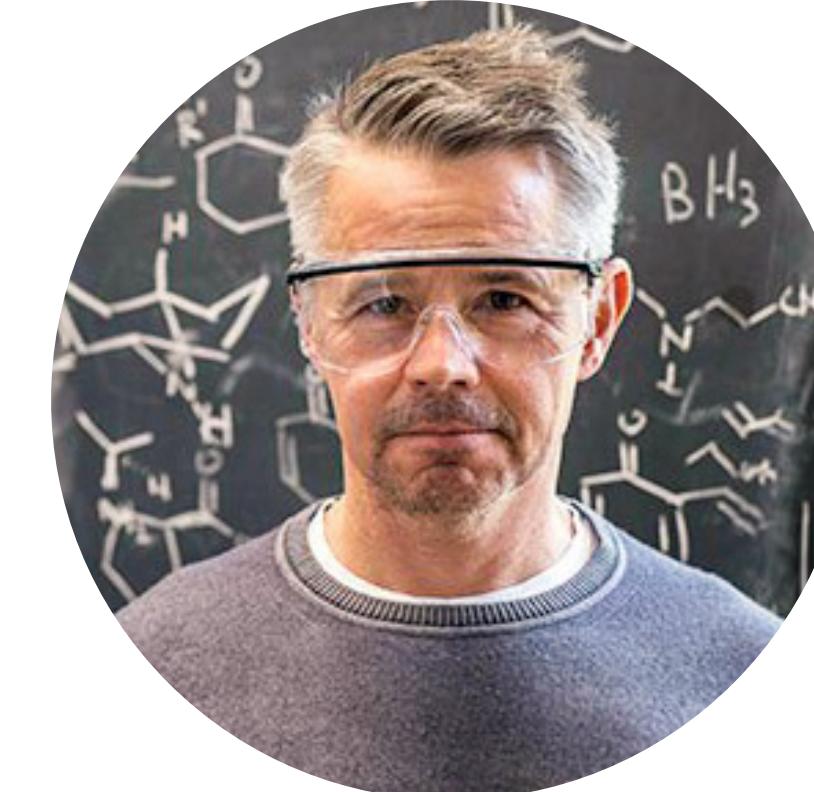
BT-33 exhibited even lower MICs than CRM against a broader range of multi-drug resistant Gram-positive and Gram-negative bacteria

BT-33 also proved to be more metabolically stable, with a half-life in vivo (6.80 hours) compared to CRM (4.66 hours) and iboxamycin (2.05 hours)

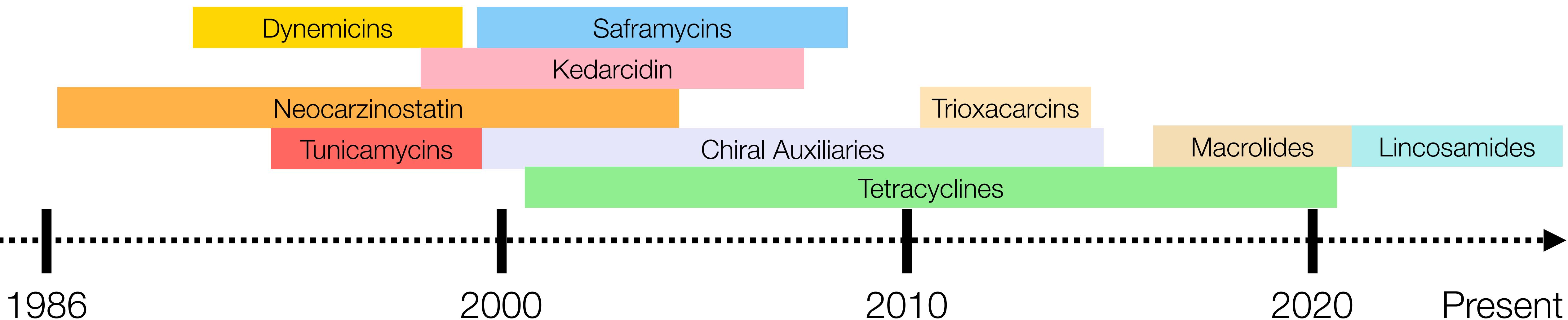
# Andrew G. Myers



Assistant, Associate, and  
then Full Professor (1994)



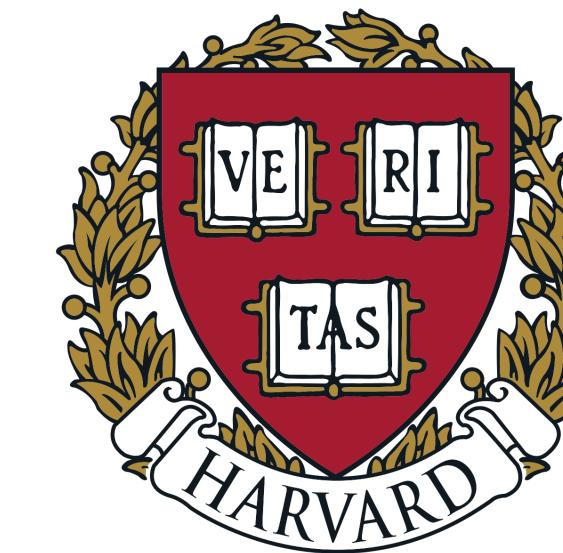
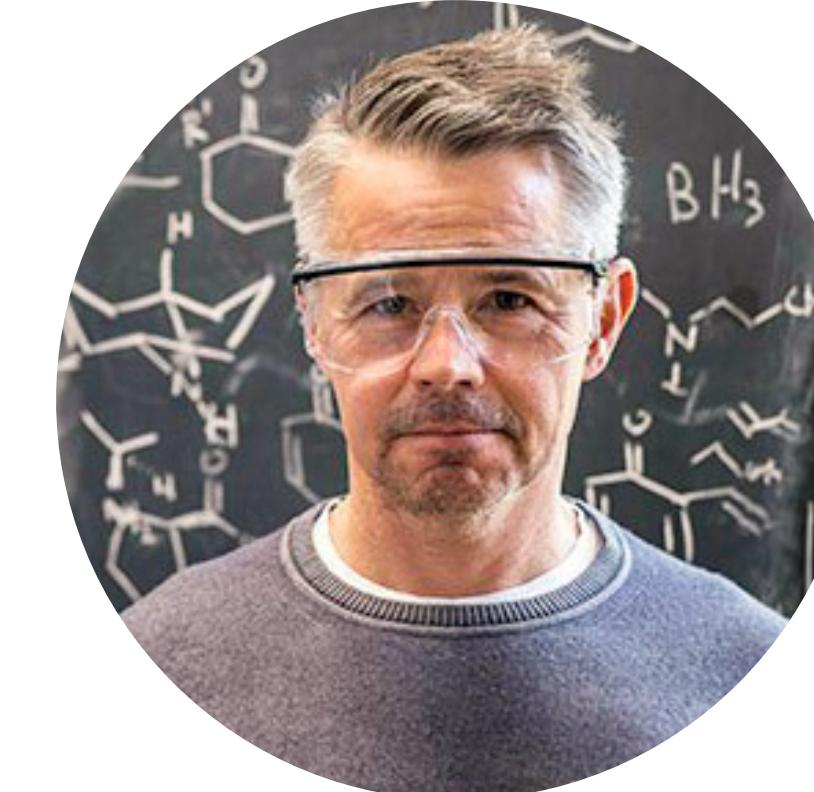
Department Chair 2007-2010



# Andrew G. Myers



# Assistant, Associate, and then Full Professor (1994)



## Department Chair 2007-2010

